

Original Article

Stability of vancomycin hydrochloride employed in antimicrobial seal solutions of central intravenous catheters*

Daniele Porto Barros^{1,2} https://orcid.org/0000-0002-4388-3793

Priscilla Sete de Carvalho Onofre^{1,2}

Fernando Luiz Affonso Fonseca³ (b) https://orcid.org/0000-0003-1223-1589

Paulo César Pires Rosa⁴ https://orcid.org/0000-0003-3665-8878

Mavilde da Luz Gonçalves Pedreira¹ https://orcid.org/0000-0002-9246-2354

Maria Angélica Sorgini Peterlini¹ bttps://orcid.org/0000-0003-1769-4662

- * Paper extracted from doctoral dissertation "Stability of vancomycin hydrochloride for antimicrobial lock technique used for decontamination of central intravenous catheters", presented to Universidade Federal de São Paulo, Escola Paulista de Enfermagem, São Paulo, SP, Brazil. This article refers to the call "Innovation in practice, teaching or research in health and nursing". Supported by Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq), Grant # 311296/2013-0 and 462183/2014-9, Brazil.
- ¹ Universidade Federal de São Paulo, Escola Paulista de Enfermagem, São Paulo, SP, Brazil.
- ² Scholarship holder at the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES), Brazil.
- ³ Universidade Estadual de Campinas, Faculdade de Ciências Farmacêuticas, Campinas, SP, Brazil.
- ⁴ Universidade Federal de São Paulo, Instituto de Ciências Ambientais, Químicas e Farmacêuticas, Diadema, SP, Brazil.

Highlights: (1) Both temperatures led to a change in pH and concentration of the antimicrobial. **(2)** Vancomycin hydrochloride at 22°C obtained less variation in chemical behavior. **(3)** Association of the drugs resulted in a change in physical stability. **(4)** Association of the drugs resulted in a solution with physical incompatibility.

Objective: to verify the stability of vancomycin hydrochloride in antimicrobial seal solutions with and without association of heparin sodium according to temperature and association time. Method: an experimental study designed for the analysis of hydrogenionic potential and concentration by means of high-efficiency liquid chromatography of vancomycin hydrochloride (n=06) and vancomycin hydrochloride and heparin sodium (n=06). The solutions studied were submitted to absence of light, as well as to 22°C and 37°C. Analyses in triplicate (n=192) were performed at the initial moment (T0) and three (T3), eight (T8) and 24 hours (T24) after preparation. The data were submitted to analysis of variance ($p \le 0.05$). Results: concentration of the antimicrobial at 22°C presented a reduction (T0-T8) and a subsequent increase (T24); hydrogenionic potential decreased significantly over time. At 37°C, the concentration increased up to T3 and decreased at T24, with a reduction of hydrogenionic potential up to 24 hours. Concentration of the vancomycin hydrochloride and heparin sodium solutions varied with a reduction at 22°C, accompanied by increased hydrogenionic potential. Precipitate formation was observed by visual inspection of the vancomycin hydrochloride-heparin sodium association (T3). Conclusion: pharmacological stability of vancomycin hydrochloride (5 mg/mL) and physical incompatibility with heparin sodium (100 IU/mL) were evidenced after three hours of association in the antimicrobial seal solutions studied.

Descriptor: Catheter-Related Infections; Drug Stability; Infusions, Intravenous; Vancomycin; Heparin; Nursing.

How to cite this article

Barros DP, Onofre PSC, Fonseca FLA, Rosa PCP, Pedreira MLG, Peterlini MAS. Stability of vancomycin hydrochloride employed in antimicrobial seal solutions of central intravenous catheters. Rev. Latino-Am. Enfermagem. 2022;30:e3621.

_____. https://doi.org/10.1590/1518-8345.5869.3621

Introduction

Critically-ill patients often require multiple drugs, almost all of them intravenously, in order to guarantee the plasma levels in the concentration and time required for an adequate therapeutic response⁽¹⁾. Consequently, the use of central intravenous catheters (CICs) has been indicated, due to the characteristics of most of the drugs employed, osmolarity and hydrogenionic potential (pH), among others⁽²⁻⁴⁾.

CIC preservation in patients with chronic diseases is essential. However, prolongation of the use time of these devices is a predisposing factor for central catheter-related bloodstream infection (CCRBSI)⁽⁵⁻⁶⁾, due to biofilm formation in the lumen of the catheter⁽⁷⁾. The traditional intervention involves removal of the device. However, this is not always feasible for these patients due to the restricted vascular network resulting from multiple punctures and treatment, as well as from the clinical condition, factors that hinder invasive interventions or procedures.

Thus, use of the seal technique with antimicrobials in CICs emerges as an adjuvant in the treatment^(2,5). It comprises the administration of antimicrobials with a strong recommendation for the use of vancomycin hydrochloride^(5,8-9) in the lumen of the catheter at a concentration 100 to 1,000 times higher than the minimum inhibitory value usually employed for systemic therapy⁽⁸⁾, being frequently used in combination with heparin sodium⁽¹⁰⁻¹¹⁾.

The solution used in the lumen of the CIC has the function of decontamination, being a *sine qua non* condition for maintaining stability of the drugs within the device during the period indicated for the expected effect^(8,11).

A number of studies indicate that the antimicrobial seal may remain in the lumen of the intravenous catheter for a long period of time, in order to overcome the barrier formed by the microbial biofilm^(8-9,11-12).

