

Systematic review on the biology, ecology, genetic diversity and parasite transmission potential of *Panstrongylus geniculatus* (Latreille 1811) in Latin America

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Panstrongylus geniculatus (Latreille, 1811) is the triatomine with the largest geographic distribution in Latin America. It has been reported in 18 countries from southern Mexico to northern Argentina, including the Caribbean islands. Although most reports indicate that *P. geniculatus* has wild habitats, this species has intrusive habits regarding human dwellings mainly located in intermediate deforested areas. It is attracted by artificial light from urban and rural buildings, raising the risk of transmission of *Trypanosoma cruzi*. Despite the wide body of published information on *P. geniculatus*, many knowledge gaps exist about its biology and epidemiological potential. For this reason, we analysed the literature for *P. geniculatus* in Scopus, PubMed, Scielo, Google Scholar and the BibTriv3.0 databases to update existing knowledge and provide better information on its geographic distribution, life cycle, genetic diversity, evidence of intrusion and domiciliation, vector-related circulating discrete taxonomic units, possible role in oral *T. cruzi* transmission, and the effect of climate change on its biology and epidemiology.

Key words: *Panstrongylus geniculatus* - geographic distribution - genetic diversity - oral transmission of *Trypanosoma cruzi* - climate change

Chagas disease affects around 7 million people in Latin America and is considered one of the 10 most important neglected diseases in the region.^(1,2) Currently, 154 species within the Triatominae subfamily (three extinct and 151 extant) have been described and are able to transmit *Trypanosoma cruzi*, the causative agent of Chagas disease under natural and experimental conditions.^(2,3,4,5,6,7,8)

Panstrongylus geniculatus (Latreille, 1811) is the triatomine with the greatest geographic distribution in the Americas, ranging from southern Mexico to northern Argentina and including the Caribbean islands.^(9,10) This vector occupies extremely variable areas across the wild landscape, being found in different ecosystems within the same country.⁽¹¹⁾ Most records deal with the species occupying altitudes ranging from sea level to around 2,000 metres above sea level (masl); however, the species has been found at altitudes from 2,000 to 4,000 masl in some Andean countries (i.e., Colombia, Venezuela, Ecuador, Peru and Bolivia), thereby indicating the species' great adaptive capability, probably enabling its shift in altitude as a consequence of lowland warming, regarding today's climate changes.^(12,13,14,15,16)

The last few decades have seen much interest in the biology, ecology and epidemiology of *P. geniculatus*, thereby enabling effective control strategies for this species to be designed. Therefore, this review aims to update existing knowledge about *P. geniculatus* and address current information gaps. A search was conducted in Scopus, PubMed, Scielo, Google Scholar, BibTriv3.0 databases with no filters of language or time and until August 2020. All relevant studies on taxonomy, morphological variability, life cycle, geographical distribution, genetic diversity, intrusion and colonisation of human dwellings, oral transmission and discrete taxonomic units of *T. cruzi* and possible effects of climate change on *P. geniculatus* were chosen.

Taxonomy, morphology and life cycle

Taxonomy - Currently, *P. geniculatus* (Latreille, 1811) is the accepted scientific name for this species; however, the following synonyms have also been used reviewed by Patterson et al.:⁽¹⁷⁾ *Reduvius geniculatus* (Latreille, 1811), *Conorhinus lutulentus* (Erichson, 1848), *Lamus geniculatus* (Stål, 1859), *Conorhinus corticalis* (Walker, 1873), *Conorhinus geniculatus* (Walker, 1873), *Lamus corticalis* (Lethierry & Severin, 1896), *Triatoma geniculata* (Chagas, 1912), *Triatoma tenuis* (Neiva, 1914), *Triatoma fluminensis* (Neiva and Pinto, 1922), *Mestor geniculatus* (Brindley, 1931) and *Panstrongylus parageniculatus* (Ortiz, 1971).

Fifteen species have been described in the *Panstrongylus* genus: *P. chinai*, *P. diiasi*, *P. geniculatus*, *P. guentheri*, *P. howardi*, *P. hispaniolae*, *P. humeralis*, *P. lenti*, *P. lignarius*, *P. lutzi*, *P. megistus*, *P. martinezorum*, *P. mitarakaensis*, *P. rufotuberculatus*, *P. tupynambai*.^(2,3,17)

doi: 10.1590/0074-02760200528

Financial support: Colombian Science, Technology and Innovation Department (Colciencias) project 120465843375 contract 063-2015 (<http://www.colciencias.gov.co/node/1119>) to FG, GAV and JDR; University of Tolima Research Fund (project 770115).

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Received 07 October 2020

Accepted 26 January 2021



Studies on the taxonomy of this genus and the identification of *P. geniculatus* have been problematic because most are based on morphometry and karyotyping, but few have employed molecular markers.^(17,18,19,20) The phylogenies based on morphological traits presents the *Panstrongylus* genus as a monophyletic group, whereas the ITS-2 (rDNA) phylogenetic trees suggest that it is polyphyletic group of *Triatoma* species from South, Central and North America.^(9,21) In another study, using four mitochondrial markers (16S, COI, COII, Cytb) and two nuclear (18S and 28S) ones, *Panstrongylus* fell into two groups. One of the groups contained *P. tupyambai*, *P. lutzi* and *P. geniculatus* as sister taxa of *Nesotriatoma*, whereas the other was a highly supported group that includes *Triatoma tibiamaculata* and *P. megistus*.⁽²²⁾

The use of morphological characters to classify *Panstrongylus* species has led to the description of 'new species' in the genus. This is the case for two species described in French Guiana and Venezuela; namely, *P. mitarakaensis* and *P. martinezorum*, respectively.^(23,24) They both look quite similar to *P. geniculatus* but no molecular methods have been used to confirm that they are the same species.⁽³⁾ Another species in Venezuela, *P. turpiali*, was synonymised with *P. chinai* but both were

then shown to phenotypically match *P. geniculatus*.^(25,26,27) Therefore, it is necessary to use molecular approaches to clarify the taxonomic status of *P. geniculatus* and phenotypically similar species.

Morphological variability in *P. geniculatus* - Several authors have reported on the morphological characteristics of *P. geniculatus*. Its nymph and adult stages have been described using light and scanning electron microscopy.^(9,17,19,28,29,30,31,32,33,34,35,36) Elsewhere, the geometric morphometric techniques applied to a sylvatic population and its laboratory descendants over five generations revealed size differences in *P. geniculatus* but not in its shape. Essentially, head size and wing size were both reduced from sylvatic and laboratory populations.⁽³⁵⁾ Another morphometric study on *P. geniculatus* showed a reduction in the size of its populations in Caracas city, when compared with sylvatic populations from other Venezuelan regions.⁽²⁰⁾ In this review we have included unpublished photographs representing the known morphological characteristics of the species, which we believe will help to a primary identification of individuals captured in sylvatic, peridomestic or domestic environments (Fig. 1A-G).

