Revista Brasileira de Farmacognosia Brazilian Journal of Pharmacognosy 23(1): 44-50, Jan./Feb. 2013

Article

Received 20 Mar 2012 Accepted 11 Jul 2012 Available online 11 Oct 2012

Keywords: plasma antioxidants polyphenols *Ugni molinae* populations

ISSN 0102-695X DOI: 10.1590/S0102-695X2012005000122

#### Introduction

Ugni molinae Turcz. (Bull. Soc. Imp. Naturalistes Moscou 21: 579. 1848), Myrtaceae, is a native Chilean plant commonly known as murtilla, murta or Chilean guava. It is distributed from the Maule Region to Chiloé Island, including the Juan Fernández archipelago (Hoffmann, 1991). The leaves of this plant have been infused and used as an astringent for treating diarrhea and dysentery by the continent indigenous people. Several authors have determined the chemical composition and biological activity of continental U. molinae leaves (Avello, 2000; 2004; Avello & Pastene, 2005; Aguirre et al., 2006; Rubilar et al., 2006; Suwalsky et al., 2006; 2007; Avello et al., 2009; Rubilar et al., 2011). The presence of phenol-type substances such as catechin, epicatechin, myricetin, quercetin, kaempferol, and some of their glycosylated derivatives have also been described (Avello & Pastene, 2005; Rubilar et al., 2006; 2011). Moreover, studies intended to verify the benefits of the consumption of this plant for human

### Variation in phenolic compounds of *Ugni molinae* populations and their potential use as antioxidant supplement

Marcia A. Avello,<sup>\*,1</sup> Edgar R. Pastene,<sup>1</sup> Evelyn D. Bustos,<sup>2</sup> Magalis L. Bittner,<sup>2</sup> José A. Becerra<sup>2</sup>

<sup>1</sup>Faculty of Pharmacy, University of Concepción, Chile; <sup>2</sup>Faculty of Natural Sciences, University of Concepcion, Chile.

Abstract: In the present work we carried out a comparative study of total phenolic contents and antioxidant capacity of aqueous leaf extracts of Ugni molinae Turcz., Myrtaceae (infusion and Soxhlet extracted) prepared from continent and Juan Fernández Island samples. The results revealed that total phenol content (TPC), tannins (TTC) and flavonoids (TFC) for U. molinae extracts (infusion and Soxhlet extracts) from island leaves were 38.5, 56.7 and 37.5% higher than those obtained with leaves from the continent, respectively. Also, HPLC profiles showed important differences between U. molinae populations. In vitro antioxidant capacity (scavenging of DPPH radical) for 1% infusion and aqueous extract (Soxhlet method) of U. molinae from island samples, was 15% greater than from continent samples. Further, in vivo impact of U. molinae intake (1% infusion) was studied in plasma samples obtained from healthy volunteers. Participants that consumed tea prepared with leaves from island population showed higher TBARS reduction and plasma antioxidant capacity (TEAC-CUPRAC) than those who consumed tea prepared with leaves from continental population. The conditions of the territory in which U. molinae populations growth could explain the differences in their composition and activity. According to results, island U. molinae populations could be an important source of study for the development of an antioxidant supplement, and thereby contribute to the use of this species that has becoming an ecological problem in the island.

> health have shown that polar extracts of continental U. molinae stabilize free radicals generated in different systems (Avello, 2000; 2004). In our previous work, we established that plasma antioxidant capacity was increased significantly after an acute ingest of infusions prepared with continental U. molinae leaves sampled from a single location (VIII Region, Bío-Bío), according to traditional medicine practice (Avello & Pastene, 2005). The Juan Fernández archipelago is a national park located 670 km from the continent. It was declared a Biosphere Reserve by UNESCO. U. molinae was introduced to the archipelago (possibly by birds) from the continent. U. molinae behave like invasive species, occupying and clearly expanding into the habitat of the island's native species, competing heavily with and displacing the endemic species (Skottsberg, 1953; Ricci, 1989; Stuessy et al., 1998; Greimler et al., 2002; Cuevas et al., 2004). An option for controlling invasive species it might be their requirement for the food and pharmaceutical industry or for biotechnology development. Geo climatic factors are known to alter

the synthesis of secondary metabolites. Relevant climate factors include solar radiation, temperature, water, and soil (Davies & Schwinn, 2006). Therefore, the conditions of the territory in which populations of U. molinae develop could affect their chemical composition. Given the abovementioned, quantitative differences in total phenolic contents and bio-activities are expected to occur between populations of U. molinae. Hence, the first aim of this work was to compare total phenolic contents of infusions and aqueous extracts prepared with Ugni leaves from the continent and Juan Fernández Island. In second place, we plan to evaluate the impact of both U. molinae populations on the plasma antioxidant capacity of healthy volunteers before and after they drink infusions prepared in the way they normally ingest their herbal native infusions.

