

EDITORIAL

What is the role of microbial infection in Alzheimer's disease?

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In 1907, Alois Alzheimer, a German psychiatrist and neuropathologist, described the case of Auguste Deter, a 51-year-old woman who was closely monitored for 5 years for symptoms of progressive presenile dementia. After her death, necropsy revealed several abnormalities in her brain, including cortical atrophy, arteriosclerotic changes in vascular tissues, and the abnormal deposition of "special substances" inside and outside of neurons, now identified as hyperphosphorylated tau tangles and amyloid- β (A β) plaques, respectively.¹ Presently, Alzheimer's disease (AD) is considered the leading cause of dementia worldwide and is rapidly becoming one of the most costly, deadly, and burdensome diseases.² Unfortunately, more than 100 years after its first description, we do not entirely understand how AD is initiated and what factors modulate its progression. At present, we know that sporadic AD (accounting for over 95% of all AD cases) can be linked with unmodifiable and modifiable risk factors. Aging is the most notorious risk factor for AD. Female sex and certain ethnicities have also been associated with a higher risk of developing this type of dementia. More recently, several pathological conditions such as type 2 diabetes, hypertension, and others have been linked with AD at physiological and molecular levels. Among them, an emerging risk factor for AD includes microbial infections.³ Basic and clinical data support the role of microbial infection in AD. Some of this evidence is discussed below.

One example found in the literature includes a multiscale analysis of three distinct patient cohorts demonstrating that human herpesvirus (HHV)-6A and HHV-7 disrupt molecular, genetic, and clinical networks more severely in AD patients than in non-demented individuals.⁴ Another study showed that individuals recently diagnosed with herpes simplex virus (HSV, n=8,362) had a risk ratio of 2.564 to develop dementia compared to non-HSV controls (n=25,086). Notably, this risk ratio decreased to 0.092 when the HSV-infected patients received anti-herpetic treatment,⁵ suggesting

that the removal of pathogens results in an improved prognosis.

In the preclinical field, growing evidence suggests that infections might be associated with the accumulation of amyloid plaques in the brain, one of the hallmarks and perhaps the triggering event in AD. One example includes experiments involving transgenic *Caenorhabditis elegans* expressing the human A β 1-42 residue. These animals, developing A β deposits over time, displayed reduced mortality compared to their non-transgenic counterparts when infected with *Candida albicans*.⁶ Additional support comes from experiments involving the intracerebral injection of *Salmonella* Typhimurium in transgenic 5XFAD mice, an animal model of accelerated amyloid pathology. Treated mice exhibited increased brain amyloid pathology compared to sham-treated mice. Moreover, A β deposits in *Salmonella*-treated 5XFAD co-localized with the invading bacteria, suggesting a direct interaction between both pathogenic entities (amyloid plaques and bacteria). Interestingly, the mortality rate induced by *Salmonella* sp. administration was reduced in 5XFAD mice, consistent with previous *in vitro* and *in vivo* evidence suggesting an antimicrobial role for A β aggregates.⁶ Corroborating the involvement of infection in AD pathogenesis, we have demonstrated that pneumococcal meningitis increased the expression of the receptor for advanced glycation endproducts (RAGE), a known contributor for A β production by enhancing the activity of secretases and neuroinflammation. We found that infection with *Streptococcus pneumoniae* also enhanced the generation of A β 1-42, an A β fragment associated with increased fibrillogenesis, microglial cell activation, and memory impairment.⁷

Similar results were observed in models of peripheral infection triggered by polymicrobial sepsis using cecal ligation and puncture (CLP) surgery, the gold-standard model of polymicrobial sepsis. These animals displayed an increase in A β production and higher Ser-202-phosphorylated Tau (p-TauSer-202, another indicator