However, there is no evidence related to the maximum volume to be administered, permanence time of the solution in the catheter lumen, use frequency and adequate concentration, as well as the association with an anticoagulant^(5,8,12). Most clinical studies propose a minimum of eight hours a day to achieve the expected action^(8-9,12). However, there is also the recommendation for the solution to remain between 24 and 48 hours inside the CIC⁽⁸⁾.

Thus, the concern about antimicrobial stability in association with heparin sodium remains. Although these phenomena are dependent, among other factors, on concentration of the drugs, exposure to higher temperatures and contact time between the medications should also be evaluated^(9,11-12). Thus, the objective of this study was to verify the physical-chemical stability of vancomycin hydrochloride in antimicrobial seal solutions with and without heparin sodium according to temperature and association time.

Method

Study design

An experimental research study, mimicking the clinical practice of antimicrobial seal administration in CICs.

Study locus

Laboratory of Nursing Experiments (*Laboratório de Experimentos de Enfermagem*, LEEnf) of the São Paulo Nursing School, Federal University of São Paulo, São Paulo, SP, Brazil.

Data collection period

Data collection took place between May and July 2018.

Samples

Vancomycin hydrochloride solutions in physiological serum (PhS) and in association with heparin sodium, equivalent to those employed in the antimicrobial seal technique in central intravenous catheters.

Sample definition

The sample consisted of 12 solutions, namely: three of vancomycin hydrochloride at 22°C, three of vancomycin hydrochloride at 37°C, three containing the association of antimicrobial and heparin sodium at 22°C, and three containing the association of antimicrobial and heparin sodium at 37°C. The solutions were prepared by a single researcher and the pH and concentration measurements occurred at the initial moment (T0) and at three (T3), eight (T8) and 24 hours (T24). Concentration checks were carried out in triplicates, resulting in 192 measurements (48 for pH and 144 for concentration).

Study variables

Simulating the clinical practice of antimicrobial seal administration, the experiments took place under controlled light and temperature conditions. The 37°C temperature aimed at mimicking body temperature. The drug concentrations were based on the Infectious Diseases Society of America (IDSA) Practice Guidelines⁽¹³⁾. Two different solutions were studied: vancomycin hydrochloride (5 mg/mL) and the association of the antimicrobial (5 mg/mL) with heparin sodium (100 I.U./mL). The conditions for the analysis programmed in the chromatograph were

as follows: mobile phase (MP) in isocratic mode, flow at 1 mL/min; 30°C column temperature, ultraviolet (UV) detector with wavelength at 220 nanometers (nm), injection of 20 microliters (μ L) and analytical running time of 14 minutes (min.).

Instruments used for collecting the information

The pH of the solutions was studied in a digital bench top meter (Kasvi® K39-2014B, Curitiba, Brazil) and the antimicrobial concentration by means of the highefficiency liquid chromatography analytical methodology - HPLC (HPLC Modular Agilent Technologies[®] - 1260 Infinity Series HPLC), using a 4.6 x 250 mm C18 reverse phase column and particle size of 5 micrometers (µm) (Thermo Scientific[®] - ODS Hypersil, Massachussetts, United States of America, batch: 13359). For the daily cleaning procedure of the analytical column of the HPLC equipment, a solution composed of the mixture was established in the proportion of 80% methanol and 20% deionized water, for 20 minutes, with a 1 mL/min flow. Subsequently, the chromatographic parameters were programed. The conditions for the analysis programmed in the equipment were as follows: Mobile phase - MP (acetonitrile 8% and ionic pairing solution 92%) in isocratic mode, flow at 1 mL/min; 30°C column temperature, UV detector with wavelength at 220 nm, injection of 20 µL and analytical running time of 14 minutes. Preparation of the solutions took place in a gas exhaust chapel (Sppencer® SP1050-25, São Paulo, Brazil), with aseptic technique, with polypropylene syringes (3 mL and 10 mL) and sterile stainless steel needles (30.0 x 0.8 mm). Vancomycin hydrochloride (500 mg), heparin sodium (5,000 IU/mL) and PhS in a 100 mL flexible transparent polypropylene bag were used. The drugs, diluents and disposable materials used in the experiment were from the same manufacturing batch and respected shelf life. The experimental solutions were packed in amber glass bottles in order to mimic the absence of light to which the solutions are subjected when inside the catheters.

Data collection

Validation of the analytical methodology was carried out by means of HPLC, prior to verifying the

vancomycin hydrochloride concentration, based on the premises established in RE Resolution Number 166 of 2017 of the National Health Surveillance Agency (*Agência Nacional de Vigilância Sanitária*, ANVISA) ⁽¹⁴⁾, in the ICH Q2(R1) guide of the 2005 International Conference on Harmonization⁽¹⁵⁾, and in the United States Pharmacopeia - USP 32⁽¹⁶⁾ for the selectivity, linearity, precision and accuracy parameters. After preparing the solutions, visual inspection and pH control were performed. To analyze the vancomycin hydrochloride concentrations, it was necessary to adapt to the chromatographic condition with adjustment of the drug sample for the linear working range of 0.1 mg/mL.

Data treatment and analysis

The results are presented based on the means of the triplicates of the pH checks and those of the concentrations in percentage and milligrams *per* milliliters, calculated taking into account the chromatographic peak area of vancomycin hydrochloride of each sample in the times established. Statistical analysis was performed using the multilevel linear regression model, analysis of variance (ANOVA) and multiple comparisons with Bonferroni correction. A 5% significance level was adopted for all the statistical analyses. The statistical analyses were performed using the SPSS 20.0 and Stata 12 programs.

