Revista da Sociedade Brasileira de Medicina Tropical

Journal of the Brazilian Society of Tropical Medicine Vol.:53:e20180498: 2020 AND THE PROPERTY OF THE PARTY O

doi: 10.1590/0037-8682-0498-2018

Short Communication

Pseudomonas aeruginosa in the ICU: prevalence, resistance profile, and antimicrobial consumption

Ághata Cardoso da Silva Ribeiro^[1], Márcia Terezinha Lonardoni Crozatti^[1],
Adilson Aderito da Silva^[2], Rodrigo Spineli Macedo^[3],
Antonia Maria de Oliveira Machado^[4]
and Antonio Távora de Albuquerque Silva^[1]

[1]. Universidade Federal de São Paulo, Instituto de Ciências Ambientais, Químicas e Farmacêuticas, Diadema, SP, Brasil.
[2]. Universidade Presbiteriana Mackenzie, Centro de Ciências Sociais e Aplicadas, São Paulo, SP, Brasil.
[3]. Universidade Federal de São Paulo, Hospital Universitário da UNIFESP/Hospital São Paulo, São Paulo, SP, Brasil.
[4]. Universidade Federal de São Paulo, Departamento de Medicina. Laboratório Central do Hospital São Paulo, São Paulo, SP, Brasil.

Abstract

Introduction: *Pseudomonas aeruginosa* is one of the main pathogens causing infection in intensive care units (ICUs) and usually presents antimicrobial resistance. **Methods:** Data were obtained from ICUs between 2010 and 2013. **Results:** *P. aeruginosa* had a prevalence of 14.5% of which 48.7% were multidrug resistant. We observed increasing resistance to carbapenems and polymyxin B and growing consumption of aminoglycosides, meropenem, ceftazidime, and polymyxin B. The regression impact between resistance and consumption was significant with respect to amikacin, imipenem, meropenem, and polymyxin B. **Conclusions:** Monitoring antimicrobial consumption and resistant microorganisms should be reinforced to combat antimicrobial- and multidrug resistance.

Keywords: Pseudomonas aeruginosa. Antimicrobial agents. Drug resistance.

Antimicrobial resistance is a public health concern¹. Among resistant pathogens, the bacterium *Pseudomonas aeruginosa* is of note because it is often associated with high rates of morbidity and mortality in patients in intensive care units (ICUs)². *P. aeruginosa* is resistant to several antibiotics and has the ability to rapidly develop resistance to new antimicrobials. Moreover, *P. aeruginosa* was the first bacterium to present multidrug-resistant (MDR) phenotypes³.

Therefore, it is important to monitor the consumption of antibiotics, especially in a hospital environment, to avoid the development of resistance and improve the therapeutic efficacy of these drugs against *P. aeruginosa*.

To address the above issues, this study aimed to describe the prevalence of resistant and MDR isolates of *P. aeruginosa* in nine ICUs of a university hospital, as well as to describe

Corresponding author: Ághata Cardoso da Silva Ribeiro.

e-mail: aghata.cardoso@unifesp.br Orcid: 0000-0002-4997-0505 Received 22 November 2018 Accepted 11 July 2019 the consumption of antimicrobial agents using a defined daily dose (DDD) and investigate the relationship between the consumption of and resistance to antibiotics in the nine ICUs.

This was a retrospective descriptive study, conducted from January 2010 to December 2013 in ICUs of the Hospital São Paulo - university hospital (HSP-HU/Unifesp) in the city of São Paulo (Brazil).

For resistance and susceptibility analysis, data from the Clinical Laboratory of the hospital were used. Samples were obtained from patients hospitalized in at least one of the ICUs selected during the study period.

P. aeruginosa isolates were considered to be MDR if they showed resistance to three or more classes of antimicrobial agents, with resistance to at least one antibiotic in each class⁴.

Data on the consumption of antimicrobials (in grams) were provided by the Supplies System of the hospital, and the hospitalization rate in the study period was provided by the Hospital Statistics Sector. Antimicrobial agents were classified

according to the Anatomical Therapeutic Chemical system, and consumption was measured in DDD per 100 bed-days⁵.

Susceptibility tests performed on samples from anal and rectal swabs for *Klebsiella pneumoniae* carbapenemase surveillance and vancomycin-resistant enterococci surveillance were excluded from the study.