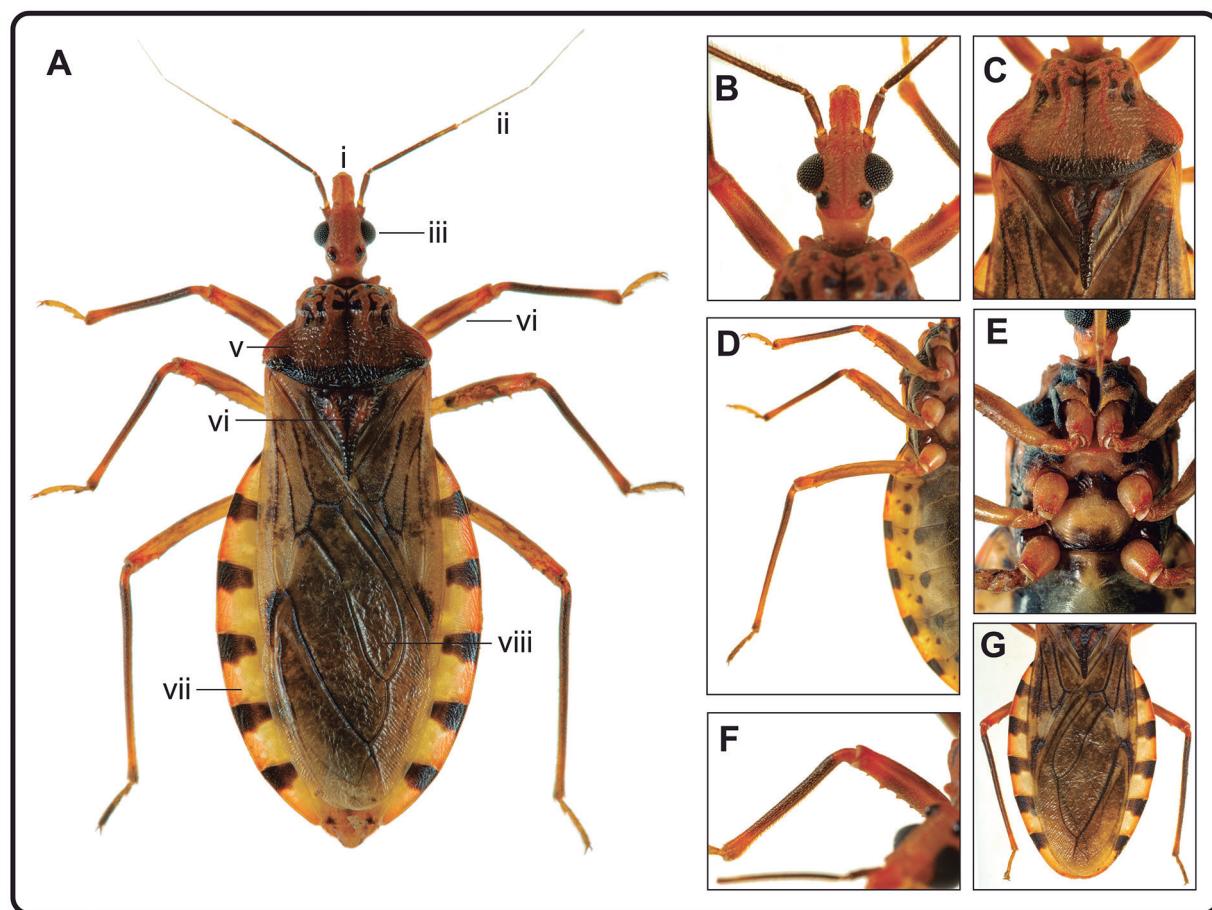


Fig. 1: (A) dorsal view of a female *Panstrongylus geniculatus*: (i) clipeus, (ii) antenna, (iii) compound eye, (iv) leg, (v) pronotum, (vi) scutellum, (vii) connexivum, (viii) wings (hemielytrons); (B) head; (C) pronotum and scutellum; (D) legs; (E) coxa and trochanter; (F) spicules; (G) connexivum. Source: authors.

Adult *P. geniculatus* insects display sexual dimorphism. The total length of the male is 22–28 mm and the larger-bodied female is 22.5–29.5 mm. The posterior end of the male's abdomen is rounded in shape, whereas it is tip-shaped in the female (Fig. 2I–J). Its colouration is light brown to orange brown and is interspersed with dark patches on various parts of the body, which do not follow a defined pattern in that the connexivum has two intercalated regions of colour: a flat black-coloured one and another coloured orange to ochre (yellow-orange) in the interior (Fig. 1A).⁽³⁶⁾

The head is sub-conical-shaped, narrow at eye level, and shorter than the pronotum; the antecular region is approximately twice as long as the postocular region. The dorsal surface is slightly rough. The general colouration is ochre, but in some individuals two dark stripes extending from the base of the jugae to the space be-

tween the ocelli are observed in the dorsal region. Anteniferous tubers are located near the anterior edges of the eyes, and the antennae vary from reddish brown to black. The first segment of the antenna subtly exceeds the apex level of the clypeus (Fig. 1A–B).

Behind the head is the orange-brown pronotum whose anterior lobe is slightly convex and rough. The pronotum features a central four-leaf clover-shaped mark and “1 + 1” side marks. The posterior lobe is irregularly rough, with a black band along the posterior margin, except in the humeral area (Fig. 1C). The femurs and tibiae are dark brown or black (Fig. 1A, D). The legs have light yellow or orange-yellow coxas and trochanters (Fig. 1E). The front and middle femurs have two to six (usually four) small subapical denticles arranged in two rows (Fig. 1F). The hemelytrons are light brown or light yellowish brown, with dark veins in the membranes (Fig. 1G).

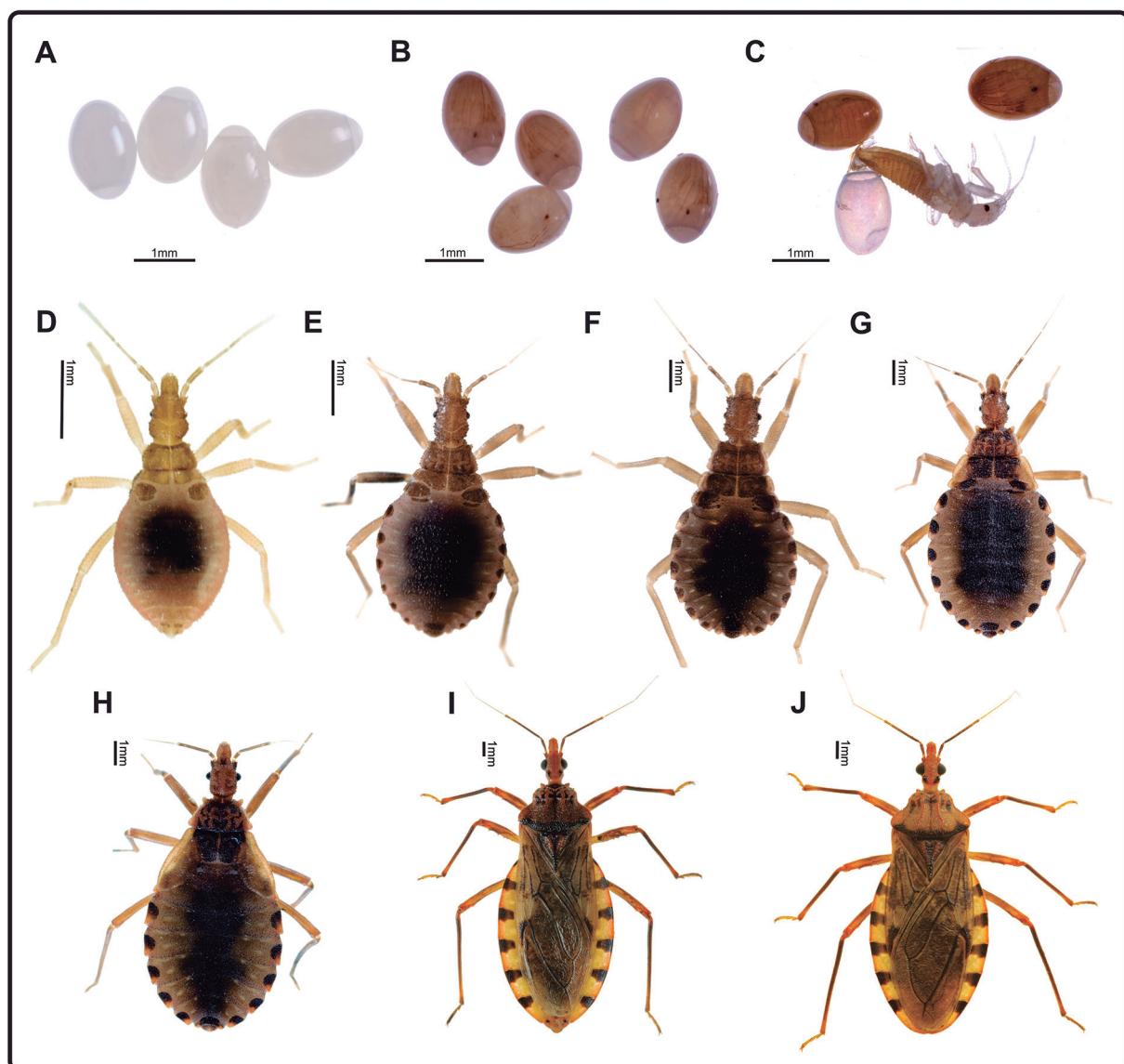


Fig. 2: stages of the life cycle of *Panstrongylus geniculatus*: (A) freshly oviposited eggs; (B) eggs near hatching; (C) hatching; (D) nymph stage N1; (E) nymph stage N2; (F) nymph stage N3; (G) nymph stage N4; (H) nymph stage N5; (I) adult (female); (J) adult (male). Source: authors.

Life cycle of P. geniculatus under laboratory conditions - *P. geniculatus* eggs do not adhere to the substrate; rather, they are oviposited free on a surface. They are oblong and symmetrically shaped, without lateral flattening or collar, and at one end is a prominent and convex operculum through which hatching is performed. The eggs are bright white for the first days after oviposition, and six-eight days later they turn pinkish yellow, pink, or pearl, becoming translucent after hatching (Fig. 2A-C). The nymphal and adult stages are shown in Fig. 2D-J.

