

Testicular structure of three species of neotropical freshwater pimelodids (Pisces, Pimelodidae)

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ABSTRACT. The testes structure of the following Siluriformes was studied: *Pimelodus maculatus* (Lacepède, 1803), *Iheringichthys labrosus* (Lütken, 1874) and *Conorhynchus conirostris* (Valenciennes, 1840). The testes of these species were dissected into cranial and caudal regions. The fringes of mature testes were counted and measured. Student's t-test was used to identify significant differences between fringe lengths of the two regions. To study the whole testes of each species in its resting and mature stage, testes were fixed in Bouin liquid for 6-8 hours and submitted to routine histological techniques. Standard histochemical techniques were used to detect carbohydrates and proteins. The testes of all species were paired and fringed organs. Histologically, cranial fringes of all species were spermatogenic with cells inside cysts at the same phase of development. Caudal fringes of *C. conirostris* were spermatogenic; while *P. maculatus* and *I. labrosus* showed caudal fringes only during secretion. Histochemically, neutral glycoproteins, acid glycoconjugates, acid carboxilates, sialomucines, and acid and sulfates glycoconjugates were detected in the caudal fringe secretions of *P. maculatus*; and neutral glycoproteins in *I. labrosus*. Significant differences between the fringe lengths of the cranial and caudal regions were found for all the species studied. **KEY WORDS.** Fringed testes, secretion, seminal vesicle, Siluriformes, testicular gland.

RESUMO. Estudou-se a estrutura testicular dos seguintes peixes Siluriformes: *Pimelodus maculatus* (Lacepède, 1803), *Iheringichthys labrosus* (Lütken, 1874) e *Conorhynchus conirostris* (Valenciennes, 1840). Os testículos foram dissecados e divididos anatomicamente em regiões cranial e caudal. As franjas dos testículos em maturação foram contadas e mediu-se o comprimento das mesmas. O teste t de Student foi utilizado para verificar diferenças significativas entre o comprimento das franjas das duas regiões. Para estudo histológico, testículos inteiros de cada espécie, em repouso e em maturação, foram fixados em líquido de Bouin por 6-8 horas e submetidos às técnicas histológicas de rotina. Utilizaram-se técnicas histoquímicas clássicas para detectar carboidratos e proteínas. Os testículos das espécies estudadas são órgãos pares e franjados. Histologicamente, as franjas da região cranial de todas as espécies são espermatogênicas, com células da linhagem em mesma fase de desenvolvimento contidas em cistos. As franjas da região caudal de *C. conirostris* são também espermatogênicas, enquanto *P. maculatus* e *I. labrosus* apresentam franjas caudais exclusivamente secretoras. Histoquimicamente detectou-se na secreção das franjas caudais de *P. maculatus* glicoproteínas neutras, glicoconjugados ácidos carboxilados, incluindo sialomucinas e glicoconjugados ácidos e sulfatados e em *I. labrosus* glicoproteínas neutras. Houve diferenças significativas entre o comprimento das franjas das regiões cranial e caudal para todas as espécies estudadas. **PALAVRAS CHAVE.** Glândula testicular, secreção, Siluriformes, testículos franjados, vesícula seminal.

The Siluriformes, among the Ostariophysi, are the most diverse fish order: 34 families, 412 genera and over 2400 species with a wide geographic distribution (NELSON 1994, HELFMAN *et al.* 2000). The family Pimelodidae is considered one of the largest and most diverse in fresh waters, presenting over 300 species (BURGESS 1989).

The morphology of the reproductive system of male Siluriformes has been shown to be variable: in some families the testes show spermatogenic lineage cells throughout their

extension; others present caudal region with seminal vesicles or accessory structures that do not show spermatogenic activity, however, they can store spermatozoa (LEGENDRE *et al.* 1996). Moreover, the caudal region of some Siluriformes' testes is known to present secretory activity without storing spermatozoa (LOIR *et al.* 1989). Recent studies have revealed that the testes of *Iheringichthys labrosus* (Lütken, 1874) are fringed, with a peculiar morphofunctional organization containing a spermatogenic cranial region and a secretory caudal region (SANTOS

et al. 2001). Fringed testes with asynchronous spermatogenic lineage cells have been found in some mature pimelodids (BAZZOLI *et al.* 1997).