For the organization and analysis of the data, Microsoft® Office Excel for Mac, version 16.27 (Microsoft Corporation) and the Statistical Package for Social Sciences version 20 (IBM Corp, Chicago, IL, USA) were used.

The resistance trend of P. aeruginosa, monthly consumption of antimicrobial agents, and the association between antimicrobial agent consumption and resistance of P. aeruginosa were analyzed using a linear regression model. The β coefficient indicates the association between a dependent variable and independent variables analyzed by the linear regression model. The significance level was set at 0.05.

The remaining analyses were for normality, linearity, and homoscedasticity. For normality analyses, the Kolmogorov-Smirnov (KS) test (significance>0.05) was performed and all the variables obtained were considered normal, except for the data on consumption of ceftazidime and polymyxin resistance. The homoscedasticity and linearity analyses were conducted by graphical analyses. Homoscedasticity was analyzed using the standardized variables against the standardized predicted values.

This study is part of the project "Prevalent microorganisms in ICU: antimicrobial agents consumption and resistance profiles," approved by the Research Ethics Committee (opinion 921.500), in accordance with Resolution No. 466 of December 12, 2012, of the National Health Council.

Of 6,473 microorganisms isolated, 939 were identified as *P. aeruginosa* (14.5%). *P. aeruginosa* was the second most prevalent bacterium during the study period, after *Acinetobacter baumannii* (17.3%). The prevalence of *P. aeruginosa* isolates resistant to antimicrobial agents by year is presented in **Table 1**.

Increasing resistance of *P. aeruginosa* to imipenem (β =0.419; p=0.003), meropenem (β =0.485; p=0.000), and polymyxin B (β =0.401, p=0.005) was statistically significant. The prevalence of resistance to imipenem was 50% in January 2010 and 71.4% in December 2013; that of resistance to meropenem was 44.4% in January 2010 and 71.4% in December 2013, while the prevalence of resistance to polymyxin B was zero in January 2010 and 7.1% in December 2013. However, significantly decreasing resistance of *P. aeruginosa* to amikacin (β =-0.345; p=0.016) and piperacillin/tazobactam (β =-0.301, p=0.038) were observed (**Table 2**).

It is noteworthy that 457 (48.7%) of the 939 isolates of *P. aeruginosa* were classified as MDR *P. aeruginosa* (PaMDR), with a higher prevalence in the years 2010 (51.2%) and 2013 (52.9%).

Consumption of amikacin (β =0.793; p=0.000), gentamicin (β =0.746; p=0.000), meropenem (β =0.571; p=0.000), ceftazidime (β =0.336; p=0.020), and polymyxin B (β =0.625;

p=0.000) were significantly increasing. In contrast, consumption of imipenem (β =-0.875; p=0.000) and cefepime (β =-0.825; p=0.000) were significantly decreasing (**Table 2**).

In addition, significant associations were observed between the consumption of and resistance of *P. aeruginosa* to amikacin (β =-0.361; p=0.012), imipenem (β =-0.316; p=0.029), meropenem (β =0.327; p=0.023), and polymyxin B (β =0.351; p=0.014) (**Table 3**).

In this study, *P. aeruginosa* was the second most frequently isolated bacterium, representing 14.5% of the total isolates, which demonstrates the clinical importance of this microorganism. The results obtained are consistent with those of other studies, such as the one performed in ICUs of a public hospital in Ceará (Brazil), wherein *P. aeruginosa* represented 33.8% of the isolates⁶, and that performed in an ICU of Anesthesiology in Turkey, wherein the prevalence of *P. aeruginosa* was 14.29%², demonstrating that the high prevalence of this microorganism is common in various countries.

The resistance profiles of *P. aeruginosa* against the tested antibiotics are presented in **Table 1**. It is noteworthy that Biswall et al. (2014), in India, reported similar resistance prevalence results for aztreonam (41.38%) and piperacillin/tazobactam (34.5%), as well as zero resistance to colistin (or polymyxin E). However, the results were different for amikacin (81.03%), gentamicin (81.03%), ceftazidime (70.68%), imipenem (18.9%), and meropenem (13.79%)⁷. Based on these data, there may be variations in microbial resistance profiles depending on the study site, which shows the importance of determining resistance profiles to be able to select the most appropriate antimicrobial therapy.

