FLAVONOIDS IN Astragalus corniculatus

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Nine flavonoids were identified in aerial parts of *Astragalus corniculatus* Bieb. (Fabaceae) by liquid chromatography coupled with ionspray mass spectrometry in the tandem mode (LC/MS/MS) with negative ion detection. Vitexin, orientin and eriodictyol-7-*O*-glucoside are obtained for the first time in genus *Astragalus* L, and isorhamnetin-3-*O*-glucoside in the species.

Keywords: Fabaceae; Astragalus corniculatus; flavonoids.

INTRODUCTION

Astragalus corniculatus Bieb. (Fabaceae) is a perennial herbaceous plant, distributed in Moldova, South Ukraine and Southeastern Romania¹. The species was recently found in Bulgarian flora². Our earlier investigations of the ethyl acetate extract of this species, containing flavonoids, resulted in a low acute oral toxicity and a remarkable antihypoxic activity, especially in a model of circulatory hypoxia³.

The aim of this study is to determine the flavonoids in the ethyl acetate extract of *A. corniculatus* using liquid chromatography coupled with ionspray mass spectrometry in tandem mode (LC/MS/MS).

EXPERIMENTAL

General procedures

LC analysis was performed on a Agilent 1100, Model G1312A O (Hewlet Packard, Palo Alto, CA, USA). An Aqua C18 125 A (150 x 3.0 mm i.d., 5 mL) (Phenomenex, Torrance, CA, USA) was used. Gradient elution was carried out with water - 0.1% formic acid and water - acetonitrile - 0.1% formic acid at a constant flow rate of 400 μL min⁻¹. The MS and MS/MS data were obtained by using an API 365 tripe-quadroupole mass spectrometer (Perkin-Elmer Sciex, Concord, ON, Canada). All the analyses were performed using a Turbo Ionspray source. The operating parameters as follows: capillary voltage – 3500 V, nebolizer gas (N₂; 10 arbitrary units), curtain gas (N2; 8 arbitrary units), draying gas (N2; 7000 cm³/min-1), collision gas (N2; 5 arbitrary units), focusing potential - 240 V and entrance potential 10 V. The collision energy (CE) and declustering potential (DP) were optimized for each standard. Thin layer chromatography (TLC) was performed on silica gel plates (Kieselgel 60 F₂₅₄, Merck, Germany). The spots were visualized by spraying with 1% methanolic solution of diphenylboric acid aminoethyl ester (NST). Column chromatography (CC) was performed on cellulose (Watman, Germany), Polyamid S (Riedelde Haën, Germany) and Sephadex LH-20 (Pharmacia, Sweden).

Plant material

Astragalus corniculatus herbs were collected in July 1999 near the town of Svishtov, Bulgaria and identified by Dr. D. Pavlova. A voucher specimen has been deposited in the Herbarium of the Faculty of Biology, Sofia University, Bulgaria (SO95265).

Extraction and obtained of purified flavonoid fractions

Air-dried plant material (800 g) was powdered and extracted with 50% EtOH under reflux. After the removal of ethanol *in vacuo* the aqueous residue was consecutively treated with CHCl₃ and EtOAc. The EtOAc extract was evaporated to dryness to give a solid residue (14 g), which was submitted to column chromatography on cellulose, using 0-95% EtOH linear gradient. Further purification of combined fractions (TLC analysis) was achieved by rechromatography over Polyamid and Sephadex LH-20. Four main purified flavonoid fractions were obtained and analysed by LC/MS/MS.

RESULTS AND DISCUSSION

Solvent partition and repeated column chromatography over cellulose, Polyamide S and Sephadex LH-20 of the ethyl acetate extract to was submitted to fractionation, and the flavonoid fractions were analysed by LC/MS/MS. Standard solutions of 12 flavonoids were studied in the negative ion mode using MS/MS product ion scans (multiple reactions monitoring (MRM). For flavonol and flavanon Oglycosides, the spectra present both the deportment molecule [M-H] of the glycosides and the ion corresponding to the deprotonated aglycone [A-H]-. The latter ion is formed by loss of the sugar residue from the glycosides4. Fragmentation of aglycones provided characteristic ions for each family of flavonoids⁵. Isorhamnetin exhibits specific fragmentation with the loss of methyl radical, thus giving m/z 315 $\rightarrow m/z$ 300⁶. Hyperoside and isoquercetin have the same molecular mass and were identified together. Different fragmentation patterns were observed in MS/MS experiments for flavone-Cglycosides. Losses of 120 and 90 u were observed, corresponding to cross-ring cleavages in the sugar unit^{7,8} (Figure 1).

