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Predation of portunid crabs *Callinectes sapidus* and *Callinectes ornatus* (Crustacea, Decapoda) on the medusa *Lychnorhiza lucerna* (Cnidaria, Scyphozoa)

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Jellyfishes (pelagic medusozoans and ctenophores) have a limited number of specialized predators, such as the leatherback turtle (Dermochelys coriacea), the ocean sunfish (Mola mola) and some comb jellyfishes (Beroe spp.); however, an increasing number of pelagic and benthic species have been reported feeding on jellyfishes (reviewed by Ates, 2017; Hays et al., 2018). Nonspecialized jellyfish consumers include fishes, birds, and marine mammals, as well as many invertebrates, such as crabs, amphipods, other jellyfish, sea anemones, etc. These recent studies have indicated that predation and scavenging on jellyfish are widespread, contradicting a previous paradigm in which jellyfish were considered the "dead ends" of marine trophic webs (Hays et al., 2018). Under this new view, the energy channelized for the production of large jellyfish blooms can be naturally driven toward multiple sets of consumers (Sweetman et al., 2014; Hays et al., 2018). This topic is, however, widely unknown and many questions remain unsolved, such as what is the importance of jellyfish for the energy intake of these consumers? Which populations of jellyfish predators take advantage of jellyfish blooms?

Many marine animals are found together with large jellyfishes in associations that may result in multiple benefits, such as protection against

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predation, transport in the water column, and food acquisition from jellyfish tissues or captured prey (Purcell and Arai, 2001; Fleming et al., 2014). When the jellyfish are not affected by their "hitchhikers", such association is considered phoresis (e.g. Martinelli et al., 2008). In some cases, these symbionts also provide benefits to their hosts, by cleaning jellyfish ectoparasites (Towanda and Thuesen, 2006). In contrast, some studies have suggested that jellyfishes are consumed by their symbionts (D'Ambra et al., 2015) or that symbionts ingest food particles captured by the jellyfish (Gonçalves et al., 2020). Brachyuran crabs are often found with scyphomedusae in coastal and estuarine areas (Farr, 1978; Moyano et al., 2012; Ates, 2017; Gonçalves, et al. 2020). These symbioses usually involve the megalopae or young crabs, but adults were also reported on top of the jellyfish bells. Information on the possible trophic role of these symbiotic relationships is scattered or unavailable in many cases. Moyano et al. (2012) reviewed 28 studies on associations involving 18 jellyfish and nine crab species; only nine studies provided evidence of predation on the jellyfish hosts, ranging from direct observations of predation in situ (Farr, 1978), laboratory experiments (Esser et al., 2004), analysis of crab gut contents (Gonçalves et al., 2020), to trophic markers such as stable isotopes (Mancinelli et al., 2017). Although these studies may reveal an important trophic link, the potential damage these symbionts may cause and whether jellyfish populations can be affected by these crabs remain unsolved.

Portunid crabs of the genus *Callinectes* are among the most common macrobenthic invertebrates in tropical and subtropical coastal areas (William, 1974; Guerra-Castro et al., 2007). The diets of these opportunist omnivorous crabs are influenced by food availability (Oliveira et al., 2006; Silveira et al., 2020). Common Callinectes prey include benthic organisms, coastal vegetation, and small fish (Branco et al., 2002; Oliveira et al., 2006). Species such as Callinectes sapidus Rathbun, 1896 are considered estuarine dependent, because their adults inhabit estuarine zones, whereas larvae development occurs in coastal and ocean areas (Epifanio, 2007; Yeager et al., 2007). Callinectes ornatus Smith 1869 inhabits sand and mud bottoms in brackish waters (Melo, 1996). Because of their abundance, these crabs play an important role as consumers in coastal food webs, potentially controlling populations of their prey (Hines, 2007).