#### **Materials and Methods**

#### Vegetable matter

The biological material was collected from Bio-Bío Region, (36°00' and 38°30' S) and from Juan Fernández archipelago (Robinson Crusoe Island, 33°36' and 33°46' S), Chile. The leaves of the plants were collected in november-december 2008, when the plant was flowering. The species was identified by the taxonomist, Dr. Roberto Rodríguez, of the Department of Botany, Faculty of Natural and Oceanographic Sciences, Universidad de Concepción (CONC 146511 y 116887, respectively).

#### Obtaining aqueous extracts

Dried leaves from both *Ugni molinae* Turcz., Myrtaceae, populations were washed, air-dried, and ground to a fine powder. Powdered *U. molinae* leaves (50 g) were extracted in a Soxhlet apparatus with water until exhausting the vegetable matter. The mass: solvent ratio was 1:6. The extracts obtained from the samples of both populations were freeze-dried and stored in a dry place, protected from the light, until their use.

#### Obtaining infusions

Infusions were prepared with 1 g of dried, ground leaves from each population. For this, 100 mL of water (100  $^{\circ}$ C) were added to the leaves, which were left to steep for 5 min. The samples to be used for qualitative and quantitative analyses were freeze-dried.

#### Total polyphenol contents

Total polyphenol contents (TPC) were

determined spectrophotometrically according to Velioglu et al. (1998), using the Folin-Ciocalteu reagent (Sigma, MO, USA). Briefly, aliquots (0.5 mL) of test samples were mixed with 25 mL water, 2.5 mL Folin-Ciocalteu reagent (Merck, Germany), and 10 mL 20% Na<sub>2</sub>CO<sub>3</sub>, and then completed to 50 mL with water. The mixtures were shaken for 30 min, then allowed to stand for 30 min. Absorbance was registered at 765 nm using gallic acid as a standard. Total tannin (TTC) and flavonoid (TFC) contents were determined, respectively, according to Lastra et al. (2000) and Salamanca et al. (2007).

#### HPLC analysis

Extracts (3 mg/mL) were separated by RP-HPLC using a Lachrom instrument, equipped with a  $250 \times 4.6$  mm, 5 µm, Kromasil KR100-5C18 column (Eka Chemicals AB, Bohus, Sweden). A gradient elution was performed by varying the proportion of solvent A (double distilled water containing 0.1% TFA, v/v) to solvent B (acetonitrile containing 0.1% TFA) with a flow of 1 mL/min. The following gradient was used: 0-25 min, 10-25% of B in A; 25-30 min, 25-75% of B in A and then bring mobile phase composition back to the initial condition in 5 min to the next run. For flavonoids, detection was at 350 nm using a diode array detector. For hydrolysable tannins (gallic acid derivates) and catechins, detection was done at 280 nm. When reference substances were available some compounds were grouped and tentatively identified by matching their retention times  $(t_p)$  and online UV spectra. Although the other compounds (particularly some rare myricetin and quercetin-3-O-glycosides) could not be wholly identified, they were characterized according to their class on the basis of their UV-VIS spectra and data reported previously in literature (Rubilar et al., 2006). The reagents used for analysis were all HPLC grade (Merck, Germany). Peak assignment was done by comparing the tR with those of pure standard substances, all purchased from Sigma (gallic acid, catechin, epicatechin, isoquercitrin, isorhamnetin, quercetin, myricetin, myricitrin, kaempferol and ellagic acid).

## Antioxidant capacity in vitro: stabilization of the 1, 1-diphenil-2-picryl-hydrazyl (DPPH) radical

The DPPH radical was stabilized as described by Joyeux et al. (1995) for both extracts and the infusions made from the two *U. molinae* populations. The results were expressed as a percentage of the discoloration of the radical. Gallic acid (Merck, Germany) was used as a standard.

#### Administration of infusions

The infusions made with the two *U. molinae* populations were administered twice daily (11 am and 17 pm) for three days, simulating the dosage used in popular medicine (1%). This study was carried out with the approval of the Ethics Committee of University of Concepción (N°VRID 290/2012), and the volunteers were recruited with informed consent, and following guidelines of the Declaration of Helsinki and Tokyo for humans.