Ethical aspects

The study was submitted to the evaluation of Unifesp's Research Ethics Committee (*Comitê de Ética em Pesquisa*, CEP) and approved under opinion No. 8942030117.

Results

The results of the concentration and pH study were obtained by analyzing 12 vancomycin hydrochloride solutions and solutions containing the vancomycin hydrochloride-heparin sodium association. Table 1 shows the results of the concentration and pH study corresponding to the solutions according to time and temperature variations. 4

Table 1 – Concentration and hydrogenionic potential of vancomycin hydrochloride solutions (n=6) and of the vancomycin hydrochloride-heparin sodium association (n=6) according to temperature and time. São Paulo, SP, Brazil, 2018

Temperature	Type of solution	Moment	Concentration (%) Mean ± SD (Min-Max)	Concentration (mg/mL) Mean ± SD (Min-Max)	Hydrogenioninc potential Mean ± SD (Min-Max)
22°C	Vancomycin hydrochloride solution	TO	101.93 ± 1.66*	5.10 ± 0.08*	3.76 ± 0.05*
			(100.75-104.33)	(5.04-5.22)	(3.73-3.82)
		Т3	101.45 ± 1.46	5.07 ± 0.07	3.73 ± 0.01*
			(99.54-103.58)	(4.98-5.18)	(3.73-3.74)
		Т8	100.31 ± 0.87 [†]	$5.02 \pm 0.04^{+}$	3.74 ± 0.01*
			(99.39-101.39)	(4.97-5.07)	(3.73-3.75)
		T24	102.50 ± 1.46*	5.12 ± 0.07*	$3.59 \pm 0.03^{\dagger}$
			(100.52-104.44)	(5.03-5.22)	(3.56-3.62)
		р	<0.05	<0.05	<0.05
	Vancomycin hydrochloride and heparin sodium solution	ТО	101.73 ± 3.81*	5.09 ± 0.19*	$3.63 \pm 0.03^{\dagger}$
			(95.50-106.64)	(4.78-5.33)	(3.60-3.65)
		ТЗ	99.01 ± 1.56 [†]	$4.95 \pm 0.08^{\dagger}$	$3.59 \pm 0.02^{\dagger}$
			(97.49-101.24)	(4.87-5.06)	(3.58-3.61)
		Т8	97.38 ± 1.20 [†]	4.87 ± 0.06 [†]	$3.64 \pm 0.04^{\dagger}$
			(95.60-98.80)	(4.78-4.94)	(3.61-3.69)
		T24	96.34 ± 1.70 ⁺	$4.82 \pm 0.08^{\dagger}$	4.00 ± 0.02*
			(94.36-99.44)	(4.72-4.97)	(3.99-4.02)
		р	<0.05	0.05	<0.05
37°C	Vancomycin hydrochloride solution	ТО	97.79 ± 0.69 [†]	4.89 ± 0.03 [†]	3.73 ± 0.02*
			(96.99-98.97)	(4.85-4.95)	(3.71-3.74)
		Т3	101.96 ± 1.13*	5.10 ± 0.06*	$3.69 \pm 0.02^{\dagger}$
			(100.25-102.92)	(5.01-5.15)	(3.68-3.71)
		Т8	100.73 ± 1.69*	5.04 ± 0.08*	3.67 ± 0.01 [†]
			(98.77-102.58)	(4.94-5.13)	(3.66-3.68)
		T24	98.35 ± 2.80 ⁺	$4.92 \pm 0.14^{\dagger}$	$3.68 \pm 0.02^{\dagger}$
			(95.41-101.83)	(4.77-5.09)	(3.66-3.69)
		р	<0.05	<0.05	0.05
	Vancomycin hydrochloride and heparin sodium solution	то	94.91 ± 1.76 ⁺	$4.75 \pm 0.09^{\dagger}$	3.80 ± 0.02
			(92.68-96.80)	(4.63-4.84)	(3.78-3.82)
		Т3	95.12 ± 1.29 ⁺	$4.76 \pm 0.06^{\dagger}$	3.75 ± 0.07
			(93.19-96.80)	(4.66-4.84)	(3.68-3.82)
		Т8	95.05 ± 3.15 ⁺	4.75 ± 0.16 [†] (4.54-4.97)	3.75 ± 0.04
			(90.84-99.43)		(3.72-3.79)
		T24	96.94 ± 1.23*	$4.85 \pm 0.06^{*}$	3.78 ± 0.01
			(94.94-98.42)	(4.75-4.92)	(3.78-3.79)
		р	0.001	0.001	0.389

Analysis of variance (ANOVA); 'Multiple comparisons Bonferroni correction; () and (*) present different means between the evaluation moments for each. The results are expressed through mean ± standard deviation

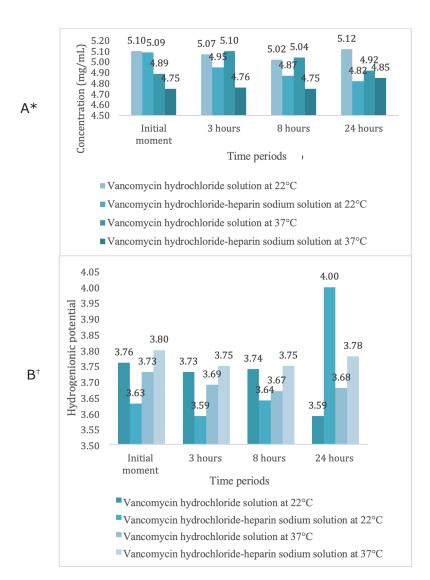
Table 1 shows a reduction in the concentration of vancomycin hydrochloride at 22°C at T0 and T8 with a subsequent increase in T24. Regarding the pH, there was a reduction over time.