The life cycle period under laboratory conditions has been shown to be dependent on the experimental conditions employed, such as temperature, relative humidity and food source in the colonies. Depending on the management of these variables, very heterogeneous records have been observed from 149.5 to 531 days, and even up to two years. When the experimental conditions are 21-

26°C and 90-100% for temperature and relative humidity, respectively, life cycle durations of 269, 297, 275.8, 273, 275.4 days have been reported (average, 278 days).^(32,37,38,39,40,41,42,43,44) One of the first studies to investigate the life cycle of *P. geniculatus* reported that the total development time is approximately 2 years.⁽⁴⁵⁾ Another study reported on the difficulties of establishing the species as a colony and observed that the time taken for the life cycle is 531 days (75.8 weeks).⁽³⁷⁾

The 1990s brought advances in biological knowledge. The life cycle was studied under controlled laboratory conditions and reported that the average development time was 269 days (38.4 weeks) and 297 days (42.4 weeks) for nymphs fed on chickens (*Gallus gallus*) and opossums (*Didelphis marsupialis*), respectively.⁽³⁸⁾ In a life-cycle study using uncontrolled conditions, the insects were subjected to temperatures of 21-25°C, relative

TABLE I
Geographic distribution of *Panstrongylus geniculatus* in Latin America

Country	Location	Reference
Argentina	Chaco, Corrientes, Entre Ríos, Formosa, Misiones, Santa Fe and Santiago Del Estero	9,10
Bolivia	Beni, Cochabamba, La Paz, Santa Cruz and Tarija	9,10,48,49,50,51
Brazil	Acre, Amapá, Amazonas, Bahia, Brasília DF, Ceará, Espírito Santo, Goiás, Maranhão, Mato Grosso, Minas Gerais, Pará, Paraná, Piauí, Rio de Janeiro, Rondônia, Roraima, São Paulo and Tocantins	9,10, 46, 52,53,54,55,56
Colombia	Amazonas, Antioquia, Arauca, Atlántico, Bolívar, Boyacá, Caquetá, Casanare, Cauca, Cesar, Choco, Córdoba, Cundinamarca, Guainía, Guaviare, Huila, La Guajira, Magdalena, Meta, Norte de Santander, Putumayo, Risaralda, Santander, Tolima, Sucre, Valle del Cauca, Vaupés and Vichada	9,10,15,57,58,59,60,61,62,63
Costa Rica	Alajuela, Cartago, Guanacaste, Heredia, Limón, Puntarenas and San José	9,11,64
Ecuador	Esmeraldas, Imbabura, Manabí, Napo, Orellana, Pastaza, Pichincha and Sucumbíos	9,10,65,66,67,68,69
Guatemala	ND	9
British Guiana	ND	9
French Guiana	Apotou, Awala-Yalimapo, Cayenne, Comopi, Grand-Santi, Kourou, Iracoubo, Macouria, Mana, Maripasoula, Matoury, Montsinery, Ouanary, Papaïchton, Roura, Remire-Montjoly, Regina, Saint-Elie, Saint-Georges, Saint-Laurent, Sinnamary and Saul	9,70,71
Mexico	Chiapas, Veracruz and Yucatan	10
Nicaragua	Boaco and Zelaya	9,72,73
Panama	Panama	9,74
Paraguay	Alto Parana, Boqueron, Concepcion, Caaguazu, Nueva Asuncion and Paraguari	9,10,46
Peru	Amazonas, Ayacucho, Bagua, Cajamarca, Cerro de Pasco, Cusco, Huanuco, Jaén, Junín, Loreto, Madre de Dios, Martín, Puno, San Ignacio, San Pasco, Ucayali and Utcubamba	9,16,46,75,76,77
Suriname	Commewijne, Para, Paramaribo, Saramacca, Sipaliwini and Wanica	9,78,79
Trinidad y Tobago	San Patricio and ST. George	9,80
Uruguay	ND	9
Venezuela	Amazonas, Anzoátegui, Aragua, Bolívar, Carabobo, Distrito federal, Delta Amacuro, Falcon, Guarico, Lara, Mérida, Miranda, Monagas, Táchira, Yaracuy and Zulia	9,10,81

humidities of 90-100%, and were fed *ad libitum* weekly on chickens (*G. gallus*). An N1 nymph stage to adulthood duration of 275.8 days (39.4 weeks) was reported.⁽³⁹⁾ The duration of the life cycle for specimens from the municipality of Amalfi in Antioquia, Colombia, which were subject to controlled temperature ($29 \pm 1^\circ\text{C}$) and relative humidity ($95 \pm 1^\circ\text{C}$) conditions, was 149.5 days (21.4 weeks).⁽³²⁾ Later on, evidence of the colonisation processes in Amalfi in Antioquia, Colombia, and estimated transmission risk indicator values were obtained. They also reported on the time required for blood intake by adults (mean, 67.8 minutes for males and 80.27 minutes for females). Blood consumption was on average 0.12 g for males and 0.26 g for females, with defecation times post-feeding of 55 minutes for females and 60 minutes for males.⁽⁴⁰⁾ Sylvatic intrusive insects captured in domestic environments have had high *T. cruzi* infection prevalence (see Table III in this review). In addition, a post-feeding time close to one hour for both males and females could be a determining factor for this species regarding the oral transmission of Chagas' disease due to food contamination. As *P. geniculatus* has been repeatedly associated with orally transmitted outbreaks, this hypothesis could be ascertained by comparing this species' post-feeding time characteristics and natural infection frequencies to those for other species invading human dwellings.

The first statistical evaluation of the population dynamics of *P. geniculatus* under controlled laboratory conditions studied a cohort of 60 eggs placed at constant temperature ($26 \pm 3^\circ\text{C}$) and relative humidity ($90 \pm 10\%$) that were fed every 15 days on chickens (*G. gallus*). The researchers observed that the hatching percentage was 88.9%, the average nymph development time was 273 days (39 weeks), and the longevity of the adults was 504 days (72 weeks).⁽⁴¹⁾

In contrast, the need of optimising the ovipositors of female *P. geniculatus* to enable massive breeding of this species in the laboratory and for use in bioassays, compared the ovipositors of laboratory and field populations fed on chickens (*G. gallus*) and mice (*Mus musculus*,

strain BALB/c). The authors found that the fertility of the first-generation laboratory population was twice that of the wild population when both were fed on chickens.^(42,43)

The most recent and detailed study on the vital statistics of *P. geniculatus*, estimated its development times by stages, vital statistics (mortality and fertility), and population growth parameters (intrinsic natural growth rate, finite population growth rate, net reproduction rate, and generational time). In a cohort of 100 eggs kept under constant temperature ($26 \pm 1^\circ\text{C}$) and relative humidity ($60 \pm 10\%$) and fed every eight days with chicken blood, it was observed that the total development time by stage was 275.4 days (39.4 weeks), and mortality reached 58% in the initial stages (I, II, and III), which represents 60% of the total mortality.⁽⁴⁴⁾

Geographical distribution, ecotopes and genetic variability

Geographical distribution and ecotopes - *P. geniculatus* has the greatest geographical distribution of species belonging to this genus. It is found exclusively in Latin America between 21.1°N , 30.3°S , 38.8°E , and 94.9°W , covering an estimated 12 million km². To date, its presence has been reported in 18 countries (Table I, Fig. 3), from southern Mexico to northern Argentina, including the Caribbean islands.^(9,12,45,46,47)

With its wide geographical distribution, *P. geniculatus* is found from sea level to 2,000-4,000 m above sea level in the Andean countries (Colombia, Venezuela, Ecuador, Peru and Bolivia)^(14,15,16) and in places where the average annual temperature (minimum and maximum) ranges between -3.9 and 34.3°C and precipitation reaches up to 4,000 mm/year.⁽⁹⁾ Some authors have suggested that the morphological variations found among *P. geniculatus* populations result from the demands of the different environments.⁽¹⁸⁾ Table II shows the presence of *P. geniculatus* above 2,000 masl in Colombia and Peru.

Natural ecotopes vary from dry or very dry tropical forests to savanna and humid tropical forests, and *P. geniculatus* can be found in caverns, caves, and burrows

TABLE II
Records of *Panstrongylus geniculatus* presence above 2,000 masl in Colombia and Peru

Country	Locality	State	Coordinates	masl	Reference
Colombia	Boavita	Boyaca	6.333333 N -72.666667 W	2,350	14,15
	Susacon	Boyaca	6.2298 N -72.6901 W	2,480	
	San Mateo	Boyaca	6.3303 N -72.585 W	2,200	
	Casabianca	Tolima	5.0796 N -75.1206	2,100	
Peru	Ninacaca	Pasco	-10.75819 S -76.17819 W	4,100	14,16
	Tarma	Junin	-11.419335 S -75.68849 W	3,050	
	Cholon	Huanuco	-8.690057 S -76.733652 W	2,350	
	Cutervo	Cajamarca	-6.376045 S -78.821275 W	2,600	
	San Miguel	Ayacucho	-12.932081 S -73.747786 W	2,660	
	Huanta	Ayacucho	-12.938411 S -74.250212 W	2,630	

masl: metres above sea level.