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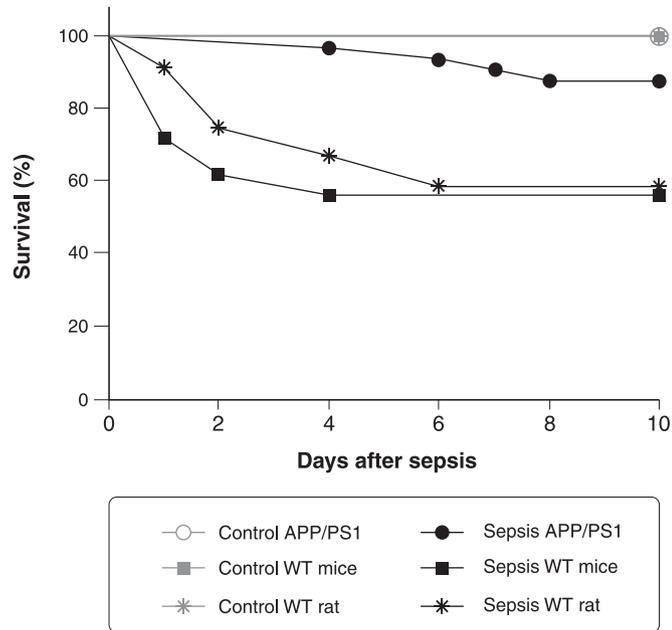


Figure 1 Kaplan-Meier survival curve of APP/PS1 mice, WT C57BL/6 mice, and Wistar rats subjected to experimental polymicrobial sepsis triggered by CLP. Control groups subjected to sham surgery were included for comparison. The animals were followed for 10 days and mortality was recorded (APP/PS1, sepsis $n=32$ vs. control $n=16$, $p = 0.153$; C57BL/6 mice, sepsis $n=50$ vs. control $n=50$, $p < 0.001$; Wistar rats, sepsis $n=12$ vs. control $n=10$, $p = 0.02$). The APP/PS1 mice subjected to sepsis had an 87.88% survival rate, WT mice had a survival rate of 56%, and Wistar rats had a survival rate of 58.33%. APP/PS1 = amyloid- β precursor protein/presenilin 1; CLP = cecal ligation and puncture.

of AD pathology) and RAGE followed by cognitive impairment.⁸ In another study, APP/PS1 double transgenic mice that express human amyloid- β precursor protein (APP) and a mutant human presenilin (PS1) associated with early-onset AD, were subjected to CLP and their survival was measured over time. Usually, CLP triggers sepsis of moderate severity, with mortality rate in rodents of about 40%.^{8,9} Some crucial variables that influence the mortality rate of the animals include fluid resuscitation procedures, CLP conditions, the position of the ligation, the postoperative antibiotic regimen, and parenteral nutrition.⁹ Surprisingly, we found that APP/PS1 mice subjected to polymicrobial sepsis had a mortality rate of only 12.12%, while the mortality of CLP in wild-type mice was 44% (Figure 1). This was also observed in a rat model of sepsis, where wild-type Wistar rats had a mortality rate of 41.6% compared with the respective controls. In summary, our data show that wild-type C57BL/6 mice and Wistar rats had a threefold increase in mortality compared to APP/PS1 mice. Our results are in agreement with those found in the literature, demonstrating that A β provides protection during infection.

Neuroinflammation is a central component of AD pathogenesis; it contributes to disease development and neurodegeneration.^{2,3} Infection or inflammatory conditions derived from aging, as well as stroke, have been identified as AD risk factors,³ triggers of pro-inflammatory cytokine production in the brain, and drivers of neuroinflammation. Recently, an essential mechanism of cell signaling that associates neuroinflammation to A β production was identified. In the brain, insults or infection lead to production of pro-inflammatory cytokines, which in turn promote the formation of interferon-induced transmembrane protein 3 (IFITM3) in neurons and astrocytes. IFITM3 binds to γ -secretase, increasing its activity, and upregulates A β production.¹⁰ Figure 2 provides a summary of the molecular cascade associated with this pathway.