Under the influence of the 37°C temperature, the vancomycin hydrochloride concentration was increased at the initial moment, as well as at three hours of preparation with a subsequent decrease at 24 hours (Table 1). Regarding pH, there was no significant variation in relation to the values found in the vancomycin hydrochloride solutions at 22°C with a reduction in the pH scale from the initial moment up to 24 hours, as shown in Table 1.

In relation to the vancomycin hydrochloride-heparin sodium association solution at 22°C, regarding pH, a lower value was observed at three hours of preparation, and a higher value at 24 hours. Regarding the concentration, it presented a drop of around 5% in the antimicrobial values over time (Table 1).

In the vancomycin hydrochloride with heparin sodium solutions, when submitted to 37°C and with regard to pH, it is noted that they presented a higher value at T0, although with a smaller variation. As for the concentration, they presented a drop at the initial moment, reaching their maximum concentration 24 hours after preparation of the solutions (Table 1).

To graphically show the variation in concentration and pH obtained in the study, the results, according to the preparation time of the solutions, are presented in Figure 1.

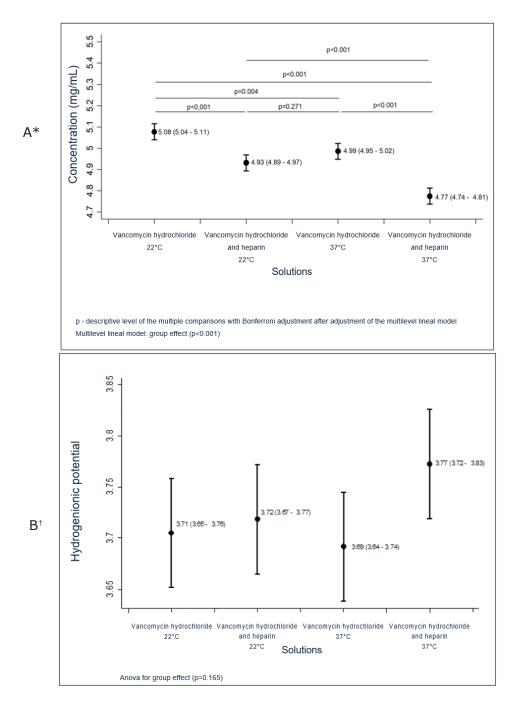


*A = Concentration; * B = Hydrogenionic potential

Figure 1 – Mean values of concentration and hydrogenionic potential of the vancomycin hydrochloride solutions and of those of the vancomycin hydrochloride-heparin sodium association, according use time and exposure temperature. São Paulo, SP, Brazil, 2018

Figure 1 shows that the 22°C temperature caused less variation in the concentrations; however, in the solutions containing the association of the antimicrobial with heparin sodium, it was possible to observe a reduction in concentration, even at 22°C. However, in the vancomycin hydrochloride-heparin sodium solution at 37°C, immediately after preparation, the concentration started with a value lower than the expected theoretical concentration (5 mg/mL). Regarding the analysis of antimicrobial pH at both temperatures, it was observed that the values remained similar, except for 24 hours at 22°C, which proved to be more acidic (Table 1).

Multiple comparisons were made with vancomycin hydrochloride solutions and solutions containing vancomycin hydrochloride in association with heparin sodium, at both temperatures and in the four time intervals established, as shown in Figure 2.



*A = Concentration; ⁺B = Hydrogenionic potential

Figure 2 – Concentration and hydrogenionic potential of the vancomycin hydrochloride solutions and of those containing the vancomycin hydrochloride-heparin sodium association, according to the influence of time and temperature on multiple paired comparisons (*a posteriori* Bonferroni test). São Paulo, SP, Brazil, 2018

The multiple mathematical comparisons performed in the study of the vancomycin hydrochloride solutions showed a statistically significant reduction in concentration of the antimicrobial in association with heparin sodium with pH elevation in the solutions submitted to 37°C (Figure 2). Visual inspection of the solutions evidenced precipitate formation in the vancomycin hydrochlorideheparin sodium association at three hours, regardless of temperature (Figure 3).

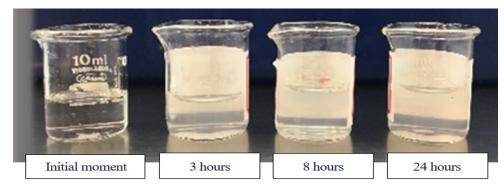


Figure 3 - Vancomycin hydrochloride and heparin sodium solution at the different moment: initial; and after three, eight and 24 hours of use of the solutions at a temperature of 22°C; and with formation of a cloudy solution at three hours of preparation. São Paulo, SP, Brazil, 2018

Discussion

The current study was carried out by means of a mimicking process of the clinical practice of the seal technique with antimicrobials in CICs, at temperatures of 22°C and 37°C. The concentrations of the vancomycin hydrochloride solutions at both temperatures were close to 100% (theoretical expected concentration of 5 mg/mL); they remained equivalent during the 24 hours after preparation, which corroborates the research regarding the evaluation of vancomycin hydrochloride concentration by the HPLC method that revealed pharmacological stability of the antimicrobial for up to 24 hours at room temperature⁽¹⁷⁾.