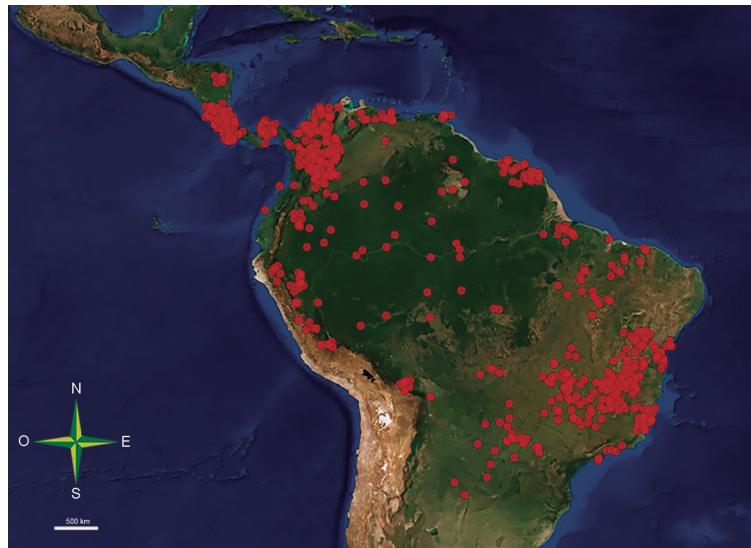


Fig. 3: continental distribution of *Panstrongylus geniculatus*. Maps were made with the database reported by Ceccarelli et al.⁽⁴⁷⁾

in these environments where it is mainly associated with armadillos (Dasypodidae), anteaters (Myrmecophagidae), bats (Chiroptera), and a wide variety of other vertebrates. It is also found under bark, tree trunks and fallen trees and near bird nests in palm trees (*Acrocomia aculeata*, *Syagrus romanzoffiana*, *Elaeis oleifera*, and *Leopoldinia piassaba*) and in epiphytes.^(9,17,82,83,84,85) Interestingly, it was reported that the species displays entomophagic behavior towards moths (*Eacles* spp.) in the forests of French Guyana for the first time, which confirms its versatility for survival in environments with low food availability.⁽⁸⁶⁾

Although the presence of *P. geniculatus* has been recorded in 18 countries, the recent availability of predictive maps can produce a robust summary of its distribution and show its spatial variation so that regions with a high probable risk of vector transmission and transmission through food contamination can be identified.⁽⁸⁷⁾

Genetic diversity in *P. geniculatus* - The *Panstrongylus* genus is classified as belonging to the Triatomini, a monophyletic tribe. However, the phylogenetic relationships inside this group are far to be cleared because current knowledge is limited and messy.⁽³⁾ The Triatomini tribe is divided into three lineages: *T. dispar*, North American, and South American lineages. *Panstrongylus* is included in the Antillean Triatoma + *Panstrongylus* clade within the North American lineage. Three groups were tentatively distinguished in the *Panstrongylus* sub-clade, but only one of them includes *P. geniculatus* and its closest relatives *P. mitarakaensis*, *P. martinezorum*;⁽³⁾ it also includes a set of south Amazonian species as *P. lutzi* (and its synonym *P. sherlocki*), *P. lenti*, *P. diasi*, *P. guentheri* and *P. tupynambai*.^(9,17,23,24,88,89)

Regarding karyotype variations in *Panstrongylus* species, it has been established that the species (*P. chinai*, *P. geniculatus*, *P. lignarius*, *P. rufotuberculatus*, *P. tupynambai*, *P. herreri*) have 20 autosomes (except *P. megistus* that has 18 autosomes), and X_1X_2Y chromo-

somes for males and $X_1X_1X_2X_2$ for females. In a study, three clusters were defined based on the C-band pattern and meiotic chromosomal behavior where *P. geniculatus* was not located in any of the groupings because it shares characteristics with each group. Consequently, the authors proposed that *P. geniculatus* should be considered as a complex of species comprising at least two different species because of its high chromosomal variability, wide distribution, and high phenotypic variation;⁽¹⁸⁾ molecular evidence, however, is needed to confirm this hypothesis.

A Venezuelan study conducted a genetic variability analysis on *P. geniculatus* using *Cytb* as the molecular marker because it is extensively used to determine genetic polymorphisms in triatomines. The results showed that this marker is useful for describing genetic heterogeneity in *P. geniculatus* and revealed spatial and genetic patterns that agree with domiciliation processes for this species, but it could not resolve the evolutionary relationships. Even though this marker is informative about the population structure, other genes must be evaluated to study the genetic variability.⁽⁹⁰⁾

A very recent study used DNA markers and molecular data to study genetic diversity in *P. geniculatus* with the aim obtaining information about the processes that have shaped genetic diversity in this species.⁽⁹¹⁾ Using mitochondrial DNA (mtDNA) fragments, 16S rRNA and *Cytb*, the authors reconstructed a phylogenetic tree that showed that *P. geniculatus* from Colombia and Venezuela form a monophyletic species with four clades concordant with its geographic distribution, which is partly explained by the Andes uplift (Fig. 4). However, other factors, including anthropogenic and eco-epidemiological effects must also be investigated to explain the existence of recent geographic *P. geniculatus* lineages. The authors also reconstructed the relationships within the *Panstrongylus* genus using various genetic markers. Using mtDNA markers (16S, ND4 and *Cytb*), they detected two clades: one includes *P. lutzi* and *P. tupynambai*,

TABLE III

Panstrongylus geniculatus adult specimens reported inside human dwellings in Latin American countries, without evidence of colonization (eggs, nymphs and exuviae)

Country	Department/State	<i>Trypanosoma cruzi</i> infection frequency	Reference
Argentina	Corrientes	ND	95
	La Paz	33%	49
Bolivia	Cochabamba	62,5%	51
	Paraná	Negative	96
	Piauí	Positive	54
	Goiás	1,40%	97
	Mato Grosso do Sul	3,20%	98
Brazil	Rio de Janeiro	Negative	99,100
	Amazonas	Positive	81,101
	Amazonas	Negative	102,103
	Minas Gerais	ND-Negative	104,105,106
	Distrito Federal	Negative	107
	Goiás	7,70%	108
	São Paulo	Negative	109,110
	Acre	Negative	111
	Bolívar	ND	112
	Meta	70,6-100%	93,113
Colombia	Magdalena	83%	114
	Casanare	ND	115
	Cordoba	30-50%	92,116
	Santander	56%	117
	Sucre	58,82%	118
Peru	Amazonas, Cajamarca	ND	76
	Pasco	18,20-50%	119,120
	Cusco	ND	121
	Loreto	ND	122
	San Martín	ND	123
Surinam	Saramacca, Wanica	ND	124
	Caracas	38,86-53%	125,126
Venezuela	Merida	Positive	127
	Caracas, Merida, Vargas	76,1% (average)	128
	Lara	ND	129
	Aragua	Negative	130
	Aragua, Caracas, Miranda, Vargas	50,3% (average of the four states)	84
	Sucre	ND	131

ND: no data.

which is the sister clade of *P. geniculatus* (*geniculatus* group), and the other one includes *P. lignarius* and *P. megistus*, which represent the sister clade of *P. rufotuberculatus*, *P. howardi* and *P. chinai* (*megistus* group).

To conclude, further studies using more genetic markers and genomic approaches, along with widespread sampling from different geographic locations are needed

to unravel the inter- and intra-specific relationships of the *Panstrongylus* genus, and specially for *P. geniculatus* considering its domiciliation reports, high infection rates with *T. cruzi* and incrimination in oral outbreaks of Chagas disease in Colombia and Venezuela. All this knowledge is relevant for the development of control and surveillance strategies for Chagas disease vectors.