The rhizostome medusa Lychnorhiza lucerna Haeckel, 1880 is one of the most common jellyfish in the Southwest Atlantic, occurring from Colombia to the northern Argentinean coast (Schiariti et al., 2008). The species is particularly abundant in coastal areas under the influence of estuaries, and can be dominant in the pelagic environment during its seasonal population peaks (Colombo et al., 2003; Schiariti et al., 2008; Nagata et al., 2009). Newly released ephyrae were recently found in a Brazilian estuary (Nogueira Jr. et al., 2019), which reinforces the importance of such ecosystems to the species life cycle. L. lucerna is a key consumer of mesozooplankton and captures thousands of prey in their filtrating oral arms (Nagata et al., 2016; Nagata and Morandini, 2018); however, the species' predatory impacts and role in coastal trophodynamics are widely unknown. Large individuals of *L. lucerna* often reach 30 cm in bell diameter; their oral arms and subgenital cavities offer many microhabitats that can be inhabited by innumerous symbionts, such as cleaner shrimp Periclimenes paivai (Martineli et al., 2008) and the spider crabs Libinia ferreirae (Nogueira Jr. and Haddad, 2005) and Libinia spinosa (Moyano et al., 2012). To the best of our knowledge, this study is the first record of an association between the medusa L. lucerna and portunid crabs of the genus Callinectes. We also analyzed the gut contents of the crabs, providing evidence of medusae consumption.

One aggregation of *L. lucerna* was found on the surface water of an embayment on the southernmost part of the Patos Lagoon, south Brazil (Figure 1A), on 25 March 2019. Medusae (N = 33) and associated crabs (N = 4) were collected individually using a dip net aboard small boats. Specimens were photographed, immediately preserved in 4% formaldehyde solution and transported to the Laboratory of Zooplankton at the Universidade Federal do Rio Grande (FURG). Biometric data of the carapace length between consecutive spines (LC2) was taken with the aid of a pachymeter (\pm 0.01 mm). The weight of the crabs was estimated (\pm 0.01 g). The stomachs of crabs were removed through a section on their carapace and the gastric contents were analyzed on slides under an inverted microscope at 1,000× magnification. After the identification of cnidae in the gastric content, these were compared with the cnidae of L. lucerna, obtained from tissues of the medusa oral arms and marginal lappets and analyzed on slides under a microscope at 1,000× magnification. These cnidae were compared and classified according to Östman (2000). The analyzed material was stored in the collection of the Laboratory of Zooplankton of FURG.

We found three medusae of L. lucerna in association with individuals of the blue crab Callinectes sapidus and one medusa with one crab of the species Callinectes ornatus. In all cases, only one crab was found per medusa (Figure 1 B and C). Both crab species were found on the subumbrella of the medusae and were juveniles, two females and two males. The mean carapace length (LC2) was 1.70 cm (0.97–2.05 cm) and the mean weight was 0.65 g (0.09–0.97 g). Gut content analyses of crabs revealed nematocysts of L. lucerna in all stomachs. These nematocysts were classified as oval-shaped small isorhiza and rhopaloids (Figure 1 C). The small (<10µm) isorhizae were characterized by an isodiametric tubule, without a proximal shaft, but the presence of spines could not be confirmed at 1000x magnification of the compound light microscope employed. The rhopaloids were characterized by a clearly distinct shaft and a coiled thinner tubule (Figure 1 D). The same nematocysts were found on oral arms and bell margins of *L*. *lucerna* (Figure 1 E) and both were similar in shape and size to those described for the rhizostome species Catostylus mosaicus and Phyllorhiza puncatata (Peach and Pitt, 2005). These

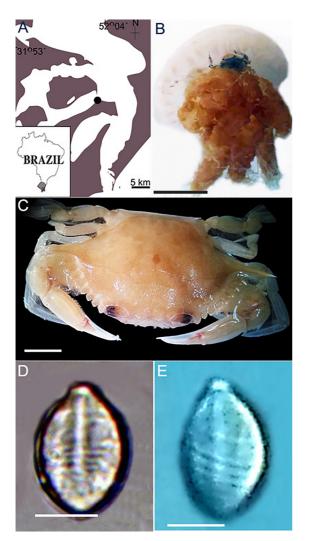


Figure 1 - A - Study site, in the Patos Lagoon Estuary, south Brazil. **B** - Specimen of *Lychnorhiza lucerna* with a crab *Callinectes sapidus* on its bell, scale: 4 cm. **C** - Detail of one specimen of *Callinectes sapidus*, found with *L. lucerna*, scale: 0.25 cm. **D** - Rhopaloid nematocyst from oral arms of *Lychnorhiza lucerna*, scale bar: 5 μm. E - Rhopaloid nematocyst found in gut content of *Callinectes sapidus*, scale bar: 5 μm.

nematocysts were the only prey item found in the gut contents of crabs, which characterized an opportunistic predation of scyphozoan jellyfish.

