Patterns of Infection with the Nematodes *Syphacia obvelata* and *Aspiculuris tetraptera* in Conventionally Maintained Laboratory Mice

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Data on the frequency, distribution and mean intensity of the helminth fauna recovered from outbred and inbred mice conventionally maintained in Brazilian animal houses, are reported. The oxyurid nematodes Syphacia obvelata and Aspiculuris tetraptera presented overall frequencies of 91.5% and 8.5%, respectively. The frequency of S. obvelata in animals of three groups out of the four investigated ranged from 9% to 74% and A. tetraptera from 17% to 83%, since animals of one of the groups were negative for helminths. Infections due to a single species were observed in 62% of the animals, compared to 16% related to associations. The frequency of single infections in each group varied from 58.6% to 100% whereas associations varied from 24.1% to 41.4%. The analysis of specific mean intensities showed that S. obvelata was represented by 13.35 to 66.58 specimens/host and A. tetraptera by 5.85 to 16.75 specimens/host.

Key words: nematodes - Syphacia obvelata - Aspiculuris tetraptera - laboratory mice - Mato Grosso do Sul - Brazil

Laboratory animals are suitable and necessary for the proper development of several biological assays. The utilization of these standard models is recommended aiming at the attainment of reliable and reproductible results. In despite of this approach, laboratory animals are seldom investigated for autochthonous ecto and endoparasites prior to their utilization in experiments. In conventional, semi-open facilities, rodent colonies are frequently infected with helminths. These parasites, if undetected, can interfere in the development of protocols and alter the interpretation of final results (Pinto et al. 1994, 2001, Gonzalez 1996, Luca et al. 1996, Gonçalves et al. 1998). The analysis of different biological parameters related to the presence of nematodes in mice conventionally kept in animal houses in which sanitary conditions and barriers have not been properly controlled, is presented.

MATERIALS AND METHODS

From August 1999 to January 2000, 116 adult male mice [(Mus musculus (Linnaeus, 1758)] 42 days old, mean weight 28 g, conventionally maintained in four institutional facilities in Campo Grande, State of Mato Grosso do Sul, Brazil, were randomly chosen to be investigated for helminths. For ethical reasons, suppliers were not named and are identified as groups A, B, C, D, according

Animals were sacrificed in an ether chamber according to ethical procedures (Apa 1989) and necropsied in accordance to Pinto et al. (1994). Cecum and colon were opened longitudinally in individual Petri dishes with an 0.85% NaCl solution. Nematodes were recovered alive and identified under a stereoscope microscope and fixed in a hot 10% formaldehyde solution. Classification of the nematodes follows Pinto et al. (1994). Deposited material: Helminthological Collection of the Oswaldo Cruz Institute 34667, 34668 (wet material).

For the quantification of specific worm burdens, six ranges of distribution were considered (Pinto et al. 1994) as follows: I: 1-20, II: 21-40, III: 41-60, IV: 61-80, V: 81-100, VI: +100. Ecological terms (frequency, distribution and mean intensity) are in accordance to Bush et al. (1997).

to their source. Each group contained 29 animals: groups A, B, D of outbred Swiss Webster (SW) mice and group C of inbred BALB/c mice. Climatization in the four animal facilities (A, B, C and D), at the moment that the animals were received, was achieved by means of air conditioning devices and natural ventilation. For animals of group A, values of temperature and relative humidity were obtained with a thermohygrometer and daily measurements registered in the morning and afternoon. The values were included in charts so to obtain daily, monthly, semestral and annual profiles. Semestral values from August 1999 to January 2000 ranged from 25.14°C (morning) to 26.46°C (afternoon) whereas the relative humidity values were of 59.7% (morning) and 57.3% (afternoon). Sanitary control of animals of group A consisted in semestral ecto and endo parasitological examination. Temperatures of B and D facilities were obtained daily with a thermometer and mean temperatures were of 24 ± 2 °C. General husbandry, maintainance procedures and sanitary conditions are in

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TABLE I
Husbandry and maintenance of mice

Maintenance	Animal		Facility	
Groups	A	В	С	D
Ambiental control				
Temperature	Yes	Yes	No	Yes
Relative humidity	Yes	No	No	No
Periodical sanitary control	Yes	No	No	No
Hygiene and biosecurity				
Material sterilization	No	No	Yes (bottles)	No
Disinfection	Yes	Yes	Yes (cages)	Yes (cages)
Cages cleaning	2x/week	2x/week	2x/week	2x/week
General cleaning	Weekly	2x/month	2x/month	2x/month
Room cleaning	3x/week	Weekly	weekly	Weekly
Appropriate clothing	Yes	No	No	No
Restrictions (jewelery, cosmetics)	Yes	No	No	No
Bath	2x/day	No	No	No
Overall changing	2x/day	Daily	Daily	Daily
Shoes protector changing	2x/day	Daily	Not utilized	Not utilized
Staff				
Professionals in animal care	Yes	No	No	No
Trainees	Yes	Yes	No	No

The age, mean weight of animals and mean intensities of infections were analyzed by means of Graph Pad Instant statistical program and the ANOVA test.