Figure 1. Fragmentation of vitexin

After MRM analysis of flavonoid fractions, nine flavonoids were identified as quercetin, quercetin-3-*O*-rutinoside (rutin), quercetin-3-*O*-galactoside (hyperoside), quercetin-3-*O*-glucoside (isoquercitrin), kaempferol, isorhamnerin, isorhamnetin-3-*O*-glucoside, isorhamnetin-3-*O*-rutinoside and eriodictyol-7-*O*-glucoside (Table 1).

Apigenin-8-C-glucoside (vitexin) and luteolin-8-C-glucoside

Table 1. Compounds identified by LC-MS/MS in the negative mode, compared with a standard

Flavonoids	Structure (m/z)	MS/MS ions (min)	t _r
Kaempferol	HO OH OH	285/151	21.20
Isorhamnetin	HO OH	315/300/151	21.50
Isorhamnetin -3- <i>O</i> -rutinoside	HO OL-Rha-D-Glc	623/315	14.10
Quercetin	HO OR OR	301/151	19.05
Quercetin-3- <i>O</i> -glucoside/galactoside	HO OH OH R- D-Gle R-D-Gal	463/301	13.93
Quercetin -3- <i>O</i> -rutinoside	HO OH O	609 / 301	13.54
Eriodyctiol -7- <i>O</i> -glucoside	Gic-D-O OH	449 /287	13.94

(orientin) were identified on the bases of the product ion spectrum and comparison with literature data⁹. Vitexin shows the ions at m/z 431 (deprotoned molecule), m/z 341 (loss of 90 u) and m/z 311 (loss of 120 u) as characteristic ions in the MS/MS mode (Table 2). The product ion spectra of the ion m/z 447 of orientin differ in relative abundance of the m/z 357 (loss of 90 u) and m/z 327 (loss of 120 u) ions (Table 2). Moreover, the ion spectra of the ion at m/z 429 in the spectrum of isoorientin, was not present in that of orientin and for isovitexin – the ion at m/z 353 wasn't absent in the spectrum of vitexin¹⁰.

Table 2. Compounds identified by LC-MS/MS in the negative mode, compared with literature data

Flavonoids	Structure	MS/MS ions m/z (relative abundance, %)	t _r (min)
Vitexin	HO GE	431 (43), 341(17), 311 (100), 269 (5)	12.80
Orientin	HO OH OH	447 (46), 357 (38), 327 (100), 285 (6)	13.62

CONCLUSIONS

LC/MS/MS analysis of ethyl acetate fractions from the aerial parts of *Astragalus corniculatus* led to the identification of nine flavonoids. Vitexin, orientin and eriodictyol-7-*O*-glucoside are identified for the first time in genus *Astragalus* L, and isorhamnetin-3-*O*-glucoside – in the species.

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REFERENCES

- Tutin, T. G.; Heywood, V. H.; Burges, N. A; Mooze, D. M.; Valeutine, D. H.; Walters, S. M.; Webb, D. A.; *Flora Europea*, Cambridge Univ. Press: Cambridge, 1972, Vol. 2, p. 108.
- 2. Pavlova, D.; Kozhuharov, S.; Herb J. Syst. Bot. 1994, 1, 17.
- Krasteva, I.; Nikolova, I.; Danchev, N.; Nikolov, S.; Acta Pharm. 2004, 54, 151.
- 4. Cuyckens, F.; Rosenberg, R.; de Hoffman, E.; Claeys, M.; J. Mass Spectrom. 2001, 36, 1203.
- Fabre, N.; Rusta, I.; de Hoffman, E.; Quetin-Leclercq, J.; J. Am. Soc. Mass Spectrom. 2001, 12, 707.
- 6. Justesen, U.; J. Mass Spectrom. 2001, 12, 169.
- 7. Becchi, M.; Fraisse, D.; Biomed. Environ. Mass Spectrom. 1989, 18, 122.
- Li, Q. M.; van den Heuvel, H.; Delorenzo, O.; Corthout, J.; Pieters, L. A.; Vlietinck, A. J.; Claeys, M.; J. Chromatogr. 1991, 562, 435.
- Sanchez-Rabaneda, F.; Jauregui, O.; Casals, I.; J. Mass Spectrometry 2003, 38, 35.
- Waridel, P.; Wolfender, J. L.; Ndjoko, K.; Hobby, K. R.; Major, H. J.; Hostettmann, K.; J. Chromatogr. 2001, 926, 29.