It is also worth noting that the prevalence of resistance of P. aeruginosa to imipenem and meropenem showed an increasing trend in the study period. These data concur with the results of a study carried out at a hospital in the state of Santa Catarina (Brazil), which indicated that resistance to carbapenems was increasing. Thus, in 2004, the resistance of *P. aeruginosa* to imipenem was 6.06%, increasing to 48.09% in 2008; for meropenem, the resistance increased from 6.89% in 2004 to approximately 20% in 20088. Similarly, Xu et al. (2013) in a study performed at a hospital in China found increasing resistance to imipenem $(\beta=4.620; p<0.001)$ and meropenem $(\beta=4.624; p<0.001)^9$. Although carbapenems are one of the most potent classes of antimicrobials against P. aeruginosa, increasing resistance to these drugs is an endemic problem in several countries; therefore, the World Health Organization (WHO) considers the development of new drugs for this microorganism a priority¹⁰.

In addition, the present study showed a decreasing prevalence of resistance of *P. aeruginosa* to amikacin and piperacillin/tazobactam, which differs from the results of Xu et al. (2013), who showed that resistance to amikacin (β =0.488; p=0.332) and piperacillin/tazobactam (β =4.346; p<0.001) was increasing⁹.

Although the resistance of *P. aeruginosa* to polymyxin B was found to be low (0.9%) in the present study, resistance to

TABLE 1: Prevalence of *P. aeruginosa* isolates resistant to the tested antibiotics, based on laboratory findings of patients in nine ICUs from January 2010 to December 2013.

	Total Total resis		resistant	ntResistant isolates								
Class / Antimicrobial agent	isolates (S+I+R)	isolates		2010			2011		2012		2013	
	n	N	%	n	%	N	%	n	%	n	%	
Aminoglycosides												
Amikacin	925	263	28.4	98	37.3	67	25.5	63	24.0	35	13.	
Streptomycin	11	6	54.6	1	16.7	0	0.0	4	66.7	1	16.	
Gentamicin	938	434	46.3	146	33.6	92	21.2	87	20.0	109	25.	
Tobramycin	53	34	64.2	0	0.0	4	11.8	25	73.5	5	14.	
Carbapenems												
Ertapenem	23	10	43.5	0	0.0	0	0.0	1	10.0	9	90	
Imipenem	931	477	51.2	127	26.6	116	24.3	108	22.6	126	26	
Meropenem	930	473	50.9	129	27.3	112	23.7	103	21.8	129	27	
Cephalosporins												
Cefepime	928	410	44.2	132	32.2	92	22.4	81	19.8	105	25	
Cefotaxime	26	26	100.0	8	30.8	6	23.1	4	15.4	8	30	
Cefoxitin	1	1	100.0	0	0.0	0	0.0	0	0.0	1	100	
Ceftazidime	920	366	39.8	124	33.9	73	19.9	67	18.3	102	27	
Ceftriaxone	35	32	91.4	2	6.3	1	3.1	5	15.6	24	75	
Fluoroquinolones												
Ciprofloxacin	928	459	49.5	155	33.8	103	22.4	85	18.5	116	25	
Levofloxacin	3	3	100.0	2	66.7	0	0.0	0	0.0	1	33	
Moxifloxacin	2	0	0.0	0	0.0	0	0.0	0	0.0	0	0.	
Norfloxacin	90	49	54.4	27	55.1	12	24.5	7	14.3	3	6.	
Monobactams												
Aztreonam	924	377	40.8	101	26.8	87	23.1	83	22.0	106	28	
Polymyxins												
Polymyxin B	766	7	0.9	0	0.0	1	14.3	0	0.0	6	85	
Penicillins												
Amoxicillin	31	28	90.3	1	3.6	1	3.6	3	10.7	23	82	
Ampicillin	15	12	80.0	2	16.7	1	8.3	6	50.0	3	25	
Oxacillin	2	2	100.0	2	100.0	0	0.0	0	0.0	0	0.	
Penicillin	9	7	77.8	0	0.0	1	14.3	4	57.1	2	28	
Piperacillin/Tazobactam	929	334	36.0	130	38.9	83	24.9	58	17.4	63	18	
Macrolides												
Erythromycin	2	2	100.0	2	100.0	0	0.0	0	0.0	0	0.	
Lincosamides												
Clindamycin	2	2	100.0	2	100.0	0	0.0	0	0.0	0	0.	
Oxazolidinones		_										
Linezolid	11	0	0.0	0	0.0	0	0.0	0	0.0	0	0.	
Sulfonamides					0.0							
Sulfamethoxazole/												
	7	6	85.7	2	33.3	0	0.0	2	33.3	2	33	
Trimethoprim												
Glycopeptides		_		_		_		_		_	_	
Teicoplanin	10	5	50.0	0	0.0	0	0.0	2	40.0	3	60	
Vancomycin	11	8	72.7	0	0.0	1	12.5	3	37.5	4	50	
Tetracyclines												
Tigecycline	17	2	11.8	1	50.0	0	0.0	0	0.0	1	50	
Others												
Ethambutol	2	0	0.0	0	0.0	0	0.0	0	0.0	0	0.	
Isoniazid	2	0	0.0	2	0.0	0	0.0	0	0.0	0	0.	
Nitrofurantoin	2	1	50.0	0	0.0	0	0.0	0	0.0	1	100	
Pyrazinamide	2	0	0.0	0	0.0	0	0.0	0	0.0	0	0.	
Rifampicin	4	1	25.0	1	100.0	0	0.0	0	0.0	0	0.	