However, the association of vancomycin hydrochloride with heparin sodium resulted in precipitate formation at three hours at both temperatures, as well as in reduced antimicrobial concentration when at 37°C.

It is known that the predominant profile of drug incompatibilities with potential risks for the patient is related to the class of antimicrobials, and the association between drugs may inactivate the active ingredient of the product or lead to toxicity, resulting in uncertainty of therapeutic efficacy⁽¹⁸⁻¹⁹⁾. Recognition of pharmacological incompatibilities makes it possible to avoid adverse situations, as well as the emergence of toxicity⁽²⁰⁻²¹⁾.

To ensure the efficacy of vancomycin hydrochloride in decontamination of CICs, concentration of the drug should be maintained at a variation of less than 10% throughout the period in which the solution remains inside the intravenous device, also taking into account the materials of the CICs⁽²²⁾. Analyzing the concentrations of the vancomycin hydrochloride solution at 22°C in the current experiment, in which the solutions remained packed in amber glass vials, it is noted that, despite the tenuous change in the values, the greatest variation occurred within periods of eight and 24 hours with a 2% increase in concentration of the antimicrobial at 24 hours of use of the solution (p>0.05), which decants the probability of the influence of time on stability of the drug.

Regarding pH, the vancomycin hydrochloride solution at 22°C presented a reduction over time, accentuating the acid profile of the drug. In this situation, an extreme acidity value of all measurements was obtained; however, pH behavior over time was similar. Such values comprise the antimicrobial reference range from 2.5 to 5.5 according to the manufacturer⁽²³⁾.

A study on drug stability states that extreme pH values can cause instability of the solutions and are considered key elements for physical-chemical compatibility of the solutions⁽²⁴⁾.

Changes in the temperature to which the solutions are submitted can cause pH changes and consequent pharmacological instability^(18,21,25). However, in this research, regarding pH, the vancomycin hydrochloride solutions remained equivalent over time at 37°C. In addition, when comparing the vancomycin hydrochloride solutions at 22°C with the antimicrobial solutions at 37°C, a similar behavior is noted with reduced values over time and with a variation of less than 0.1 points in the pH scale.

When submitted to the 37°C temperature, the solution containing vancomycin hydrochloride in

association with heparin sodium presented a higher pH mean value than the others found in the study, although with a reduction of 0.02 pH points from the initial moment to 24 hours after preparation. The greatest variation occurred in the solutions containing the association of the antimicrobial with heparin sodium at 22°C, with an increase of 0.37 pH points over time. Therefore, it can be suggested that in this experiment, high temperature did not affect the degree of concentration of hydrogen ions of the vancomycin hydrochloride solutions; however, there was a change in relation to the reduction of the antimicrobial concentration when in association with heparin sodium after 24 hours.

According to the American Pharmacopoeia⁽¹⁶⁾, the drug should comprise 90% to 110% of the labeled concentration. When comparing the concentrations of the solutions immediately after preparation, it was possible to observe a variation in the vancomycin hydrochloride concentration at the initial moment of preparation of the vials, although within the tolerance limit established. However, when evaluating the vancomycin hydrochloride concentration when in association with heparin sodium at 22°C, it was possible to observe a reduction of more than 5% in the values from the initial moment to 24 hours of use of the solutions.

When establishing a comparison in this research between the solutions submitted to 22°C, it was possible to observe that the greatest variation in concentration occurred in the association with heparin sodium, with decreased antimicrobial concentration over time. When submitted to 37°C, the solution containing vancomycin hydrochloride and heparin sodium presented lower concentration values than the others found in the experiment. Regarding pH, the highest value obtained in the experiment was achieved in the vancomycin hydrochloride and heparin sodium solutions at 22°C at 24 hours, reaching a value of 4.0; therefore, 0.41 points higher than the value obtained in the antimicrobial solution at the same temperature and in the same period of time, a fact that suggests the interference of the anticoagulant in stability of vancomycin hydrochloride.

Solutions of vancomycin hydrochloride isolated or associated with heparin sodium were identified as the best alternatives for the treatment of CCRBSIs for Grampositive microorganisms⁽²⁶⁻²⁷⁾. However, with regard to drug stability, the results were contradictory, especially for antimicrobial-heparin sodium association solutions since, although in one study⁽²⁸⁾ vancomycin hydrochloride (5 mg/ mL) was compatible with heparin sodium (2.5000 IU/ mL), others, in turn, indicated that high concentrations of antimicrobial (10 mg/mL) can increase the risk for precipitation of the solutions, even with low doses of anticoagulant (100 IU/mL)⁽²⁹⁾. In this study, when at 22°C, there was a reduction in the vancomycin hydrochloride concentration after the association with heparin sodium in the evaluation of the solution after 3 hours of the 2.72% preparation. In a similar research study, a reduction in antimicrobial concentration was demonstrated in solutions containing vancomycin hydrochloride and heparin sodium inside CICs after 72 hours, suggesting, among other things, the interaction between the drugs as well as between the solutions and the intravenous device material⁽³⁰⁾.