Overall, several mechanisms link microbial infections and AD, including but not limited to A β misfolding and neuroinflammation. Additional research will help us clarify the specific molecular links between pathological events, explore the specificity of certain microbes with pathological variations in AD, and identify new preventive and therapeutic strategies to combat this fatal form of dementia.

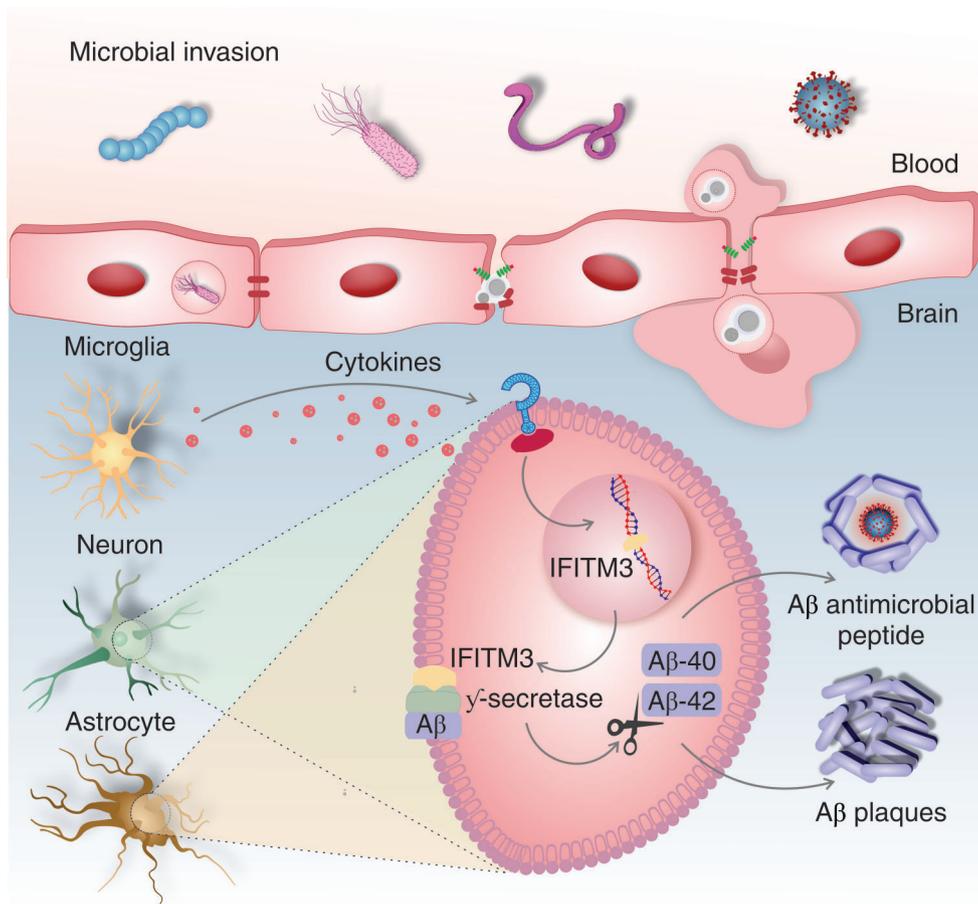


Figure 2 Infection-induced inflammation as a potential trigger of Alzheimer’s disease pathology. Infection or inflammation activates microglial cells, triggering the release of pro-inflammatory cytokines. In sequence, the pro-inflammatory cytokines promote the formation of interferon-induced transmembrane protein 3 (IFITM3) in neurons and astrocytes, which binds to γ -secretase, increasing its activity and upregulating A β production. Released A β peptides have antimicrobial effects, preventing pathogen development by forming amyloid plaques. Brain A β deposition exacerbates the cerebral inflammatory response, accelerating AD pathological cascades. A β = amyloid- β ; AD = Alzheimer’s disease.

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Disclosure

The authors report no conflicts of interest.

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