The development of the present protocol has been authorized by the Committee of Ethics for the Use of Animals (CEU-Fiocruz), no. P0072-01.

RESULTS

Mean values of weight and age of animals were evaluated in the different groups (Table II). The nematodes Syphacia obvelata (Rudolphi, 1802) Seurat, 1916 (Figs 1a,b) and Aspiculuris tetraptera (Nitzsch, 1821) Schulz, 1924 (Figs 2a,b) were recovered from the investigated mice. Total number of worms was of 2,885. S. obvelata was represented by 2,613 specimens with a frequency of 91.5%, compared to 242 A. tetraptera worms with a frequency of 8.5% (Fig. 3). The highest frequency of *S. obvelata* occurred in the SW mice of group B with 1,931 worms (74%) followed by those of group D, with 455 worms (17%). In the group C of BALB/c mice, 227 worms (9%) were recovered; the highest frequency of A. tetraptera was observed in mice of group B, with 201 worms (83%), whereas in mice of group D, with 41 worms, the frequency was of 17% (Fig. 4). Animals of group A were not parasitized.

According to the pattern of worm burden distribution, *S. obvelata* appeared in a larger number of mice in range I. Parasitism in mice of group B, was distributed in all ranges; in mice of group C worm burdens were represented in ranges I-III (Fig. 5b) and those of group D, in ranges I-III and V (Fig. 5c). Similar patterns were observed for infections with *A. tetraptera*: the largest number of parazitized mice harbored worm burdens included in range I; animals of group B presented burdens in ranges I, IV and V (Fig. 5a). Parasitism in mice of group D was represented only in range I (Fig. 5c).

Statistical analysis of mean intensities of parasitism with *A. tetraptera* did not show significant differences between the groups B and D (16.75 \pm 28.59 and 5.85 \pm 5.46) and between the groups C and D (13.35 \pm 14.17 and 17.5 \pm 19.90) with mice infected with *S. obvelata*; nevertheless, significant differences related to the mean intensities of parasitism by *S. obvelata* were observed in the comparison of mice of groups B and C (66.58 \pm 82.75 and 13.35 \pm 14.17) and those of groups B and D (66.58 \pm 82.75 and 17.5 \pm 19.90) (Table III).

DISCUSSION

The improvement of laboratory animals either considering genetic approaches in the utilization of homogeneous strains or on the basis of sanitary conditions in an attempt to maintain these animals free from pathogens is a priority, since undetected autochthonous parasitic infections in laboratory animals, even in the absence of clinical signs, may act as variables, during experimental assays (Eaton 1972, Saiz-Moreno et al. 1983, Jacoby & Fox 1984, Huerkamp 1993, Pinto et al. 1994, 2001, Sato et al. 1995, Rehbinder et al. 1996, Rosas 1997, Gonçalves et al. 1998, Ortiz et al. 2000). In conventional animal facilities, rodent colonies are frequently infected with helminths or become infected in the laboratories where they are mantained in the course of experiments.

The age of the mice to be utilized is a parameter to be considered in the evaluation of pre-existing worm burdens in these experimental hosts. According to Flynn (1973) and Jacoby and Fox (1984), infections with *S. obvelata* generally occur in young mice, since adults seem to be more resistant; thus, 4-5 weeks mice should be investigated considering that prevalences of worm burdens are higher at this time. Panter (1969) and Taffs (1976) reinforce the acquired resistance to infection between the 4th

TABLE II Values (mean \pm standard deviation) of weight and age of mice

Groups	A	В	С	D
Weight	27.62 ± 4.87a	33.34 ± 3.86b	23.76 ± 3.60c	$29.72 \pm 4.07a$
Age (days)	$41.34 \pm 11.68a$	$48.00 \pm 4.81b$	$40.72 \pm 8.51a$	$45.45 \pm 4.17b$

ANOVA test (p < 0.05), n = 29; values in the same line, with the same letter are not significantly different.

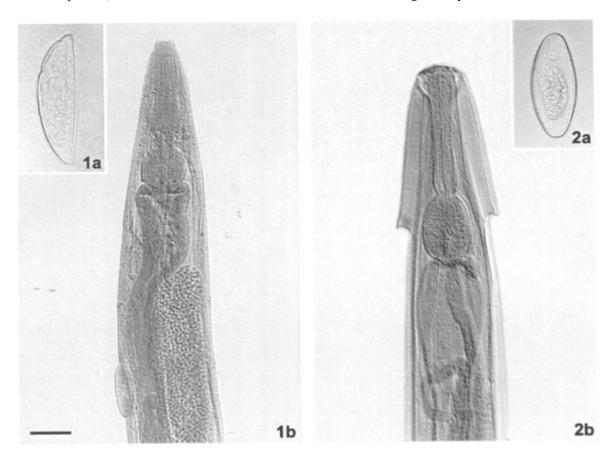


Fig. 1: Syphacia obvelata. a: egg (bar = 0.03 mm); b: anterior extremity (bar = 0.02 mm). Fig. 2: Aspiculuris tetraptera. a: egg (bar = 0.02 mm); b: anterior extremity (bar = 0.05mm). Bar common to Figs 1 and 2. Figs 1b and 2b were obtained in a Differential Interference Contrast (DIC) system.