A study conducted in a hospital found that most of the medication errors observed were related to drug incompatibility or to lack of evidence for administration associated to the drugs⁽³¹⁾. Drug incompatibility occurs when two or more drugs react or interact in a way that there is a change in the normal activity of one or more components, a fact that can derail clinical therapy, resulting in alteration of the active ingredient, precipitation or clouding of the solution and change in the color of the drug^(20-21,32).

A research on study vancomycin hydrochloride stability showed precipitate formation and turbidity when in association with heparin sodium from five minutes of solution preparation⁽²⁹⁾. In the current study, changes were observed in the physical aspect of the solutions containing vancomycin hydrochloride in association with heparin sodium, evidenced by turbidity with precipitate formation at three hours at temperatures of 22°C and 37°C, suggesting interaction between the drugs. In addition, the formation of precipitate may result in obstruction of the intravenous device and/or risk of causing embolism in the patient, and the association between them is not recommended.

Feasibility of the occurrence of drug incompatibility and the scarcity of scientific evidence are difficulties found in nurses' daily practice. It is to be remembered that performing drug associations without scientific evidence incurs in medication errors⁽³³⁻³⁴⁾. With regard to concentration of the drugs and solutions, it is possible that this may cause pharmacological instability and compromise effectiveness of the seal technique in CICs. Therefore, in order to better portray the clinical practice, it is suggested to investigate stability of vancomycin hydrochloride with a possible interference of different CIC materials, linked to the permanence time in the administration system as well as in the association between drugs and solutions.

The following stand out as study limitations: conduction of the experiments in glass vials, as well as the use of a single dosage of heparin sodium. Additionally, only one concentration of the antimicrobial (5 mg/mL) was performed. For future studies, it is suggested to investigate stability and concentration of vancomycin hydrochloride, when in association with heparin sodium under the possible interference of the material of intravenous devices and at a concentration of 10 mg/mL.

Conclusion

There was a change in physical stability in the vancomycin hydrochloride (5 mg/mL) and heparin sodium (100 I.U./mL) solution at three hours of association with a reduction of less than 10% in antimicrobial concentration, evidencing chemical instability with drug degradation, although with pharmacological stability.

The vancomycin hydrochloride solutions maintained at 22°C presented the smallest variations in pH. However, temperature does not seem to be an environmental factor triggering considerable differences in the chemical behavior of the solutions studied.

Thus, this study evidenced that, regardless of temperature, the association of vancomycin hydrochloride with heparin sodium in solutions used in seals for CIC decontamination presented chemical instability and drug incompatibility. Despite maintenance of pharmacological stability, it is therefore recommended to adopt non-use of these compounds in association in the seal technique with an antimicrobial.

Acknowledgments

We thank the members of the SEGTEC - Segurança e Tecnologia Research Group - Nursing research study group on patient safety, intensive care and intravenous therapy in pediatrics (Brazil) for their support during data collection.

To the team of the Laboratory of Nursing Experiments (LEEnf).

References

1. Sandoval LJS, Lima FET, Gurgel SS, Freitas I, Barbosa LP, Almeida PC. Translation and cross-cultural adaptation of the instrument patient safety in pediatric drug administration: spanish version. Esc Anna Nery. 2021;25(4). https://doi.org/10.1590/2177-9465-EAN-2020-0333

 Anttila VJ. Central venous catheter care for children with cancer should focus on early infections. Acta Paediatr.
Feb;108(2):204-5. https://doi.org/10.1111/ apa.14548

3. Benlabed M, Perez M, Gaudy R, Genay S, Lannoy D, Barthélémy C, et al. Clinical implications of intravenous drug incompatibilities in critically ill patients, Anaesth Crit Care Pain Med. 2019;38(2):173-80. https://doi. org/10.1016/j.accpm.2018.04.003

4. Manrique-Rodríguez S, Heras-Hidalgo I, Pernia-López MS, Herranz-Alonso A, Pisabarro MCDR, Suárez-Mier MB,

et al. Standardization and chemical characterization of intravenous therapy in adult patients: a step further in medication safety. Drugs RD. 2021;21:39-64. https:// doi.org/10.1007/s40268-020-00329-w

5. Ardura MI. Overview of infections complicating pediatric hematopoietic cell transplantation. Infect Dis Clin North Am. 2018;32(1):237-52. https://doi.org/10.1016/j. idc.2017.11.003

6. Torre FPF, Baldanzi G, Troster EJ. Risk factors for vascular catheter-related bloodstream infections in pediatric intensive care units. Rev Bras Ter Intensiva. 2018;30(4):436-42. https://doi.org/10.5935/0103-507X.20180066

7. Higgins M, Zhang L, Ford R, Brownlie J, Kleidon T, Rickard CM, et al. The microbial biofilm composition on peripherally inserted central catheters: a comparison of polyurethane and hydrophobic catheters collected from paediatric patients. J Vasc Access. 2021;22(3):388-93. https://doi.org/10.1177/1129729820932423

8. Qureshi S, Fatima P, Mukhtar A, Zehra A, Qamar FN. Clinical profile and outcome of antibiotic lock therapy for bloodstream infections in pediatric hematology/oncology patients in a tertiary care hospital, Karachi, Pakistan. Int J Pediatr Adolesc Med. 2019 Mar;6(1):25-8. https://doi. org/10.1016/j.ijpam.2019.01.004

9. Liang H, Zhang L, Guo X, Sun L. Vancomycin-lock therapy for prevention of catheter-related bloodstream infection in very low body weight infants. BMC Pediatr. 2021;21(3). https://doi.org/10.1186/s12887-020-02482-2