#### Plasma preparation

Blood was obtained from healthy volunteers (n=24, 20-30 years of age, non-smokers, normal range of body mass index, normolipemic, non-diabetic, following a normal diet) through venipuncture before and after drinking the teas (1%) made with leaves from continental (n=12) and island (n=12) populations of *U. molinae*. EDTA (2.7 mM) was used as an anticoagulant in the samples. The plasma was separated after centrifuging at 800 x g at 4 °C for 15 min according to Avello & Pastene (2005).

#### Conjugated dienes

The formation of conjugated dienes was determined by UV absorption at 234 nm (Esterbauer et al., 1989) using a spectrophotometer (Shimadzu UV-VIS 1601).

#### Thiobarbituric acid reactive substances (TBARS)

Plasma samples (500  $\mu$ L) were mixed with 1 mL of the TBARS reagent and the mixture was incubated at 100 °C for 30 min. After cooling on ice and centrifuging at 1800 g for 15 min (Kubota, Japan), the absorbance of the supernatants was measured at 532 nm (Jasco, Japan). The results were expressed in terms of percentage of protection (Gugliucci, 1996).

# Trolox equivalent antioxidant capacity-Cupric ion reducing antioxidant capacity (TEAC-CUPRAC assay).

TEAC-CUPRAC indexes were determined according to Apak et al. (2007). For this, a CuCl<sub>2</sub> solution (1.0 x  $10^{-2}$  M) was prepared by dissolving 0.4262 g CuCl<sub>2</sub> x 2H<sub>2</sub>O in water and diluting to 250 mL. Ammonium acetate buffer (1.0 M, pH 7.0) was prepared by dissolving 19.27 g NH<sub>4</sub>Ac in water and diluting to 250 mL. Neocuproine (Nc) solution (7.5 x  $10^{-3}$  M) was prepared daily by dissolving 0.039 g Nc in 96% ethanol and diluting to 25 mL with ethanol. Trolox

 $(1.0 \times 10^{-3} \text{ M})$  was prepared in 96% ethanol. The assay was performed by adding 1 mL  $10^{-2}$  M Cu<sup>2+</sup>+1 mL 7.5 x  $10^{-3}$  M neocuproine+1 mL 1M NH<sub>4</sub>Ac. The antioxidant sample (or standard) solution (x mL) and H<sub>2</sub>O (1-X mL) were added to the initial mixture in order to make a final volume of 4.1 mL. The final absorbance was measured at 450 nm.

#### Statistical analysis

Statistical analyses were performed using one-way analysis of variance (ANOVA) followed by Dunnett's test for multiple comparisons. Differences were considered significant at p < 0.01.

#### **Results and discussion**

The total phenol contents of the aqueous extracts (Soxhlet method) differed significantly. So, the island population of Ugnimolinae Turcz., Myrtaceae, contained 38.5% more total phenols (TPC) than continental population (Figure 1A). Likewise, compared with the continental population, aqueous extracts from the island population showed 56.7% and 37.5% more total tannins (TTC) and flavonoids (TFC), respectively (Figure 1B). In vitro antioxidant capacity (scavenging of DPPH radical) for 1% infusion and aqueous extract (Soxhlet method) of U. molinae from island samples, was 15% greater than from continent samples (Figure 1C). These differences were clearly illustrated in HPLC profiles depicted in Figures 2A and B. In the extracts from the continental population, HPLC analysis suggested the presence of phenolic acids, flavan-3-ols and flavonols derivates (Figure 2A). However, its identity could not be assigned in all cases because certain gallotannins and myricetin and quercetin derivates currently were not available in our laboratory. Nevertheless, it was possible to tentatively assign the identity of gallic acid  $(t_p=4.7 \text{ min})$ , catechin  $(t_p=9.8)$ , epicatechin  $(t_p=12.5)$ , myricetin 3-O-rhamnoside (myricitrin;  $t_p=19.2$ ), quercetin-3-O-glucoside (isoquercitrin;  $t_p=22.0$ ) and ellagic acid ( $t_p=31.8$ ). In the aqueous extract of the island population, we could clearly identify some of the polyphenols described previously by Rubilar et al. (2006; 2011) such as gallic acid derivates (hydrolysable tannins), flavan-3-ols (epicatechin) and glycosides of myricetin, quercetin (Figure 2B). These substances are widely known for their antioxidant, antinociceptive, antiallergic and antitumoral activities (Yokoshimo & Moriwaki, 2005; Meotti et al., 2006; Shimosaki et al., 2011). Altogether, HPLC registers suggest that insular population have similar flavonoids and tannins than continental population but its levels, particularly flavonoids, are highest. These results are in accordance with the observed difference in the total