It has been suggested that, knowledge of anatomical differences of the reproductive system of teleosts may offer evidence for establishing phylogenetic relationships between similar species (MEINSNER *et al.* 2000). Thus, MATTEI (1991) suggested the use of spermatozoa morphology to help studies on taxonomy and phylogeny of fish.

In consideration of the fact that few studies have investigated Siluriformes' testes, especially those of the family Pimelodidae, which is an economically important group, the present study investigated anatomical, biometrical, histological and histochemical differences in the testes of three pimelodids species: with the aim of establishing testicular patterns, that may provide important information to phylogenetic studies.

MATERIAL AND METHODS

Animals

Twenty-five males of *Pimelodus maculatus* (Lacepède, 1803), *Iheringichthys labrosus* (Lütken, 1874) and *Conorhynchos conirostris* (Valenciennes, 1840) were used. For each species five males were in the resting stage and 20 in reproductive activity. Fishes were captured, using gill nets with 3-12 cm mesh size, from November 2001 to October 2002 in the reservoirs of Furnas and Três Marias and also in the São Francisco river, Minas Gerais, Brazil.

Anatomy and biometry

For each species 15 males in reproductive activity were fixed in 10% formaldehyde for 24 hours and stored in 70% alcohol. From each specimen we measured the distance from the rostral edge of the head to the base of the caudal fin: standard length (mm). Testes were removed, measured (longitudinal length) and maintained in 70% ethanol. Maturing and resting testes were each divided into a cranial and caudal region. The fringes were manually counted and their lengths, in each of two regions of maturing testes, were determined using a caliper.

Statistical analyses

Significant differences in the length of the fringes between the cranial and caudal regions of the testes were determined using Student's *t*-test, with a statistical significance level of $p \leq 0.05$.

Light microscopy

For each species fragments of cranial and caudal regions of five testes in the resting stage and five in reproductive activity, were fixed in Bouin's fluid for 6-8 hours, embedded in paraffin, sectioned at 3-5 μm , and stained with hematoxylin and eosin or Gomori trichrome. To detect carbohydrates and proteins on the testes caudal region, the following histochemical techniques were used: PAS (Periodic Acid-Shiff); Alcian blue pH 0.5; Alcian blue pH 2.5; and ninhydrin-Shiff (PEARSE 1985).

RESULTS

Biometric results of the fishes, and mature testes in this study are showed in table I. A Student's *t*-test showed that the cranial region was significantly longer than the caudal region for all species studied ($p \leq 0.05$). Fringed mature testes and their respective histological section of cranial and caudal regions are presented in figures 1 to 9.

The testes of all species were paired organs related to the swim bladder cranially and to the kidneys caudally. They showed fringes or lobes that communicated singly, in pairs or in groups of three, through the whole testes (Figs 1, 4 and 7). Mature testes showed prominent fringes in two distinct regions: cranial with approximately 3/4 of the testes total length and caudal with 1/4. Conversely, testes in the resting stage presented smaller fringes in both regions. Each testis had a spermatid duct that joined at their caudal portion, forming the common spermatid duct, which extended to the urogenital papillae.