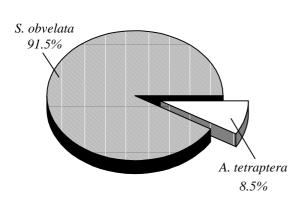


Fig. 3: frequency of Syphacia obvelata and Aspiculuris tetraptera.

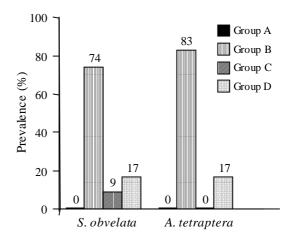
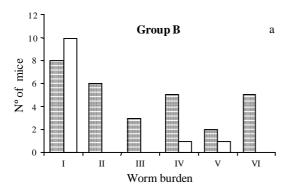
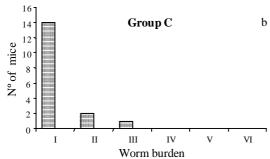


Fig. 4: frequencies of *Syphacia obvelata* and *Aspiculuris tetraptera* in mice of different groups.

and 9th weeks after birth, whereas infections with *A. tetraptera* affect older mice. Taffs (1976) reported to the age related resistance in mice, associated to an increase of mucus production and to the natural aging physiological process; nevertheless, no specific immune response was detected. Mean values of ages, related to worm burdens recovered, are in accordance with data previously reported (Flynn 1973, Jacoby & Fox 1984).





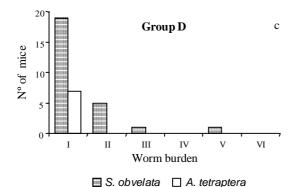


Fig. 5a-c: distribution of worm burdens, according to the different ranges. I: 1-20 worms; II: 21-40 worms; III: 41-60 worms; IV: 61-80 worms; V: 81-100 worms; VI: +100 worms.

The high frequency of *S. obvelata* in comparison with *A. tetraptera* worm burdens presently observed is not in accordance with Rosas (1977) that reports higher frequencies of the latter species; nevertheless, this high frequency of *S. obvelata* in rodent colonies is justified considering the nematode life-cycle, that is shorter in this species, thus inducing the infection in a larger number of mice in short periods (Flynn 1973, Jacoby & Fox 1984, Scott & Gibbs 1986, Coghlan et al. 1993, Moulia et al. 1993, Klement et al. 1996, Zenner 1998).

Gonçalves et al. (1998) observed associations of *S. obvelata* and *A. tetraptera* in outbred and inbred mice; this parameter is to be analyzed with basis on the population dynamics of pinworms in mice, as previously reported (Scott & Gibbs 1986). Hayunga (1991) and Gonçalves et al. (1998) agree in the statement that associations of different helminth species in a same host specimen induce the establishment of competition for nutrients and overgrowth of the parasites; however, according to Scott and Gibbs (1986) this competition is not related to the habitat since the site of infection of *S. obvelata* is the cecum, while *A. tetraptera* is recovered mainly from the colon; thus, this factor is not to be considered as the principal in these associations.

In the comparison of the present results with those reported by Gonçalves et al. (1998) referring to worm burdens, inbred mice investigated here, harbored smaller parasite loads and are in accordance with Higgins-Opitz et al. (1990) and Moulia et al. (1995) that refer to lower prevalences of *S. obvelata* in inbred mice strains.

The distribution of worm burdens evaluated in the present study agrees with previous data (Anderson & Gordon 1982, Pinto et al. 1994). According to Anderson and Gordon (1982), the aggregation of helminths in the host populations are common to occur under laboratory and field conditions. These aggregates are due to the heterogeneous susceptibility of mice to infections, related either to behavior, genetics background, immunological status or to the concentration of infective eggs in the environment increasing the possibility of accidental infections (Scott & Gibbs 1986).

The absence of parasites in mice from the animal facility A is, undoubtely, due to the more adequate adopted procedures and qualified working staff that act as barriers against helminth infections, when compared to animal houses B, C and D (Table I). Despite the fact that cages in the animal facility A are openly cleaned, disinfected and dried under the sun, this aspect seems to improve hygiene conditions, since according to Taffs (1976) oxyurid eggs although highly resistent to the cold and disinfection, are extremely susceptible to sunlight and heat.

TABLE III

Mean intensities and standard deviation of the parasitism by Syphacia obvelata and Aspiculuris tetraptera in necropsied mice

		Groups			
	A	В	С	D	
S. obvelata	-	$66.58 \pm 82.75a$	$13.35 \pm 14.17b$	$17.5 \pm 19.30b$	
A. tetraptera	-	$16.75 \pm 28.59a$	-	$5.85 \pm 5.46a$	

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