The extent of the damage caused by decapods feeding on jellyfishes can be varied. For spider crabs of the genus *Libinia*, it has been suggested that the damage to the hosts is not substantial and that nematocysts of *L. lucerna* found in the gut contents could have been incidentally ingested along with prey captured by the jellyfish (Moyano et al., 2012; Gonçalves et al., 2020). In contrast, decapods such as *Carcinus maenas*, *Crangon crangon*, and *Pagurus bernhardus* were able to actively capture and

consume comb jellyfish in laboratory experiments (Esser et al., 2004) or in the field (Ramussen, 1973; Farr, 1978). Predation by decapod crabs on jellyfish was reported even on occasions when other prey items were available (Ates, 2017). This is in contrast with the idea that jellyfish are consumed only in the absence of energy-richer prey (Farr, 1978). Specifically, regarding Callinectes crabs, Farr (1978) described individuals of C. sapidus actively feeding on the bells and oral arms of the medusae Cyanea capillata and Stomolophus meleagris. Based on this evidence, as well as the data, it was concluded that portunid crabs may consume large jellyfish, but it is not possible to determine whether the large number of dead or damaged jellyfish found without oral arms during sampling was a consequence of such predation.

Another non-exclusive explanation for the large number of dead/damaged jellyfish is the senility of individuals in this population during late summer. Although there is no study on the population dynamics of L. lucerna at the Patos Lagoon Estuary, other populations along the Southwest Atlantic are seasonal, as are most scyphozoan species (Schiariti et al. 2008; Nogueira Jr. & Haddad 2017). An evaluation of the jellyfish feeding structures suggested that most individuals showed a non-feeding post-reproductive condition, because of the lack or reduction of oral arms and the absence of digitata, the finger-like projections on oral arms that capture prey. This feature is common in scyphozoan jellyfishes that shed their reproductive and feeding structures and rapidly became senescent (Henschke et al. 2017). It is possible, therefore, that both factors (predation and senescence) contributed to the mortality of these jellyfishes. To clarify the role of predation by crabs and the interplay of this factor with the senescence of L. lucerna, further studies should investigate crab feeding rates and the extent of their damage to the jellyfishes. Even if the predation is less important to the jellyfish mortality, as suggested by the low frequency of crabs found on the latter (~12%), our study demonstrates an important fate for the large biomass that is seasonally produced by scyphozoan species in coastal areas.

The predation of benthic organisms may be increased when jellyfish aggregations are stuck in shallow enclosed waters and become more vulnerable to crabs, as well as to other consumers, such as hyperiid

amphipods, shrimps, free-living or parasitic anemones, and echinoderms (reviewed by Ates, 2017). In the Patos Lagoon Estuary, the tides are small amplitude (<40 cm); therefore, the entrance of coastal/oceanic waters with jellyfishes depends on the wind stress and lower rainfall (Odebrecht et al., 2017). After salinization of the estuary, the southernmost part of the Patos Lagoon can hold pockets of saltwater with aggregations of L. lucerna for weeks within small shallow (<2 m) embayments. In the Patos Lagoon Estuary, such episodes may be more frequent during the warmer months (December to May) when the rainfall is lower. These enclosed/shallow areas may act as sinks for aggregations/blooms of gelatinous organisms. In addition, research has shown that benthic consumers can control jellyfish populations (Esser et al., 2004) and alter pelagic food web structures (Sullivan et al., 1991). These interactions can be enhanced by turbulence (Esser et al., 2004), by the behavior of L. lucerna aggregating close to the bottom (Alvarez Colombo et al., 2003), and by the fact that these crabs can feed on pelagic prey (Rady et al., 2018).