I: intermediate; $\mbox{\bf R} :$ resistant; $\mbox{\bf S} :$ susceptible.

TABLE 2: Trends in the prevalence of resistance of *P. aeruginosa* to and consumption of antimicrobial agents of clinical interest in nine ICUs from January 2010 to December 2013.

Antimicrobial —— agent	Trend in	n the prevaler	nce of P. aeruginos	sa resistance	Trend in antimicrobial agent consumption				
	β	р	Trend	KS significance	β	р	Trend	KS significance	
Amikacin	-0.345	0.016	decreasing	0.974	0.793	0.000	increasing	0.445	
Gentamicin	0.037	0.802	increasing	0.769	0.746	0.000	increasing	0.845	
Ciprofloxacin	0.040	0.786	increasing	0.966	0.188	0.200	increasing	0.341	
Imipenem	0.419	0.003	increasing	0.927	-0.875	0.000	decreasing	0.904	
Meropenem	0.485	0.000	increasing	0.754	0.571	0.000	increasing	0.301	
Cefepime	0.163	0.269	increasing	0.470	-0.825	0.000	decreasing	0.575	
Ceftazidime	0.217	0.138	increasing	0.967	0,336	0.020	increasing	0.016	
Piperacillin/ Tazobactam	-0.301	0.038	decreasing	0.673	0.245	0.094	increasing	0.889	
Polymyxin B	0.401	0.005	increasing	0.010	0.625	0.000	increasing	0.210	

β coefficient: indicates the association between a dependent variable and independent variables by a linear regression model. The significance level was set at 0.05. **KS significance**: indicates the significance of the Kolmogorov-Smirnov normality test. Variables were considered normal when KS>0.005.

TABLE 3: Association between the consumption of antimicrobial agents of clinical interest and the prevalence of *P. aeruginosa* resistance in nine ICUs from January 2010 to December 2013.

Antimicrobial agent	β	р	Impact	KS significance		
Amikacin	-0.361	0.012	Negative	0.774		
Gentamicin	0.082	0.579	Positive	0.858		
Ciprofloxacin	-0.002	0.992	Negative	0.881		
Imipenem	-0.316	0.029	Negative	0.350		
Meropenem	0.327	0.023	Positive	0.534		
Cefepime	-0.239	0.102	Negative	0.399		
Ceftazidime	-0.199	0.176	Negative	0.982		
Piperacillin/ Tazobactam	-0.062	0.675	Negative	0.953		
Polymyxin B	0.351	0.014	Positive	0.001		

β coefficient: indicates the association between a dependent variable and independent variables by a linear regression model. The significance level was set at 0.05. **KS significance**: indicates the significance of the Kolmogorov-Smirnov normality test. Variables were considered normal when KS>0.005.

this antibiotic was observed to increase (β =0.401, p=0.005). This result is worth mentioning as polymyxins are considered the last line of defense against gram-negative bacteria and have been classified by the WHO as critically important for human medicine¹¹.