Epidemiology, intrusion, colonisation of human habits and possible role in the oral transmission of *T. cruzi*

Evidence of dwelling intrusion by *P. geniculatus* in Latin America - Although *P. geniculatus* is considered a species of wild habits, several cases of intrusion into homes have been reported in different Latin American countries in recent decades. This new scenario implies an increased risk of transmission in endemic and non-endemic areas of Chagas disease, either by direct contact or via food contamination.^(92,93) Table III shows 45 registered reports of intrusion by adult *P. geniculatus* in Bolivia, Brazil, Colombia, Peru, Surinam and Venezuela. Despite growing awareness of the relevance of the transmission dynamics of *P. geniculatus*, the drivers of house invasion remain poorly understood. However, it was observed that invasion by this species decreased with higher landscape disturbance levels and in hotter-day municipalities and increased somewhat in more places with intermediate disturbance, peaking in municipalities with average rainfall.⁽⁹⁴⁾

It is accepted that the increased interactions occurring between triatomines and humans are mainly related to losses in vegetation cover and increases in urban

and rural populations invading natural ecotopes. Most of these intrusions occur at night when *P. geniculatus* fly in from wild environments to inhabited areas under the attraction of artificial lighting in search of food.⁽⁸⁴⁾ The author concluded that this occurred regardless of the type of housing, and that the invasion increased when high numbers of domestic animals were present and the distance to the forest was small.⁽⁸⁴⁾ Recently, 11 *P. geniculatus* specimens were captured in the city of Bucaramanga, a Colombian neighborhood, of which five out of nine tested were positive for *T. cruzi* (56%), which drew attention to the risk of infection by this vector settling in the periphery of homes and adjacent to the natural ecotopes where its life cycle in the wild occurs.⁽¹¹⁷⁾

The ability of *P. geniculatus* to fly up to 2 km away has been verified.⁽¹²⁵⁾ In other study, was reported that *P. geniculatus* is attracted to artificial light sources. These researchers installed a light trap positioned 45 m above sea level in a wild area of Manaus in the Amazonas State of Brazil, and they operated it monthly for three consecutive nights over the course of a year. They found that the most commonly captured triatomine was *P. geniculatus* (38 individuals) in addition to six other species.⁽¹³²⁾ Likewise, Jacome et al., determined the risk associated with the dispersive nocturnal flights of sylvatic triato-

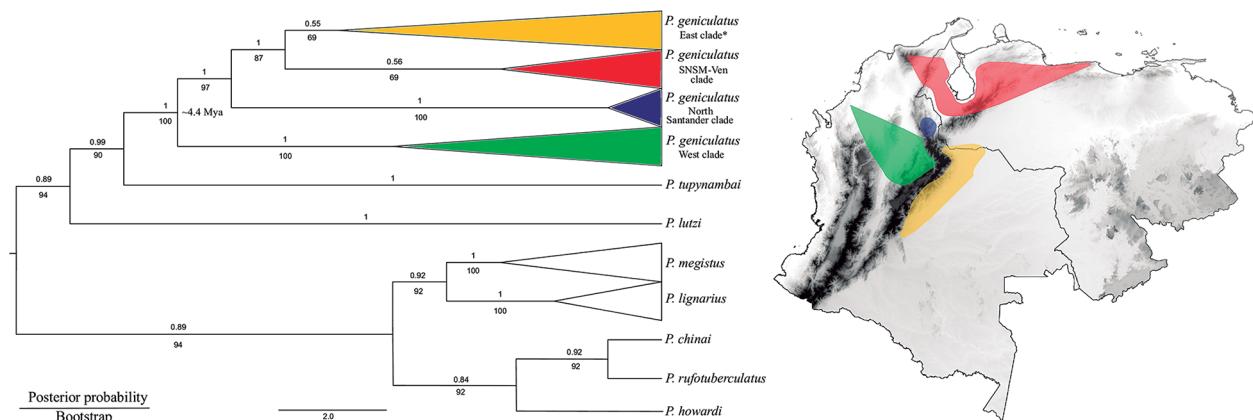


Fig. 4: genetic differentiation of *Panstrongylus geniculatus* in Colombia and Venezuela reported by Caicedo-Garzón et al.⁽⁹¹⁾

TABLE IV

Registration of *Panstrongylus geniculatus* adults, nymphs and eggs inside houses or peridomestic in Latin American countries

Country	Department/State	ID	PD	Association with	Natural infection	Reference
Bolivia	La Paz	X	X	ND	Negative	48
Brazil	Pará	X	X	Pigs	17%	134
	Antioquia	X	ND	ND	21,2-50%	40
Colombia	Atlántico	X	ND	ND	ND	135
	Casanare	X	ND	ND	Negative	136
Venezuela	Miranda	X	ND	Rats	20%	137
	Lara	X	ND	ND	9,0-5,07%	138,139
	Metropolitan District of Caracas	X	X	ND	59,5%	140

ND: no data; ID: intradomically; PD: peridomically.

mines by installing artificial lights in a model house in the northeastern plains of Colombia and concluded that this nocturnal dynamic poses a risk for the domestic introduction of discrete typing units (DTUs) of *T. cruzi* associated with sylvatic transmission foci.⁽¹³³⁾

Evidence of intradomiciliary and peridomiciliary colonisation of *P. geniculatus* in Latin America

Although *P. geniculatus* is registered as an intrusive wild species in some regions of the Americas, in other regions it is registered as a species capable of colonising human dwellings. These differences in the behavior of *P. geniculatus* probably reflect the genetic differences in this species across Latin American populations. Generally, the reported number of *P. geniculatus* nymphs associated with human dwellings varies, and Table IV lists nine publications that have reported on the presence of *P. geniculatus* eggs and nymphs. Furthermore, two intradomiciled adults and three peridomiciled nymphs in Muñecas, La Paz (Bolivia) were recorded.⁽⁴⁸⁾

Fifteen pigsties were examined in Muana, Pará State, Brazil, contained 207 females, 168 males, 2 N5, 7 NIII, 16 NII, 28 NI stages and 385 unhatched eggs, thereby evidencing a domiciliation process for *P. geniculatus*. In 10 of the 15 houses located up to a 10 m distant from the pigsties, 21 adults were collected in total.⁽¹³⁴⁾

The presence of *P. geniculatus* adults in 80 homes in Amalfi, Antioquia, Colombia, but only in one of the homes did they find nymphal stages of the insect was recorded.⁽⁴⁰⁾ Thirty-seven adults and a nymph of *P. geniculatus* in the intra-domicile of the studied dwellings in Atlántico, Colombia were reported.⁽¹³⁵⁾ The presence of adults and nymphs in development stages IV and V of *P. geniculatus* in the intra-domiciles in six of seven villages in the municipality of Tamara in Casanare Department (Colombia) was recorded.⁽¹³⁶⁾

Two males and one female *P. geniculatus*, 5NV, 8NIV, 3NIII, and 1NII stages, and seven eggs were captured in the tunnel of a rat (*Rattus rattus*) inside a house in Miranda, Venezuela. All the triatomines were abundantly blood-fed, with the exception of one recently molted male and one female, whose emergences were confirmed by the presence of NV-stage exuviae. Fecal examination of the three NV-stages and an adult male revealed the presence of *T. cruzi*.⁽¹³⁷⁾ In three houses examined in El Guamito, Lara State, Venezuela, were found one female, six N5-, two NIII- and two N2-stages and an overall *T. cruzi* infection rate of 9%.⁽¹³⁸⁾ In another study in the same state, *P. geniculatus* was found in 11 homes, registering 136 adults and two NIII stages, with a colonisation index of 18.18% and a 5.07% infection rate for *T. cruzi*.⁽¹³⁹⁾ 3390 adult *P. geniculatus* specimens, 26 NV, 71 NIV and 27 NIII stages were captured in the Metropolitan District of Caracas between 2007 and 2013, of which 59,5% carried *T. cruzi*. The authors warned that dramatic modifications to the surrounding natural habitats had led to the establishment of a *T. cruzi* urban enzootic cycle, resulting in a high risk of transmission of Chagas disease in this capital city.⁽¹⁴⁰⁾

T. cruzi isolates from *P. geniculatus*, *R. rattus* and from humans infected during an orally-acquired Chagas disease outbreak in a non-endemic region of Venezuela

belong to a same *T. cruzi* I genotype (TcI). The similarity in the parasite isolates from the three patients affected by the urban oral outbreak, the triatomine, and the rat captured at the guava juice preparation site allowed the authors to suggest that the source of infection was common among these persons and the vector, providing indirect evidence of *P. geniculatus* domiciliation in this region.⁽¹⁴¹⁾

The colonisation capacity of *P. geniculatus* has been studied under laboratory conditions. One such study reported a ro/b index of 0.77, but concluded that as a "K" strategist with a low reproduction rate and a prolonged generational time, it was unlikely that *P. geniculatus* was an important disease vector at the household level, a finding that does not presently reflect the epidemiological importance of the species.⁽⁴¹⁾ Nevertheless, another study compared the results they obtained from *P. geniculatus* (ro/b = 0.74) with two species known for their high colonising capacity, *Rhodnius prolixus* (0.74) and *Triatoma infestans* (0.65), and concluded that *P. geniculatus* has a remarkable colonising capacity, which should alert the relevant institutions to the risk presented by this species and the need to design new control strategies aimed at the wild species that in recent decades have been increasingly reported as having home intrusion and colonisation behavior.⁽⁴⁴⁾

The presence of sexual dimorphism was an indicator of home adaptation and the authors concluded that populations of *P. geniculatus* from Caracas, Venezuela, are morphologically adapted to home environments.⁽²⁰⁾ That the ability of the species to adapt to different environments in a few generations as was recently confirmed by Nakad-Bechara et al.⁽⁹⁰⁾ and poses a future risk to humans and should alert health institutions, especially taking into account the possible new scenarios of climate change and the specific anthropic processes of deforestation and landscape fragmentation.