Finally, the results demonstrate that portunid crabs of the genus Callinetes actively feed on L. lucerna and may benefit from blooms/ aggregations of this species. The possible role of these crabs in controlling jellyfish populations is still unknown and should be further investigated. Because these crabs are key species in estuaries and coastal areas, they may act as intermediate consumers, driving the energy produced by jellyfish to other consumers, such as the fishes Micropogonias furnieri, Genidens spp., and Paralonchurus brasiliensis, which are known predators of Callinectes (Figueiredo and Vieira, 2005; Bemvenuti and Colling, 2010). This trophic relationship illustrates examples of benthicpelagic coupling and the contribution of coastal/ oceanic production to the estuarine food web, which can also be found in many similar coastal ecosystems.

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AUTHOR CONTRIBUTIONS

- T.L.V.: Conceptualization; Data curation; Formal analysis; Writing - Original draft; Writing - review & editing.
- S.S.: Formal analysis; Investigation; Writing review & editing;
- R.M.N.: Investigation; Methodology; Formal Analysis; Supervision; Writing - review & editing.

REFERENCES

- ATES, R. M. L. 2017. Benthic scavangers and predators of jellyfish, material for a review. *Plankton Benthos Research*, 12(1), 71-77.
- BEMVENUTI, C. E. & COLLING L. A. 2010. Relações tróficas na comunidade bentônica da região estuarina da Lagoa dos Patos, RS, Brasil. *Cadernos de Ecologia Aquática*, 5(1), 1-8.
- BRANCO, J. O., LUNARDON-BRANCO, M. J., VERANI, J. R., SCHVEITZER, R., SOUTO, F. X. & VALE, W. G. 2002. Natural Diet of Callinectes ornatus Ordway, 1863 (Decapoda, Portunidae) in the Itapocoroy Inlet, Penha, SC, Brazil. Brazilian Archives of Biology and Technology, 45(1), 35-40.
- COLOMBO, G. A., MIANZAN, H. & MARIOLAS, A. 2003. Acoustic characterization of gelatinous-plankton aggregations: four case studies from the Argentine continental shelf. *ICES Journal of Marine Science*, 60, 650-657.
- D'AMBRA, I., GRAHAM, W. M., CARMICHAEL, R. H. & HERNANDEZ JUNIOR, F. J. 2015. Fish rely on scyphozoan hosts as a primary food source: evidence from stable isotope analysis. *Marine Biology*, 162, 247-252.
- EPIFANIO, C. E. 2007. Biology of larvae. *In:* KENNEDY, V. S. & CRONIN, L. E. (eds.). *The blue crab Callinectes sapidus*. Maryland: Maryland Sea Grant College.
- ESSER, M., GREVE, W. & BOERSMA, M. 2004. Effects of temperature and the presence of benthic predators on the vertical distribution of the ctenophore *Pleurobrachia pileus*. *Marine Biology*, 145(3), 595-601.
- FARR, J. A. 1978. Blue crab predation on jellyfish. *Florida Scientist*, 41(4), 217-218.
- FIGUEIREDO, G. M. & VIEIRA J. P. 2005. Diel feeding, daily food consumption and the predatory impact of withemouth croaker (*Micropogonias furnieri*) in an estuarine environment. *Marine Ecology*, 26(2), 130-139.
- FLEMING, N. E. C., HARROD, C., GRIFFIN, D. C., NEWTON, J. & HOUGHTON, J. D. R. 2014. Scyphozoan jellyfish provide short-term reproductive habitat for hyperiid amphipods in a temperate near-shore environment. *Marine Ecology Progress Series*, 510, 229-240.
- GONÇALVES, G. R. L., NEGREIROS-FRANSOZO, M. L., FRANSOZO, A. & CASTILHO, A. L. 2020. Feeding ecology and niche segregation of the spider crab *Libinia ferreirae* (Decapoda, Brachyura, Majoidea), a symbiont of *Lychnorhiza lucerna* (Cnidaria, Scyphozoa, Rhizostomeae). *Hydrobiologia*, 857(4), 1013-1025.
- GUERRA-CASTRO, E., CARMONA-SUÁREZ C. A. & CONDE J. E. 2007. Activity patterns and zonation of the swimming crabs Arenaeus cribrarius and Callinectes ornatus. Journal of Crustacean Biology, 27(1), 49-58.
- HAYS, G. C., DOYLE, T. K. & HOUGHTON, J. D. R. 2018. A paradigm shift in the trophic importance of jellyfish? *Trends in Ecology* and Evolution, 33(11), 874-884.