Among the isolates of *P. aeruginosa*, 48.7% were classified as MDR. This result is similar to the prevalence of 30.5% observed in a multicenter study of patients admitted to hospitals in Europe and the United States⁴. Similarly, a study carried out at a Spanish hospital indicated that 33.3% of *P. aeruginosa* isolates were classified as MDR¹². It is also worth noting that in Brazil, Neves et al. (2010) reported a frequency of 23.1% of PaMDR, in a study carried out at the Clinics Hospital of Botucatu of the School of Medicine of the State of São Paulo¹³.

Therefore, it is important to note that the treatment options for PaMDR are limited. Most PaMDR are susceptible to polymyxins (polymyxin B and colistin). Aminoglycosides may be another alternative³.

Considering the consumption of antimicrobial agents, increasing tendency for the consumption of amikacin and gentamicin was observed in the present study. However, Vega et al. (2015) observed a decrease in the consumption (in DDD/100 patients-day) of amikacin (from 0.87 in 2008 to 0.31 in 2011) and an increase in the consumption of gentamicin (from 1.17 in 2008 to 10.67 in 2011) in a study carried out in an adult ICU in Argentina¹⁴. In another study, an increase in the consumption (in DDD/100 patients-days) of both amikacin (from 1.48 in 1999 to 1.91 in 2008) and gentamicin (from 5.02 in 1999 to 6.88 in 2008) was observed in ICUs in the Czech Republic¹⁵.

In the present study, a downward trend in imipenem consumption was identified, while meropenem consumption was increasing. In the study by Vojtová et al. (2011), increasing consumption of both imipenem (from 0.33 in 1999 to 1.60 in 2008) and meropenem (from 0.88 in 1999 to 3.56 in 2008) was observed¹⁵.

In addition, a decreasing trend in the consumption of cefepime was observed in the present study, whereas ceftazidime showed an increasing trend. In the study by Vega et al. (2015), the consumption (in DDD/100 beds-day) of cefepime and ceftazidime decreased between 2008 and 2011 from 10.19 to 1.48 and 2.2 to 0.25, respectively¹⁴.

Furthermore, a growing trend of consumption was found for polymyxin B, perhaps because of the increased prevalence of PaMDR in the study period and because polymyxins are currently the last option for monotherapy against resistant and PaMDR³, and are therefore extensively used for this purpose.

By analyzing the correlation between the prevalence of resistance to and the consumption of antimicrobial agents, we observed a negative association between the consumption of amikacin and imipenem and prevalence of *P. aeruginosa* resistance to these antibiotics. However, even though the consumption of amikacin increased during the study period, we cannot conclude that the decrease in the prevalence of resistance was only due to the possible effectiveness of the antimicrobial

since it is known that the decrease in prevalence of bacterial resistance also depends on other factors¹ such as increased surveillance, campaigns to combat the spread of resistance, and changes in microbial resistance profiles.

It is still noteworthy that a positive impact and association between the consumption of meropenem and polymyxin B and the prevalence of resistance of *P. aeruginosa* to these antibiotics were detected. Xu et al. (2013) obtained similar results for meropenem (β =1.241; p<0.001) but different results for imipenem (β =1.238; p<0.001)⁹. As for polymyxin B, the result obtained is worrisome as polymyxins are the last option for monotherapy against resistant and PaMDR³. However, it is important to note that in the present study, *P. aeruginosa* showed a low resistance profile to polymyxin B.

The present study has the following limitations: (i) the presented consumption data were general data for the nine selected ICUs, not specific data for patients with *P. aeruginosa* infection; and (ii) other factors that influence resistance were not considered, although antimicrobial consumption is one of the most important factors¹.

Thus, a prospective study using specific antimicrobial consumption data and data from each patient with *P. aeruginosa* infection, as well as the analysis of all associated factors and/or resistance influencers, may help better establish the correlations between bacterial resistance and antimicrobial consumption, as well as between bacterial resistance and other associated factors and/or resistance influencers.

In the present study, we determined the prevalence of resistant and MDR isolates of *P. aeruginosa*, as well as the levels of antibiotic consumption in ICUs of a university hospital in São Paulo (Brazil). The results of this work emphasize the importance of monitoring the consumption of antimicrobial agents and combating the increase of microbial resistance, including multidrug resistance.

ACKNOWLEDGMENTS

We express our gratitude to Universidade Federal de São Paulo (UNIFESP) which supported this work by providing the data of the study population.

Conflict of Interest

The authors declare that there are no conflicts of interest.