10. Bookstaver PB, Premnath P, Wright K, McInnes C. Compatibility and stability of daptomycin lock solutions in combination with gentamicin, azithromycin, heparin and trisodium citrate. Preprints 2019;2019090107. https://doi.org/10.20944/preprints201909.0107.v1

11. Ranch-Lundin M, Schedin A, Björkhem-Bergman L. Equal effect of vancomycin lock with or without heparin in treatment of central venous catheter related blood stream infections - an observational study in palliative home care. Infect Dis (Lond). 2021 Sep;53(9):719-23. https://doi. org/10.1080/23744235.2021.1922752

12. Fatima M, Akash MSH, Rasool MF, Rehman K. Chemical kinetics and its applications in drug stability. In: Akash MSH, Rehman K, editors. Drug Stability and Chemical Kinetics [Internet]. Singapore: Springer; 2020 [cited 2021 Dec 9]. Available from: https://www.semanticscholar.org/paper/Drug-Stability-and-Chemical-Kinetics-Akash-Re hman/2eed3743025cb69bdc4fd360c3fa459a433eb97b 13. Liu C, Bayer A, Cosgrove SE, Daum RS, Fridkin SK, Gorwitz RJ, et al. Diretrizes práticas da Infectious Diseases Society of America para o tratamento de infecções por *Staphylococcus aureus* resistente à meticilina em adultos e crianças. Clin Infect Dis [Internet]. 2011 Fev [cited

2021 Dec 9];52(3):e18-55. Available from: https://www. sbp.com.br/fileadmin/user_upload/pdfs/diretrizes_idsa_ tratamento_infec_mrsa_adul_crian.pdf

14. Ministério da Saúde (BR), Agência Nacional de Vigilância Sanitária. Resolução RDC no 166, de 24 de julho de 2017. Dispõe sobre a validação de métodos analíticos e dá outras providências [Internet]. Diário Oficial da União, 25 jul. 2017 [cited 2021 Dec 9]. Available from: https://www.in.gov.br/materia/-/asset_publisher/Kujrw0TZC2Mb/ content/id/19194581/do1-2017-07-25-resolucao-rdc-n-166-de-24-de-julho-de-2017-19194412

15. International Conference on Harmonisation (ICH), Expert Working Group. Harmonised tripartite guideline validation of analytical procedures: text and methodology Q2(R1). In: International Conference on Harmonisation of Technical Requirements for Registration of Pharmaceuticals for Human Use [Internet]. Geneva: ICH Expert Working Group; 2005 [cited 2021 Dec 9]. Available from: https:// database.ich.org/sites/default/files/Q2%28R1%29%20 Guideline.pdf

16. United States Pharmacopeial Convention. USP 32-NF 27 [Homepage]. Rockville, MD: USP Convention; 2009 [cited 2021 Dec 9]. Available from: https://www.uspnf. com/official-text/proposal-statuscommentary/usp-32-nf-27 17. Masse M, Genay S, Mena AM, Carta N, Lannoy D, Barthélémy C, et al. Evaluation of the stability of vancomycin solutions at concentrations used in clinical services. Eur J Hosp Pharmacy. 2020;27:e87-e92. https:// doi.org/10.1136/ejhpharm-2019-002076

18. Kuscu F, Ulu A, Inal AS, Suntur BM, Aydemir H, Gul S, et al. Potential drug-drug interactions with antimicrobials in hospitalized patients: a multicenter point-prevalence study. Med Sci Monit. 2018 Jun 20;24:4240-7. https://doi.org/10.12659/MSM.908589

19. Smeringaiova I, Nyc O, Trosan P, Spatenka J, Burkert J, Bednar J, et al. Antimicrobial efficiency and stability of two decontamination solutions. Cell Tissue Bank. 2018;19:581-9. https://doi.org/10.1007/s10561-018-9707-0

20. Begum S, Reddy Y, Divya B, Komali P, Sushmitha K, Ruksar S. Pharmaceutical incompatibilites: a review. Asian J Pharm Res Dev. 2018;6(6):56-61. https://doi.org/https://doi.org/10.22270/ajprd.v6i6.448

21. Wilder AG, Foushee JA, Fox LM, Navalle J, Wright AM, Greer MA. Physical compatibility of medications used in critically ill patients with balanced fluid solutions. Int J Pharm Compd [Internet]. 2020 May-Jun [cited 2021 Dec 9];24(3):238-41. Available from: https://ijpc.com/Abstracts/Abstract.cfm?ABS=4804

22. Trissel LA Handbook on Injectable Drugs. 19. ed. Bethesda, MD: ASHP; 2017.

23. Antibióticos do Brasil Ltda. Vancomicina: o antibacteriano glicopeptídeo mais utilizado no mundo

tem novas diretrizes de uso. Rev Hospital in Foco. 2009 Set;1(3).