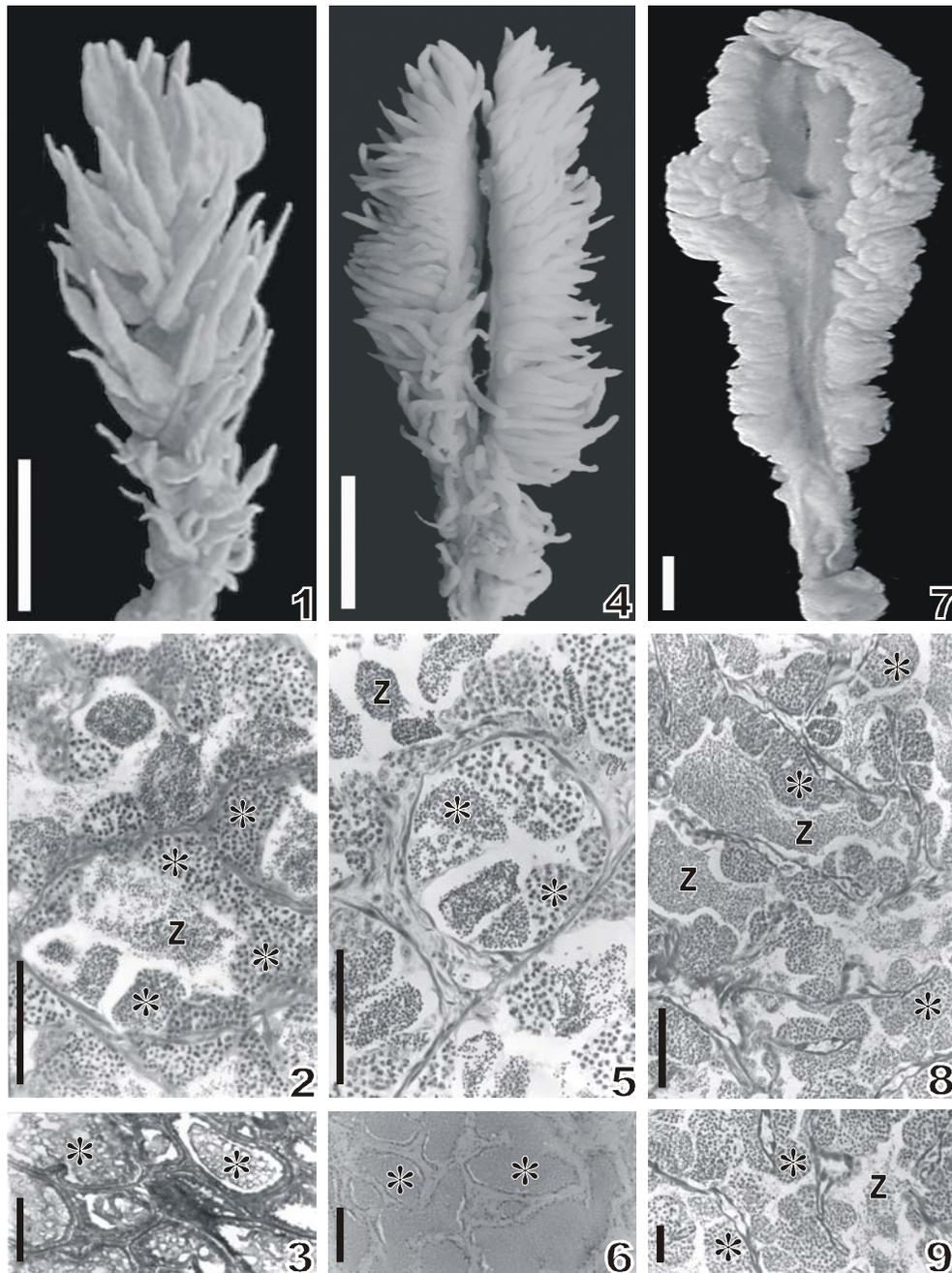
Cranial fringes of mature testes of *P. maculatus* (Fig. 2) and *I. labrosus* (Fig. 5) showed seminiferous tubules with an open lumen, filled with spermatozoa, and the wall of the seminiferous tubules was composed of cysts, each of which contained spermatogenic cells in the same stage of development. The caudal fringes showed tubules with prismatic secretory cells and an open lumen filled with secretion (Figs 3 and 6). Cranial fringes of testes in resting stage showed on the wall of the seminiferous tubules cysts only spermatogonia cells and an occluded lumen. However, the tubules on the caudal fringes did not show any secretion, even though they had an open lumen.