P. geniculatus and oral transmission of *T. cruzi*

Over the last two decades, the oral form of disease transmission has changed from being a rare and unusual event to a public health concern after causing numerous deaths.^(142,143)

The difficulty in obtaining direct evidence incriminating the vector species responsible for the oral transmission of *T. cruzi*, is related to the fact that oral Chagas disease outbreaks occur several days after food contamination. Despite this difficulty, in some reported cases strong suspicions exist about the role of *P. geniculatus* in this transmission mechanism, because it has been observed as an abundant species in the area of the outbreak and it has been found invading the houses where contaminated food was prepared and, although in some cases the captured insects did not carry *T. cruzi* parasites, in other cases high levels of natural infection were observed. Table V lists 11 publications that report indirect evidence involving *P. geniculatus* in the oral transmission of *T. cruzi*.

Venezuela has experienced the most numerous and acute cases of parasite transmission by the oral route, with the source of infection generally being the intake of juices contaminated with triatomine feces. Infections have been

TABLE V
Cases where *Panstrongylus geniculatus* has been reported as a highly probable transmitter of oral *Trypanosoma cruzi* infection in Latin America

Country	Location	Contaminated food	Cases/deaths	Reference
Colombia	Restrepo (Meta)	Arepas and/or pineapple juice	4/0	113
	Antioquia (Turbo)	ND	11/1	144
	Santander (Bucaramanga)	tangerine juice and/or orange juice	9/0	143
	Santander (Lebrija)	tangerine juice and/or orange juice	10/2	145
	Santander (Piedecuesta)	ND	5/0	146
	San Vicente de Chucuri	ND	3/0	143
Bolivia	Cesar (Aguachica)	ND	11/0	147
Bolivia	Beni (Guayaramerín)	majo juice	14/0	148
	Caracas	guava juice	103/1	149
Venezuela	Vargas	ND	85/4	150
	Mérida (Antonio Pinto Salinas)	ND	5/0	151

ND: no data.

reported in greater Caracas and the states of Falcon, Mérida, Tachira, and Vargas where *P. geniculatus* has been the species linked with most of the outbreaks. Because it is the most abundant species in these localities, it has high *T. cruzi* infection rates and has been observed to defecate while returning to its shelter after feeding.^(128,140,152)

In the municipality of Chacao in Caracas in 2007, 103 people from a school were infected with *T. cruzi*, and of those infected, 19% needed hospitalisation and one child died. The infection source was juice made in a house where *T. cruzi*-infected *P. geniculatus* insects were captured.⁽¹⁴⁹⁾ During 2009, another outbreak occurred in a school environment that affected 82 people, from which three children and one adult died. Again, the evidence from the case indicates that the infection source was juice contaminated with the feces of *P. geniculatus*.⁽¹⁵⁰⁾

In Colombia, several cases of infection by oral transmission have occurred in areas with low endemicity and presence of domiciled vectors. Thus, wild vectors such as *P. geniculatus* have become important in relation to this mechanism of infection and may be associated with outbreaks related to oral transmission in several Colombian municipalities.⁽¹⁵³⁾ In Lebrija, Santander, an acute case was presented, where ten infections and two deaths were reported.⁽¹⁴⁵⁾ A report on a new scenario for transmission in the department of Córdoba-Colombia in an area that was not considered to be a region at risk of *T. cruzi* transmission. This report was based on an acute case of the disease in which the patient did not present lesions on the skin or in the periocular region that indicated the bite of the insect.⁽⁹²⁾

The transmission sources for two oral outbreaks were identified, one of them in the Colombian town of Restrepo, Meta. By epidemiological analysis, genotyping and allele identification of seven *T. cruzi* microsatellites in the samples obtained from patients, as well as insects and reservoirs implicated in the outbreaks, they determined that this outbreak was caused by fecal con-

tamination of arepa and pineapple juices by *P. geniculatus*. Their distance analysis allowed them to observe the grouping of patients with insects rather than with the reservoirs. Their findings were further explained by recent housing construction in the outbreak area, which altered the natural habitat of *P. geniculatus* and caused its intrusion into homes.⁽¹¹³⁾

DTUs of *T. cruzi* detected in *P. geniculatus*

Natural infections of *T. cruzi* in *P. geniculatus* have been widely reported and quantified in different Latin American countries. Despite no clear trend, high rates of natural infection occur with this parasite, reaching 44% on average. Brazilian rates are 16.5%,⁽¹⁵⁴⁾ 50%,⁽¹⁵⁵⁾ 4.7%⁽¹⁵⁶⁾ and 27.3%.⁽¹⁵⁷⁾ The Trinidadian rate is 42.5%,⁽⁸⁰⁾ the Colombian rates are 50%,⁽⁴⁰⁾ 70.6%⁽⁹³⁾ and 58.3%,⁽¹¹⁸⁾ the Venezuelan rates are 20%⁽¹³⁸⁾ and 76.1%,⁽¹²⁸⁾ the Peruvian rate is 50%,⁽¹²⁰⁾ and the Bolivian rate is 62.5%.⁽⁵¹⁾

Currently, seven DTUs, TcI to TcVI and T.c.bat, are known to circulate in the vectors in different proportions.^(158,159) The *T. cruzi* DTUs in *P. geniculatus* correspond to six of the seven described DTUs (TcI, TcII, TcIII, TcIV, TcV and TcVI), the geographic distributions of which in America for *P. geniculatus* are shown in Fig. 5. In Bolivia, insects infected with TcI and TcIII have been reported inside homes.⁽⁵¹⁾ In Brazil, insects infected with TcII and TcIII have been identified.⁽¹⁵⁷⁾ In Colombia, the transmission dynamics of the primary and secondary vectors in six departments (Antioquia, Cesar, Guajira, Huila, Meta, and Norte de Santander) were studied and, with six circulating DTUs (TcI, TcII, TcIII, TcIV, TcV and TcVI). *P. geniculatus* was observed to be the species with the highest number of DTUs.^(93,160) While TcI, TcIII and TcIV genotypes were isolated from the insects collected in the Colombian department of Casanare,⁽¹³³⁾ insects from Peru, Ecuador and French Guyana were only infected with TcI.^(120,161) Last, TcI, TcIII and TcIV DTUs were identified in *P. geniculatus* collected from Venezuela.⁽¹⁴⁰⁾



Fig. 5: geographical distribution in America of *Trypanosoma cruzi* discrete typing units (DTUs) detected in *Panstrongylus geniculatus*.

Possible effects of climate change on *P. geniculatus*

Although controversy persists about the consequences of climate change on vector-transmitted infectious diseases, there is full consensus that the global temperatures are closely linked with the physiological processes of arthropod vectors.^(162,163) Indeed, it has been shown that temperature and relative humidity can affect a vector insect by changing its original geographical distribution, altering the duration of its life cycle, or modifying its vector capacity, all which have consequences for the prevalence of infectious diseases.⁽¹⁶⁴⁾

To date, several studies have aimed to establish the effect of environmental variables on triatomines; however, attention has been mainly directed to *T. infestans* and *R. prolixus*.^(165,166,167) No studies on *P. geniculatus* that assess the effect of climatic conditions on its life cycle or vector capacity have been reported under controlled laboratory conditions in the available databases. Only estimates of its potential distribution based on ecological niche modeling have been reported for some Latin American countries.

In Brazil, the geographic distribution of triatomine species in the Central West region was estimated and the climatic factors that influence their occurrence were analysed. The results indicate that among the variables analysed, temperature seasonality better explains the models of occurrence. The authors also reported that almost the entire region has the climatic conditions appropriate for at least one species and, according to the distribution maps, *P. geniculatus* was the most widespread species with the greatest potential for occurrence.⁽¹⁶⁸⁾ In Venezuela, the possible effects of climate change on the potential distribution of five species (*Eratyrus mucronatus*, *P. geniculatus*, *T. maculata*, *R. robustus*, and *R. prolixus*) were analysed. It was found that the variables best explaining the model were seasonal temperature (47.9%) and isometry (26.5%), and the highest relative vector competence values were those for *R. prolixus* (0.8) and *P. geniculatus* (0.1). How-

ever, they concluded that the possible future effects of climate change on the vulnerability of the Venezuelan *P. geniculatus* population show a slight downwards trend.⁽¹⁶⁹⁾

In Colombia, the geographical distribution of four species (*P. geniculatus*, *R. pallescens*, *R. prolixus*, and *T. maculata*) was estimated and identified a relationship between landscape structure and climate factors that influence their occurrence. According to the predictive maps on potential species distribution, the variables that best explain the model were altitude (26%) and precipitation seasonality (18%).⁽¹³⁾ In light of the potential future geographical distributions, expressed as the suitability of a climate niche, several papers agree that *P. geniculatus* has a wide geographic distribution and has a greater distribution potential in possible climate change scenarios.^(13,168,169)

We recently observed that when *P. geniculatus* colonies are kept at 30°C, they have higher mortality and lower fertility than those kept at 26 and 28°C (Unpublished observations). Thus, it seems possible that under natural conditions, this species probably moves to areas where temperatures and altitudes favor its survival.