phenolic, tannins and flavonoids contents (Figure 1A). It should be noted that the study carried out by Rubilar et al. (2006) was done using a continental population collected in the Araucanía Region (IX). In the present work, our continental sample was extracted in Bío-Bío region (VIII). In both cases, there is more than 800 km between insular and continental sampling zones. Geologically, Juan Fernández is an archipelago of volcanic origins, and it has a warm, temperate, subtropical, marine climate with high environmental humidity. This climate creates special conditions in the sector and allows the development of exuberant, unique vegetation. In turn, the Bío-Bío Region is a transition zone between the temperate climates of central Chile and the rainy climates found southern of the "rio Laja". The predominant climate in this region is temperate Mediterranean and, in general, the soils are classified as miscellaneous (www.meteochile.cl). The high humidity and volcanic soils of the Juan Fernández archipelago could influence the biosynthesis of phenolic compounds. Correlations have been reported between humidity conditions of a territory and the levels of phenolic compounds. On the other hand, a highly fertile environment with elevated iron hydroxide and aluminum hydroxide contents (Ritter et al., 2003). Although contributions of nitrogen, phosphorous, potassium, and sulfur are described as important, their deficiency in volcanic soils could stimulate the biosynthesis of phenolic compounds. These mechanisms could be associated with the activity of key enzymes in phenolic biosynthesis such as phenylalanine ammonia-lyase (PAL) (Davies & Schwinn, 2006). The mechanisms used by plants to adapt to stressful situations (e.g., an introduced species in an unknown environment) are also important; the resulting defense mechanisms have a high participation of phenolic compounds, as do the allelopathic mechanisms used to protect against pathogens and predators (Waniska, 2000; Vivanco et al., 2005; Vermerris & Nicholson, 2006).

In order to evaluate if the acute oral intake of teas prepared from two populations of U. molinae could modify antioxidant status in humans we design a pilotscale study. In this trial, plasma antioxidant capacity of 24 healthy volunteers was measured before and after the administration of infusions (1%) of U. molinae leaves from the continent and island through the formation of conjugated dienes, TBARS, and TEAC-CUPRAC. None of the volunteers reported adverse effects. TBARS formation was reduced in a 31.22% for those participants who drank tea from the island population, as opposed to 19.33% for the group that drank the tea made using the continental population (Figure 3A). Although our measurements of conjugated dienes did not reveal a general overview of the influence of the assayed infusions on plasma antioxidant protection at this level, we did note a slight protection



**Figure 1.** Total polyphenol contents (TPC), Total tannin contents (TTC), and Total flavonoid contents (TFC) of aqueous extracts (A) and infusions (B) from island and continental populations of *Ugni molinae*. (C) DPPH scavenging capacity of 1 % infusions and aqueous extracts from island and continental populations of *U. molinae*. Values are the mean±SD; n=3.\*p<0.01.

(2.66%) associated with the ingestion of the teas made from the island population (Figure 3A). Fluctuations in the plasma Trolox equivalents (TEAC-CUPRAC), could be ascribed both to an increased ingest of antioxidants (from natural or synthetic sources) or to pathological states that promote depletion of endogenous antioxidants (Apak et al., 2007). The results of our study reveal that TEAC-CUPRAC indexes were improved after oral intake of U. molinae teas from both populations. Interestingly, plasma samples of participants who consumed tea prepared with island plants has 316.8 µM more Trolox equivalents than those samples analyzed in the continent group (p < 0.05) (Figure 3B). Such results suggest that positive correlation between polyphenol contents and the in vivo antioxidant activity might be associated to U. molinae intake. The phenolic compounds described for this species predominantly are flavonoid glycosides and high molecular weight tannins whose bioavailability remains uncertain. However, important evidence suggests that these compounds could be substrate for intestinal microbiota (Selma et al., 2009). Therefore, unbound polyphenols (genins) or even new molecules formed in gastrointestinal tract might be responsible of the acute increments in the antioxidant status of human plasma after intake of polyphenol-rich beverages and foods (Yokoshimo & Moriwaki, 2005; Landete, 2011). These associations are merely speculative and need to be duly clarified trough pharmacokinetic studies. Encouraged for the results of this work, we plan to lead our efforts to the study of chemical profiles for phenolic compounds and its metabolites in the blood of humans after U. moline tea intake.