In *C. conirostris* cranial and caudal fringes of mature testes showed exclusively cysts with spermatogenic lineage cells on the wall of the seminiferous tubules, which presented an open lumen with spermatozoa (Figs 8 and 9). Conversely, fringes of testes in resting stage showed only spermatogonia cysts in the wall of the tubules with an occluded lumen.

Histochemical analyses revealed that the secretion inside the tubules on caudal fringes of *P. maculatus* contained neutral glycoproteins, acid glycoconjugates, acid carboxylates, including sialomucines and acid and sulfates glycoconjugates. The secretion on caudal fringes of *I. labrosus* reacted positively to the histochemical techniques, which indicated the presence of neutral glycoproteins.

DISCUSSION

In the present study all species of pimelodids showed paired and fringed testes, joined caudally. These anatomic characteristics are common to other Siluriformes, such as: *Ictalurus punctatus* (Rafinesque, 1818) (SNEED & CLEMENS 1963); *Heteropneustes fossilis* (Bloch, 1794), *Clarias batrachus* (Linnaeus, 1758) (SIRCAR 1970); *Pimelodella cristata* (Müller & Troschel, 1848), *Rhamdia quelen* (Quoy & Gaimard, 1824), *Ageneiosus brevifilis* (Linnaeus, 1766) (LOIR *et al.* 1989); *P. maculatus* (BAZZOLI *et al.* 1997); *Trachelyopterus galeatus* (Linnaeus, 1766) (MEISNER *et al.* 2000).



Figures 1-9. Fringed maturing testes and their respective histological section of cranial and caudal regions: hematoxylin and eosin (2, 3, 6, 8, 9) and Gomori trichrome (5). (1) Fringed maturing testes of *P. maculatus*: (2) spermatogenic activity in seminiferous tubules of cranial region, with cysts of spermatogenic cells in different stages of development (*) and spermatozoa (Z); (3) secretory activity in tubules of caudal region of testes with accumulation of secretion in lumen (*); (4) fringed maturing testes of *I. labrosus*: (5) spermatogenic activity in seminiferous tubules of cranial region, with cysts of spermatogenic cells in different stages of development (*) and spermatozoa (Z); (6) secretory activity in tubules of caudal region of testes with accumulation of secretion in lumen (*); (7) fringed maturing testes of *C. conirostris*: (8-9) spermatogenic activity in seminiferous tubules of cranial and caudal region, respectively, with cysts of spermatogenic cells in different stages of development (*) and spermatozoa (Z). Scale bars: figures 1, 4, 7 = 1 cm; figures 2, 3, 5, 6, 8, 9 = 100 μ m.

Table I. Biometry of fish and mature testes. (SD) Standard deviation.

Species	Specimens' number (n)	Standard length (mm)	Testes length (mm)	Number of fringes	Cranial fringe lengths (mm)	Caudal fringe lengths (mm)
		Mean \pm SD	Mean \pm SD	Minimum – Maximum	Mean \pm SD	Mean \pm SD
<i>P. maculatus</i>	15	132.3 \pm 43.4	34.6 \pm 13.9	73 – 132	6.88 \pm 1.96	2.10 \pm 1.00
<i>I. labrosus</i>	15	126.0 \pm 9.5	33.7 \pm 4.1	178 – 204	4.32 \pm 1.02	1.60 \pm 0.45
<i>C. conirostris</i>	15	465.0 \pm 21.2	108.1 \pm 15.0	1466 – 1686	4.45 \pm 1.06	2.16 \pm 0.49

Testes of *Pimelodella cristata*, *Pimelodus ornatus* Kner, 1858, *Pimelodus blochii* Valenciennes, 1840, *Pseudopimelodus raninus* (Valenciennes, 1840) and *R. quelen* present variable fringe numbers (LOIR *et al.* 1989). In the present study three species presented testes with different fringe numbers, the highest being found in *C. conirostris* (1466 to 1686) and the lowest in *P. maculatus* (73 to 132). In relation to cranial and caudal fringe lengths, there were significant differences among the three species: *P. maculatus* presented the largest cranial (6.88 \pm 1.96 mm) and *C. conirostris* the largest caudal (2.16 \pm 0.49 mm) fringes.