- HENSCHKE, N., STOCK, C. A. & SARMIENTO, J. L. 2018. Modeling population dynamics of scyphozoan jellyfish (*Aurelia* spp.) in the Gulf of Mexico. *Marine Ecology Progress Series*, 591, 167-183.
- HINES, A. H. 2007. Chapter 14: Ecology of juvenile and adult blue crabs. *In*: KENNEY, V. S. & Cronin, E. (eds.). *Biology of the blue crab*. College Park: Maryland Sea Grant Program, pp. 575-665.
- MANCINELLI, G., GUERRA, M. T., ALUJEVIĆ, K., RAHO, D., ZOTTI, M. & VIZZINI, S. 2017. Trophic flexibility of the Atlantic blue crab *Callinectes sapidus* in invaded coastal systems of the Apulia region (SE Italy): a stable isotope analysis. *Estuarine, Coastal and Shelf Science,* 198(Pt B), 421-431.
- MARTINELLI FILHO, J. E., STAMPAR, S. N., MORANDINI, A. C. & MOSSOLIN, E. C. 2008. Cleaner shrimp (Caridea: Palaemonidae) associated with scyphozoan jellyfish. *Vie Milieu Life and Environment*, 58(2), 133-140.
- MELO, G. A. S. 1996. Manual de Identificação dos Brachyura (caranguejos e siris) do litoral brasileiro. São Paulo: Plêiade/FAPESP.
- MOYANO, M. S., SCHIARITI, A., GIBERTO, D. A., BRIZ, L. D., GAVIO, M. A. & MIANZAN, H. W. 2012. The symbiotic relationship between Lychnorhiza lucerna (Scyphozoa, Rhizostomeae) and Libinia spinosa (Decapoda, Epialtidae) in the Río de la Plata (Argentina-Uruguay). Marine Biology, 159(9), 1933-1941.
- NAGATA, R. M., HADDAD, M. A. & NOGUEIRA JÚNIOR, M. 2009. The nuisance of medusae (Cnidaria, Medusozoa) to shrimp trawls in central part of southern Brazilian Bight, from the perspective of artisanal fishermen. *Pan-American Journal of Aquatic Sciences*, 4(3), 312-325.
- NAGATA, R. M. & MORANDINI, A. C. 2018. Diet, prey selection, and individual feeding rates of the jellyfish *Lychnorhiza lucerna* (Scyphozoa, Rhizostomeae). *Marine Biology*, 165(12), 187.
- NAGATA, R. M., MORANDINI, A. C., COLIN S. P., MIGOTTO, A. E. & COSTELLO, J. H. 2016. Transitions in morphologies, fluid regimes, and feeding mechanisms during development of the medusa *Lychnorhiza lucerna*. *Marine Ecology Progress Series*, 557,145-159.
- NOGUEIRA JÚNIOR, M., COSTA, B. S. P., MARTINEZ, T. A., BRANDINI, F. P. & MIYASHITA, L. K. 2019. Diversity of gelatinous zooplankton (Cnidaria, Ctenophora, Chaetognatha and Tunicata) from a subtropical estuarine system, southeast Brazil. *Marine Biodiversity*, 49(3), 1283-98.
- NOGUEIRA JÚNIOR, M. & HADDAD, M. A. 2005. *Lychnorhiza lucerna* (Scyphozoa, Rhizostomeae) and *Libinia ferreirae* Brito Capello (Brachyura, Majidae) association in Southern Brasil. *Revista Brasileira de Zoologia*, 22(4), 908-912.
- NOGUEIRA JÚNIOR, M. & HADDAD, M. A. 2017. Seasonal distribution, abundance and biomass of large medusae in subtropical coast of Brazil. *In:* MARIOTINI, L. (ed.). *Jellyfish: ecology, distribution patterns and human interactions.* New York: Nova Publishers, pp. 3-26.