In Conclusion

All 15 of the species in the *Panstrongylus* genus have been described using morphological characters. Some of them are phenotypically similar to *P. geniculatus*, but molecular methods have not yet been used to determine whether or not they are the same species.

Geometric morphometry studies have revealed a reduction in the size of the populations studied in the urban areas of Caracas (Venezuela), when compared with sylvatic populations. The size reduction has also been verified in laboratory-raised populations, when compared with sylvatic populations; hence, this size reduction is likely associated with domiciliation processes.

Life-cycle timing under laboratory conditions has been shown to be dependent on the experimental conditions employed, such as temperature, relative humidity

and the food source for the colonies. Depending on the management of these variables, very heterogeneous data have been recorded from 149.5–531 days to even up to two years. Other studies using temperature conditions of 21–26°C and relative humidity of 90–100% have reported life cycle durations of 269, 297, 275.8, 273 and 275.4 days (average, 278 days).

Although *P. geniculatus* has been found in 18 countries, the recently generated predictive maps can produce a robust summary of its distributions. They can map its spatial variation to identify regions where a high risk of vector transmission of *T. cruzi* and transmission through food contamination are likely.

Based on the karyotype characteristics of *P. geniculatus*, which has high chromosomal variability, *P. geniculatus* is proposed to be a complex of species containing at least two different ones. The use of *Cytb* as a molecular marker has revealed spatial and genetic patterns related to domiciliation processes. The phylogenetic reconstruction of *P. geniculatus* from Colombia and Venezuela, using 16S rRNA and *Cytb*, showed that it comprises four phylogenetic clades concordant with its geographical distribution and the Andes uplift. However, it will be necessary to use other molecular markers and genomic approaches to identify intraspecific variations related to domiciliation of this species, high infection rates with *T. cruzi* and incrimination in oral outbreaks of Chagas disease in Colombia and Venezuela.

P. geniculatus is considered to have sylvatic habits; however, in recent decades at least 45 cases of intrusion into homes by adult insects have been reported in Bolivia, Brazil, Colombia, Peru, Surinam and Venezuela. This new scenario implies an increased transmission risk of the disease in endemic and non-endemic areas, either by direct contact or by food contamination. Nevertheless, in some Latin American regions the presence of nymphs, eggs and exuviae is evidence of colonisation of human dwellings, as reported in nine publications.^(40,48,134,135,136,137,138,139,140) These differences in species behavior probably reflect genetic differences between the populations.

The difficulty in obtaining direct evidence incriminating which vector species is responsible for oral transmission of *T. cruzi*, is related to the fact that orally-transmitted outbreaks of Chagas disease occur several days after food contamination. Despite this difficulty, in some reported cases there are strong suspicions about the role of *P. geniculatus* in this transmission mechanism, because it has been abundantly observed in outbreak areas. Indeed, it has been found invading houses where contaminated food is prepared and although in some cases the captured insects did not show infection with *T. cruzi*, in other cases high levels of natural infection were observed. At least 10 publications present indirect evidence involving *P. geniculatus* in the oral transmission of *T. cruzi*.^(113,143,144,145,146,147,148,149,150,151)

Although it is not completely clear what the effect of climate change will be on the biology of *P. geniculatus*, it is known that the species has optimal development between 26 and 28°C and between 90–100% relative humidity. That colonies kept at 30°C have high

mortality and low fertility means that increases in temperature may affect the distribution of the species such that it could move altitudinally to find places with more favorable temperatures.

ACKNOWLEDGEMENTS

To Sandra Cheesman, PhD, from Edanz Group (<https://en-author-services.edanzgroup.com/ac>) and Jason Garry for editing a draft of this manuscript.

AUTHORS' CONTRIBUTION

RJV and GAV conducted the review and drafted an original version of the manuscript. All authors contributed equally and approved the final version of the manuscript. The authors declare that there are no conflicts of interest related to the content of this article.

REFERENCES

- WHO - World Health Organization. Chagas disease (American trypanosomiasis). (Updated: 2020 May 23; cited 2020 Jul 15). Available from: [https://www.who.int/news-room/fact-sheets/detail/chagas-disease-\(american-trypanosomiasis\)](https://www.who.int/news-room/fact-sheets/detail/chagas-disease-(american-trypanosomiasis)).
- Justi S, Galvão C. The evolutionary origin of diversity in Chagas disease vectors. Trends Parasitol. 2016; 33(1): 42–52. <http://dx.doi.org/10.1016/j.pt.2016.11.002>.
- Monteiro F, Weirauch C, Felix M, Lazoski C, Abad-Franch F. Evolution, systematics, and biogeography of the Triatominae, vectors of Chagas disease. Adv Parasitol. 2018; 99: 265–344. <https://doi.org/10.1016/bs.apar.2017.12.002>.
- Dorn P, Justi S, Dale C, Stevens L, Galvão C, Lima-Cordón R, et al. Description of *Triatoma mopan* sp. n. from a cave in Belize (Hemiptera, Reduviidae, Triatominae). ZooKeys. 2018; 775: 69–95. <http://dx.doi.org/10.3897/zookeys.775.22553>.
- Poinar Jr. A primitive triatomine bug, *Paleotriatoma metaxytaxis* gen. et sp. nov. (Hemiptera: Reduviidae: Triatominae), in mid-Cretaceous amber from northern Myanmar. Cretaceous Res. 2019; 93: 90–7. <https://doi.org/10.1016/j.cretres.2018.09.004>.
- Nascimento J, Da Rosa J, Salgado-Roa F, Hernández C, Pardo-Díaz C, Alevi K, et al. Taxonomical over splitting in the *Rhodnius prolixus* (Insecta: Hemiptera: Reduviidae) clade: Are *R. taquaruensis* (da Rosa et al., 2017) and *R. neglectus* (Lent, 1954) the same species? PLoS One. 2019; 14(2): e0211285. <https://doi.org/10.1371/journal.pone.0211285>.
- Jurberg J, Galvão C. Biology, ecology, and systematics of Triatominae (Heteroptera, Reduviidae), vectors of Chagas disease, and implications for human health. Neue. 2006; 50: 1096–116.
- Schofield C, Galvão C. Classification, evolution, and species groups within the Triatominae. Acta Trop. 2009; 110(2–3): 88–100. <https://doi.org/10.1016/j.actatropica.2009.01.010>.
- Lent H, Wygodzinsky P. Revision of the Triatominae (Hemiptera: Reduviidae), and their significance as vectors of Chagas disease. Bull Am Museum Nat Hist. 1979; 163(3): 123–520. <http://hdl.handle.net/2246/1282>.
- Galvão C, Carcavallo R, Rocha D, Jurberg J. A checklist of the current valid species of the subfamily Triatominae Jeannel, 1919 (Hemiptera, Reduviidae) and their geographical distribution, with nomenclatural and taxonomic notes. Zootaxa. 2003; 202(1): 1–36. <http://dx.doi.org/10.11646/zootaxa.202.1.1>.
- Zeledón R, Ugalde J, Paniagua L. Entomological and ecological aspects of six sylvatic species of Triatomines (Hemiptera, Reduviidae) from the collection of the National Biodiversity Institute of Costa Rica, Central América. Mem Inst Oswaldo Cruz. 2001; 96(6): 757–64. <https://doi.org/10.1590/s0074-02762001000600002>.