24. Maison O, Tardy C, Cabelguenne D, Parat S, Ducastelle S, Piriou V, et al. Drug incompatibilities in intravenous therapy: evaluation and proposition of preventive tools in intensive care and hematology units. Eur J Clin Pharmacol. 2019 Feb;75(2):179-87. https://doi.org/10.1007/ s00228-018-2602-6 Irshad K, Akash MSH, Rehman K, Imran I. Principles of pharmaceutical analysis in drug stability and chemical kinetics. In: Akash MSH, Rehman K, editors. Drug Stability and Chemical Kinetics [Internet]. Singapore: Springer; 2020 [cited 2021 Dec 9]. Available from: https://www.semanticscholar.org/paper/Drug-Stability-and-Chemical-Kinetics-Akash-Rehman/2eed37 43025cb69bdc4fd360c3fa459a433eb97b

25. Freire MP, Pierrotti LC, Zerati AE, Benites L, Motta-Leal JM Filho, Ibrahim KY, et al. Role of lock therapy for long-term catheter-related infections by multidrug-resistant bacteria. Antimicrob Agents Chemother. 2018 Aug 27;62(9):e00569-18. https://doi.org/10.1128/AAC.00569-18

26. Alonso B, Fernández-Cruz A, Díaz M, Sánchez-Carrillo C, Martín-Rabadán P, Bouza E, et al. Can vancomycin lock therapy extend the retention time of infected long-term catheters? APMIS. 2020;128(6):433-9. https://doi. org/10.1111/apm.13033

27. Shahkarami S, Mehdizadeh M, Roshanzamiri S, Tavakoli Ardakani M. Effect of vancomycin as antibiotic lock technique in prevention of catheter-associated infection in stemcell transplantation patients: vancomycin lock after HSCT. Int Pharmacy Acta. 2021;4(1):4e10:1-8. https:// doi.org/10.22037/ipa.v4i1.35723

28. Najari Z, Rusho WJ. Compatibility of commonly used bone marrow transplant drugs during Y-site delivery. Am J Health Syst Pharm. 1997 Jan 15;54(2):181-4. http:// doi.org/10.1093/ajhp/54.2.181

29. Vercaigne LM, Sitar DS, Penner SB, Bernstein K, Wang GQ, Burczynski FJ. Antibiotic-heparin lock: in vitro antibiotic stability combined with heparin in a central venous catheter. Pharmacotherapy. 2000 Apr;20(4):394-9. https://doi.org/10.1592/phco.20.5.394.35063

30. Sabzi Z, Mohammadi R, Talebi R, Roshandel GR. Medication errors and their relationship with care complexity and work dynamics. Open Access Maced J Med Sci. 2019 Oct 10;7(21):3579-83. https://doi.org/10.3889/ oamjms.2019.722

31. Sriram S, Aishwarya S, Moithu A, Sebastian A, Kumar A. Intravenous drug incompatibilities in the intensive care unit of a tertiary care hospital in India: are they preventable? J Res Pharm Pract. 2020 Apr-Jun;9(2):106--11. https://doi.org/10.4103/jrpp. JRPP_20_11

32. Sutherland A, Canobbio M, Clarke J, Randall M, Skelland T, Weston E. Incidence and prevalence of

intravenous medication errors in the UK: a systematic review. Eur J Hosp Pharm. 2020 Jan;27(1):3-8. https://doi.org/10.1136/ejhpharm-2018-001624

33. Elliott RA, Camacho E, Jankovic D, Sculpher MJ, Faria R. Economic analysis of the prevalence and clinical and economic burden of medication error in England. BMJ Qual Saf. 2021 Feb;30(2):96-105. https://doi.org/10.1136/bmjqs-2019-010206

34. Härkänen M, Turunen H, Vehviläinen-Julkunen K. Differences between methods of detecting medication errors: a secondary analysis of medication administration errors using incident reports, the global trigger tool method, and observations. J Patient Saf. 2020;16(2):168-76. https://doi.org/10.1097/PTS.00000000000261

Authors' contribution

Study concept and design: Fernando Luiz Affonso Fonseca, Paulo César Pires Rosa, Mavilde da Luz Gonçalves Pedreira, Maria Angélica Sorgini Peterlini. Obtaining data: Daniele Porto Barros, Priscilla Sete de Carvalho Onofre, Fernando Luiz Affonso Fonseca, Paulo César Pires Rosa, Data analysis and interpretation: Daniele Porto Barros, Priscilla Sete de Carvalho Onofre, Fernando Luiz Affonso Fonseca, Paulo César Pires Rosa, Mavilde da Luz Gonçalves Pedreira, Maria Angélica Sorgini Peterlini. Statistical analysis: Daniele Porto Barros, Priscilla Sete de Carvalho Onofre, Mavilde da Luz Gonçalves Pedreira. Obtaining financing: Maria Angélica Sorgini Peterlini. Drafting the manuscript: Daniele Porto Barros, Priscilla Sete de Carvalho Onofre, Mavilde da Luz Gonçalves Pedreira, Maria Angélica Sorgini Peterlini. Critical review of the manuscript as to its relevant intellectual content: Fernando Luiz Affonso Fonseca, Paulo César Pires Rosa, Mavilde da Luz Gonçalves Pedreira, Maria Angélica Sorgini Peterlini. All authors approved the final version of the text.

Conflict of interest: the authors have declared that there is no conflict of interest.

Received: Dec 9th 2021 Accepted: Mar 28th 2022

> Associate Editor: Maria Lúcia Zanetti

Copyright © 2022 Revista Latino-Americana de Enfermagem This is an Open Access article distributed under the terms of the Creative Commons (CC BY).

This license lets others distribute, remix, tweak, and build upon your work, even commercially, as long as they credit you for the original creation. This is the most accommodating of licenses offered. Recommended for maximum dissemination and use of licensed materials.

Corresponding author: Daniele Porto Barros E-mail: danipbarros@yahoo.com.br b https://orcid.org/0000-0002-4388-3793