**Figure 2.** HPLC analysis of *Ugni molinae* aqueous extracts. A. Chromatogram of the aqueous extract of continental *U. molinae*; B. Chromatogram of the aqueous extract of island *U. molinae*.

#### Conclusions

Our results reveal that total phenolic contents and *in vitro* antioxidant capacity of infusions and



**Figure 3**. A. Protection (expressed as a percentage) following the ingestion of infusions made using island and continental populations of *Ugni molinae* in the formation of TBARS and Conjugated Dienes. *B. Trolox* equivalents (TEAC) following the ingestion of infusions of island and continental populations of U. molinae through TEAC-CUPRAC Values are the mean $\pm$ SD; n=12, by group. \*p < 0.01.

aqueous extracts prepared from *Ugni* leaves of Juan Fernández archipelago are higher than those of the continent. Conditions of the territory in which the *Ugni molinae* Turcz., Myrtaceae, populations growth may explain such differences. The plasma antioxidant capacity of the healthy volunteers differed before and after the ingestion of teas made from the leaves of the continental and island populations, following the dosages of traditional medicine, favoring the island populations. These results let us to propose the population of the island as a source of study for the development of an antioxidant supplement, and thereby contribute to the use of this species that has becoming an ecological problem in the island.

#### Acknowledgments

This work was supported by CONICYT (24091006) and the postgraduate council of the University of Concepción, (D.I. 210.074.043), and Proyecto Anillo (ADI-38), Anillo ACT-38, Proyecto Nuestra Flora de Chile, CONAF, Armada de Chile.

#### References

- Aguirre MC, Delporte C, Backhouse N, Erazo S, Letelier ME, Cassels BK 2006. Topical anti-inflammatory activity of 2 α-hydroxy pentacyclic triterpene acids from the leaves of *Ugni molinae*. *Bioorg Med Chem Lett 14*: 5673-5677.
- Apak R, Güclü K, Demirata B, Özyürek M, Celik SE, Bektasoglu B 2007. Comparative evaluation of various total antioxidant capacity assays applied to phenolic compounds with the CUPRAC assay. *Molecules 12*: 1496-1547.
- Avello M 2000. Estudio fitoquímico de hojas de *Ugni molinae* (Murtilla) y evaluación de la actividad antioxidante de sus componentes. Concepción, 100p. Trabajo de Tesis para optar al Título de Químico Farmacéutico, Facultad de Farmacia, Universidad de Concepción.
- Avello M 2004. Estudio fitoquímico de los extractos de Ugni molinae (Murtilla) y evaluación del efecto protector de sus componentes sobre endotelio vascular humano frente a lipoproteínas de baja densidad oxidadas (oxLDL). Concepción, 200p. Trabajo de Tesis para optar al grado de Magíster en Ciencias Farmacéuticas, Facultad de Farmacia, Universidad de Concepción.
- Avello M, Pastene E 2005. Actividad antioxidante de infusos de Ugni molinae Turcz. (Murtilla). Blacpma 4: 33-39.
- Avello M, Valdivia R, Mondaca MA, Ordóñez JL, Bittner M, Becerra J 2009. Actividad de Ugni molinae Turcz. frente a microorganismos de importancia clínica. Blacpma 8: 141-144.
- Cuevas J, Marticorena A, Cavieres LA 2004. New additions to

the introduced flora de of the Juan Fernández islands: origin, distribution, life history traits and, potential of invasion. *Rev Chil Hist Nat* 77: 523-538.