- ODEBRECHT, C., SECCHI, E. R., ABREU, P. C., MUELBERT, J. H. & UIBLEIN, F. 2017. Biota of the Patos Lagoon estuary and adjacent marine coast: long-term changes induced by natural and humanrelated factors. *Marine Biology Research*, 13(1), 3-8.
- OLIVEIRA, A., PINTO, T. K., SANTOS, D. P. D. & D'INCAO, F. 2006. Natural diet of the blue crab *Callinectes sapidus* (Decapoda, Portunidae) in the Patos Lagoon estuary area, Rio Grande, Rio Grande do Sul, Brasil. *Iheringia, Série Zoologia*, 96(3), 305-313.
- ÖSTMAN, C. 2000. A guideline to nematocyst nomenclature and classification, and some note on the systematic value of nematocysts. *Scientia Marina*, 64(Suppl 1), 31-46.
- PEACH, M. B., & PITT, K. A. 2005. Morphology of the nematocysts of the medusae of two scyphozoans, *Catostylus mosaicus* and *Phyllorhiza punctata* (Rhizostomeae): implications for capture of prey. *Invertebrate Biology*, 124(2), 98-108.
- PURCELL, J. E. & ARAI, M. N. 2001. Interactions of pelagic cnidarians and ctenophores with fish: a review. *Hydrobiologia*, 451, 27-44.
- RADY, A., SALLAM, W. S., ABDOU, N. E. I. & EL-SAYED, A. A. M. 2018. Food and feeding habits of the blue crab, *Callinectes sapidus* (Crustacea: Decapoda: Portunidae) with special reference to the gastric mill structure. *Egyptian Journal of Aquatic Biology and Fisheries*, 22(spe5), 417-431.
- RASMUSSEN, E. 1973. Systematics and ecology of the Isefjord marine fauna (Denmark). *Ophelia*, 11(1), 1-507.
- SCHIARITI, A., KAWAHARA, M., UYE, S. & MIANZAN, H. W. 2008. Life cycle of the jellyfish *Lychnorhiza lucerna* (Scyphozoa: Rhizostomeae). *Marine Biology*, 156(1), 1-12.
- SILVEIRA, S., ORTEGA, I. & DUMONT, L. F. C. 2020. Artisanal trawling impact over prey availability and diet of estuarine megabenthic organisms in southern Brazil. *Estuarine, Coastal and Shelf Science*, 237, e106682.
- SULLIVAN, B. K., DOERING, P. H., OVIATT, C. A., KELLER, A. A. & FRITHSEN, J. B. 1991. Interactions with the benthos alter pelagic food web structure in coastal waters. *Canadian Journal of Fisheries and Aquatic Sciences*, 48, 2276-2284.
- SWEETMAN, A. K., SMITH, C. R., DALE, T. & JONES, D. O. B. 2014. Rapid scavenging of jellyfish carcasses reveals the importance of gelatinous material to deep-sea food webs. *Proceedings of the Royal Society B*, 281, 1-8.
- TOWANDA, T. & THUESEN, E. V. 2006. Ectosymbiotic behavior of Cancer gracilis and its trophic relationships with its host Phacellophora camtschatica and the parasitoid Hyperia medusarum. Marine Ecology Progress Series, 315, 221-236.
- WILLIAM, A. B. 1974. The swimming crabs of the genus *Callinectes* (Decapodae: Portunidae). *Fishery Bulletin*, 72(3), 685-198.
- YEAGER, L. A., KREBS, J. M., MCIVOR, C. C. & BRAME, A. B. 2007. Juvenile blue crab abundances in natural and man-made tidal channels in mangrove habitat, Tampa Bay, Florida (USA). Bulletin of Marine Science, 80(3), 555-565.