REFERENCES

- World Health Organization (WHO). Antimicrobial Resistance. Global Report on surveillance [Internet]. 1th ed. Geneva: WHO Press; 2014 Apr [cited 2018 Nov 22]. 256p. Available from: http://apps.who.int/iris/bitstream/handle/10665/112642/9789241564748_eng.pdf?sequence=1
- Dereli N, Ozayar E, Degerli S, Sahin S, Koç F. Three-Year Evaluation of Nosocomial Infection Rates of the ICU. Braz J Anesthesiol. 2013;63(1):73-8.
- Fraimow HS, Tsigrelis C. Antimicrobial Resistance in the Intensive Care Unit: Mechanisms, Epidemiology, and management of Specific Resistant Pathogens. Crit Care Clin. 2011;27(1):163-205.

- 4. Micek ST, Wunderink RG, Kollef MH, Chen C, Rello J, Chastre J et al. An international multicenter retrospective study of *Pseudomonas aeruginosa* nosocomial pneumonia: impact of multidrug resistance. Crit Care. 2015;19:219-27.
- World Health Organization Collaborating Centre for Drug Statistics Methodology - WHOCC [Internet]. Norway: Norwegian Institute of Public Health; 2015 [updated 2017 Dec 20; cited 2018 Nov 22]. Available from: http://www.whocc.no/atc_ddd_index/
- Barros LM, Bento JNC, Caetano JA, Moreira RAN, Pereira FGF, Frota NM et al. Prevalência de micro-organismos e sensibilidade antimicrobiana de infecções hospitalares em unidade de terapia intensiva de hospital público no Brasil. Rev Ciênc Farm Básica e Apl. 2012;33(3):429-35.
- Biswall I, Arora BS, Kasana D, Neestushree. Incidence of Multidrug Resistant *Pseudomonas aeruginosa* Isolated from Burn Patients and Environment of Teaching Institution. J Clin Diagn Res. 2014;8(5):26-9.
- Baumgart AMK, Molinari MA, Silveira ACO. Prevalence of carbapenem resistant *Pseudomonas aeruginosa* and *Acinetobacter Baumannii* in high complexity hospital. Braz J Infect Dis. 2010;14(5):433-6.
- Xu J, Duan X, Wu H, Zhou Q. Surveillance and Correlation of Antimicrobial Usage and Resistance of *Pseudomonas aeruginosa*: A Hospital Population-Based Study. PLoS ONE [Internet]. 2013 Nov 8 [cited 2018 Nov 22]; 8(11):e78604. Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3826718/

- 10. World Health Organization (WHO). Critically important antimicrobials for human medicine [Internet]. 5rd ed. Geneva: World Health Organization; 2016; June [cited 2018 Nov 22]. 41p. Available from: http://apps.who.int/iris/bitstream/hand le/10665/255027/9789241512220-eng.pdf;jsessionid=4CA3754731 D70DB15F9C963004DE86F0?sequence=1
- World Health Organization (WHO). Global priority list of antibiotic-resistant bacteria to guide research, discovery, and development of new antibiotics [Internet]. Geneva: WHO; 2016 [cited 2018 Nov 20].
 Available from: http://www.who.int/medicines/publications/WHO-PPL-Short_Summary_25Feb-ET_NM_WHO.pdf?ua=1
- Morales E, Cots F, Sala M, Comas M, Belvis F, Riu M et al. Hospital costs of nosocomial multi-drug resistant *Pseudomonas aeruginosa* acquisition. BMC Health Serv Res. 2012;12(122):1-8.
- 13. Neves MT, Lorenzo MEP, Almeida RAMB, Fortaleza CMB. Antimicrobial use and incidence of multidrug-resistant *Pseudomonas aeruginosa* in a teaching hospital: an ecological approach. Rev Soc Bras Med Trop. 2010;43(6):629-32.
- Vega EM, Fontana D, Iturrieta M, Segovia L, Rodríguez G, Agüero S. Consumo de antimicrobianos en la Unidad de Terapia Intensiva del Hospital Dr. Guillermo Rawson-San Juan, Argentina. Rev Chilena Infectol. 2015;32(3):259-65.
- Vojtová V, Kolár M, Hricová K, Uvízl R, Neiser J, Blahut L et al. Antibiotic utilization and *Pseudomonas aeruginosa* resistance in intensive care units. New Microbiol. 2011;34(3):291-8.