- Davies K, Schwinn K 2006. Molecular biology and biotechnology of flavonoid biosynthesis. In: *Flavonoids chemistry, biochemistry and applications*. U.S: Taylor & Francis Group, LLC, editors, p. 1151-152.
- Esterbauer H, Striegl G, Puhl H, Rotheneder M 1989. Continuous monitoring of *in vitro* oxidation of human low density lipoprotein. *Free Radical Res Com* 6: 67-75.
- Greimler J, Stuessy TF, Swenson U, Baeza CM, Matthei O 2002. Plants invasions on an oceanic Archipiélago. *Biol Invasions 4*: 73-85.
- Gugliucci A 1996. Antioxidant effects of Ilex paraguayensis: induction of decreased oxidability of human LDL *in* vivo. Biochem Bioph Res Com 224: 338-44.
- Hoffmann JA 1991.Flora silvestre de Chile zona araucana. Chile: Editorial Claudio Gay.
- Joyeux M, Mobstein A, Anton R, Mortier F 1995. Comparative antilipoperoxidant antinecrotic and scavenging properties of terpenes and biflavones from Ginkgo and some flavonoids. *Planta Med 61*: 126-129.
- Landete JM 2011. Ellagitannins, ellagic acid and their derived metabolites: A review about source. metabolism, functions and health. *Food Res Int 44*: 1150-1160.
- Lastra H, Rodríguez E, Ponce H, González ML 2000. Método analítico para la cuantificación de taninos en el extracto acuoso de romerillo. *Rev Cubana Plant Med 5*: 17-22.
- Meotti FC, Luiz, AP, Pizzolatti MG, Kassuya CAL, Calixto JB, Santos AR 2006. Analysis of the antinociceptive effect of the flavonoid myricitrin: evidence for a role of the L-Arginine-nitric oxide and protein kinase C pathways. J Pharmacol Exp Ther 316: 789-796.
- Ricci M 1989. Programa de conservación y recuperación de plantas amenazadas de Juan Fernández, Informe Final, 1ª etapa. Chile: Proyecto CONAF-WWWF-3313.
- Ritter A, Muñoz-Carpena R, Regalado CM, Socorro AR 2003. Caracterización del transporte de solutos en suelos volcánicos agrícolas mediante TDR y simulación inversa. In: *Estudios de la zona no saturada del suelo*. Chile: Álvarez Benedi J, Marinero P, editors, p. 19-24.
- Rubilar M, Pinelo M, Ihl M, Scheuermann E, Sineiro J, Nuñez MJ 2006. Murta leaves (Ugni molinae Turcz.) as a source of antioxidant polyphenols. J Agr Food Chem 54: 59-64.
- Rubilar M, Jara C, Poo Y, Acevedo F, Gutiérrez C, Sineiro J 2011. Extracts of maqui (*Aristotelia chilensis*) and murta (*Ugni molinae* Turcz): sources of antioxidant compounds and α-glucosidase/ α-amylase inhibitors. J Agr Food Chem 59: 1630-1637.
- Salamanca G, Correa I, Principal J 2007. Perfil de flavonoides

e índices de oxidación de algunos propóleos colombianos. *Zootecnia Tropical 25*: 95-102.

- Selma MV, Espin JC, Tomás-Barberan FA 2009. Interaction between phenolics and gut microbiota role in human health. *J Agric Food Chem* 57: 6485-6501.
- Shimosaki S, Tsurunaga Y, Itamura H, Nakamura M 2011. Anti-allergic effect of the flavonoid myricitrin from Myrica rubra leaf extracts *in vitro* and *in vivo*. *Nat Prod Res 25*: 374-380.
- Skottsberg C 1953. Notas sobre la vegetación de las islas de Juan Fernández. *Revista Universitaria 35*: 195-207.
- Stuessy TF, Swenson DJ, Crawfford G, Anderson G, Silva M 1998. Plant conservation in the Juan Fernández Archipiélago, Chile. *Aliso* 16: 89-101.
- Suwalsky M, Orellana P, Avello M, Villena F, Sotomayor C 2006. Human erythrocytes are affected in vitro by extracts of *Ugni molinae* leaves. *Food Chem Toxicol* 44: 1393-1398.
- Suwalsky M, Orellana P, Avello M, Villena F 2007. Protective effect of *Ugni molinae* Turcz against oxidative damage of human erythrocytes. *Food Chem Toxicol 45*: 130-135.
- Velioglu YS, Mazza G, Gao L, Oomah BD 1998. Antioxidant activity and total phenolics in selected fruits,

vegetables and grain products. *J Agr Food Chem 46*: 4113-4117

- Vermerris W, Nicholson R 2006. Phenolic compound biochemistry. The Netherlands: Springer, Dordrecht.
- Vivanco J, Cosio E, Loyola V, Flores H 2005. Mecanismos químicos de defensa en las plantas. *Investigación y Ciencia 341*: 68-75.
- Waniska RD 2000. Structure, phenolic compounds and antifungal proteins of Sorghum caryotipes. In: Proceedings of International Consultation. India: Chandrashekar A, Bandyopadhyay R, Hall AJ. ICRISTAT editors, p. 72.
- Yokoshimo A, Moriwaki M 2005. Myricitrin degraded by simulated digestion inhibits oxidation of human lowdensity lipoprotein. *Biosci Biotech Bioch 69*: 693-699.

#### \*Correspondence

Marcia A. Avello Faculty of Pharmacy, University of Concepción PO BOX 237, Concepción, Chile maavello@udec.cl Tel: 56 041 2204523