Testes of *P. maculatus* and *I. labrosus* showed a spermatogenic cranial region and exclusively secretory caudal region, whereas testes of *C. conirostris* showed spermatogenic activity in both regions. Spermatogenesis occurred synchronously inside cysts throughout the whole seminiferous tubules, as is known for most teleosts (GRIER 1981). In the present study, secretory activity occurred in cells of the seminiferous tubules wall, on caudal fringes without forming conspicuous glands.

Conversely, the Siluriformes *H. fossilis*, *C. batrachus*, *Trachelyopterus lucenai* Bertolotti, da Silva & Pereira, 1995 and *T. galeatus* have been found to have a secretory caudal region and form seminal vesicles (SIRCAR 1970, MEISNER *et al.* 2000). A similar structure was also detected in species of the family Gobiidae (SIRCAR & HAR 1980, LAHNSTEINER *et al.* 1992).

Secretory activity in the caudal region of *P. maculatus* and *I. labrosus* testes was observed only during reproductive period, while in the resting stage no secretion was found inside the seminiferous tubules nor in the testicular ducts. Seasonal variations in weight and secretory activity of these vesicles, in relation to testes cycle have been described for other Siluriformes: *Mistus tengara* (Hamilton, 1822) (RASTOGI 1969), *H. fossilis* (NAYYAR & SUNDARARAJ 1970) and *C. batrachus* (SINGH & JOY 1999). However, WEISEL (1949) did not observe any seasonal changes in length nor in the structure of seminal vesicles epithelial cells of *Gillichthys mirabilis* Cooper, 1864.

Various substances and functions are related to the secretion produced by the seminal vesicles of teleosts: VAN DER HURK *et al.* (1987), LAHNSTEINER *et al.* (1990), LAHNSTEINER *et al.* (1992) mentioned that they can store spermatozoa, secrete glycoproteins, steroids and pheromones, increase semen volume and can also influence fertilization, and female attraction. The secretion observed in the present study may have a similar

function to that of the seminal vesicles of other teleosts.

The information available in the literature relating to the origin and nomenclature of fish seminal vesicles are speculative. Therefore, FISHELSON *et al.* (1994) demonstrated, through ontogenesis of seminal vesicles of *Clarias gariepinus* (Burchell, 1822) that they are formed in the beginning of spermatogenesis, as a simple protrusion on the wall of the spermatid duct and become complex with age. SUNDARARAJ (1958), LEHRI (1967), RASTOGI (1969), NAYYAR & SUNDARARAJ (1970), VAN DER HURK *et al.* (1987), ASAHINA *et al.* (1989) and LAHNSTEINER *et al.* (1990) affirmed to the presence of the seminal vesicle in some teleosts, notably in catfishes, gobiids and blennies. For the species studied in the present work, naming testes caudal fringes, as seminal vesicle is inappropriate, as according to their embryonic and ontogenetic development they are not homologous to mammal's seminal vesicle. Therefore the term a testicular gland mentioned by RICHTARSKI & PATZNER (2000) is deemed more suitable.

As herein demonstrated, the testes characteristics of the studied pimelodids presented an unusual pattern in relation to most teleosts. They showed fringes through the whole testes; with significant differences between fringe lengths of cranial and caudal regions; *P. maculatus* and *I. labrosus* presented two regions with distinct function: spermatogenic cranial region and secretory caudal region which did not constitute an individual gland, and secretory activity only occurred during the gonadal maturation period. Testes of *C. conirostris* presented cranial and caudal fringes with spermatogenic activity only. These quantitative, biometric and morphological characteristics may provide useful data for phylogenetic analyses of Siluriformes.

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