12. Carcavallo RU, Curto de Casas S, Sherlock I, Galíndez I, Juberg J, Galvão C, et al. Geographical distribution and alti-latitude dispersión. In RU Carcavallo RU, IG Girón, J Jurberg, editors. *Atlas of Chagas disease vectors in the Americas*. Rio de Janeiro: Fiocruz; 1998; p. 747-92.
13. Parra-Henao G, Suárez-Escudero L, González-Caro S. Potential distribution of Chagas disease vectors (Hemiptera, Reduviidae, Triatominae) in Colombia, based on ecological niche modeling. *J Trop Med*. 2016; 1439090: 1-10. <http://dx.doi.org/10.1155/2016/1439090>.
14. Ceccarelli S, Balsalobre A, Cano ME, Vicente ME, Rabinovich JE, Medone P, et al. Datos de ocurrencia de triatominos americanos del Laboratorio de Triatominos del CEPAVE (CONICET-UNLP). Versión 1.2. Centro de Estudios Parasitológicos y de Vectores (CEPAVE). 2020 [accessed via GBIF.org on 2021-01-09]. Available from: <https://doi.org/10.15468/fbywtn>.
15. Guhl F, Aguilera G, Pinto N, Vergara D. Actualización de la distribución geográfica y ecoepidemiología de la fauna de triatominos (Reduviidae: Triatominae) en Colombia. *Biomédica*. 2007; 27(1): 143-62.
16. Chávez J. Contribución al estudio de los triatominos del Perú: distribución geográfica, nomenclatura y notas taxonómicas. *An Fac Med*. 2006; 67(1): 65-76.
17. Patterson J, Barbosa S, Feliciangeli M. On the genus *Panstrongylus* Berg 1879: evolution, ecology and epidemiological significance. *Acta Trop*. 2009; 110(2-3): 187-99. <http://dx.doi.org/10.1016/j.actatropica.2008.09.008>.
18. Crossa R, Hernández M, Caraccio M, Rose V, Valente S, Valente V, et al. Chromosomal evolution trends of the genus *Panstrongylus* (Hemiptera, Reduviidae), vectors of Chagas disease. *Infect Genet Evol*. 2002; 2(1): 47-56. [https://doi.org/10.1016/S1567-1348\(02\)00063-1](https://doi.org/10.1016/S1567-1348(02)00063-1).
19. dos Santos C, Jurberg J, Galvão C, Rocha D. Morphometric study of the genus *Panstrongylus* Berg, 1879 (Hemiptera, Reduviidae, Triatominae). *Mem Inst Oswaldo Cruz*. 2003; 98(7): 939-44. <http://dx.doi.org/10.1590/S0074-02762003000700014>.
20. Aldana E, Heredia E, Avendaño F, Lizano E, Concepción J, Bonfante R, et al. Análisis morfométrico de *Panstrongylus geniculatus* de Caracas, Venezuela. *Biomédica*. 2011; 31(1): 108-17. <https://doi.org/10.7705/biomedica.v31i1.341>.
21. Marcilla A, Bargues M, Abad-Franch F, Panzera F, Carcavallo R, Noireau F, et al. Nuclear rDNA ITS-2 sequences reveal polyphyly of *Panstrongylus* species (Hemiptera: Reduviidae: Triatominae), vectors of *Trypanosoma cruzi*. *Infect Genet Evol*. 2002; 1(3): 225-35. [http://dx.doi.org/10.1016/S1567-1348\(02\)00029-1](http://dx.doi.org/10.1016/S1567-1348(02)00029-1).
22. Justi S, Russo C, Mallet J, Obara M, Galvão C. Molecular phylogeny of Triatomini (Hemiptera: Reduviidae: Triatominae). *Parasit Vectors*. 2014; 7(149): 1-12. <http://dx.doi.org/10.1186/1756-3305-7-149>.
23. Bérenger J, Blanchet D. A new species of the genus *Panstrongylus* from French Guiana (Heteroptera: Reduviidae: Triatominae). *Mem Inst Oswaldo Cruz*. 2007; 102(6): 733-6. <https://doi.org/10.1590/S0074-0276200700500088>.
24. Ayala L. Una nueva especie de *Panstrongylus* Berg de Venezuela (Hemiptera: Reduviidae, Triatominae). *Entomotropica*. 2009; 24(3): 105-9.
25. Valderrama A, Lizano E, Cabello D, Valera M. *Panstrongylus turpiali*, n. sp. (Hemiptera: Reduviidae: Triatominae) from Venezuela. *Caribb J Sci*. 1996; 32: 142-4.
26. Lent H. Novos sinônimos de duas espécies de Triatominae da Venezuela (Hemiptera, Reduviidae). *Entomol Vect*. 1997; 4: 67-70.
27. Ayala J. Presencia de *Panstrongylus Chinai* (Del ponte, 1929) en Venezuela, con notas aclaratorias sobre su sinonimia con *Panstrongylus Turpiali* (Heteroptera: Reduviidae: Triatominae). *Boletín de la SEA*. 2016; 59: 233-6.
28. Lent H, Jurberg J. Estudos morfológicos comparativos de *Panstrongylus geniculatus* (Latreille, 1811) e *Panstrongylus megistus* (Burmeister, 1835) e suas genitalia externas (Hemiptera, Reduviidae, Triatominae). *Rev Bras Biol*. 1968; 28(4): 499-520.
29. Galíndez I, Carcavallo R, Valderrama A. Erwinilas o cerdas interomatidiales en la subfamilia Triatominae (Hemiptera: Reduviidae). *Entomol Vect*. 1994; 1(3): 93-6.
30. Carcavallo RU, Galíndez Girón I, Catalá S, Jurberg J, Lent H, Galvão C, et al. Some anatomic structures studied with scanning electronic microscopy (SEM). In RU Carcavallo, I Galíndez Girón, J Jurberg, H Lent, editors. *Atlas of Chagas disease vectors in the Americas*. Rio de Janeiro: Fiocruz; 1998. p. 299-393.
31. Galíndez Girón I, Carcavallo RU, Jurberg J, Galvão C, Lent H, Barata J, et al. External morphology and anatomy. In RU Carcavallo, I Galíndez Girón, J Jurberg, H Lent, editors. *Atlas of Chagas disease vectors in the Americas*. Rio de Janeiro: Fiocruz; 1998. p. 53-84.
32. Wolff M, González C. Ciclo de vida de *Panstrongylus geniculatus* (Hemiptera: Reduviidae) en condiciones de laboratorio. *Caldasia*. 1998; 20(1): 75-7. <https://revistas.unal.edu.co/index.php/cal/article/view/17471>.
33. Soto-Vivas A. Clave pictórica de triatominos (Hemiptera: Triatominae) de Venezuela. *Bol Malariol Salud Amb*. 2009; 49(2): 259-74.
34. Avendaño-Rangel F, Sandoval C, Aldana E. Descripción de setas cuticulares externas de cabeza, tórax, patas, abdomen y genitales en cuatro especies de Triatominae. *Biomédica*. 2016; 36(3): 354-8. <http://dx.doi.org/10.7705/biomedica.v36i3.3122>.
35. Jaramillo N, Castillo D, Wolff M. Geometric morphometric differences between *Panstrongylus geniculatus* from field and laboratory. *Mem Inst Oswaldo Cruz*. 2002; 97(5): 667-73. <http://dx.doi.org/10.1590/S0074-02762002000500015>.
36. Jurberg J, Galvão C, Noireau F, Carcavallo R, Rocha D, Lent H. Uma iconográfica dos triatomíneos (Hemiptera: Reduviidae). *Entomol Vect*. 2004; 11(3): 457-94.
37. Lent H, Jurberg J. Observações sobre o ciclo evolutivo, em laboratorio, do *Panstrongylus geniculatus* (Latreille, 1811) (Hemiptera: Reduviidae: Triatominae). *An Acad Brasil Ciênc*. 1969; 41: 125-31.
38. Galíndez I. Colonización y ciclo de vida de *Panstrongylus geniculatus* (Latreille, 1811) (Hemiptera: Reduviidae: Triatominae). Universidad de Los Andes, Núcleo Rafael Rangel, Trujillo: 1990; 34 pp.
39. Galíndez Girón I, Torres A, Márquez J, Carcavallo R. Colonización y ciclo de vida de *Panstrongylus geniculatus* (Latreille, 1811) (Hemiptera, Reduviidae, Triatominae). *Entomol Vector*. 1997; 4(3): 83-94.
40. Wolff M, Castillo D. Evidencias de domesticación y aspectos biológicos de *Panstrongylus geniculatus* (Latreille, 1811) (Hemiptera: Reduviidae). *Acta Entomol Chile*. 2000; 24: 77-83.
41. Cabello D, Galíndez Girón I. Vital Statistics of *Panstrongylus geniculatus* (Latreille 1811) (Hemiptera: Reduviidae) under experimental conditions. *Mem Inst Oswaldo Cruz*. 1998; 93(2): 257-62. <http://dx.doi.org/10.1590/S0074-02761998000200024>.
42. Esteban L, Angulo V. Estudio comparativo de la ovipostura de una cepa de laboratorio y una de campo de *Panstrongylus geniculatus* alimentadas con *Gallus domesticus* y *Mus musculus* BALB/c en condiciones de laboratorio. XX Congreso Latinoamericano de Parasitología. *Biomédica*. 2011a; 31(3): 253.

43. Esteban L, Angulo V. Estandarización de las condiciones de ayuno y peso en ninfas de V instar de *Panstrongylus geniculatus* para su utilización en pruebas biológicas. XX Congreso Latinoamericano de Parasitología. Biomédica. 2011; 31(3): 252-3.
44. Rabinovich J, Feliciangeli M. Vital statistics of Triatominae (Hemiptera: Reduviidae) under laboratory conditions: IV. *Panstrongylus geniculatus*. J Med Entomol. 2015; 52(5): 797-805. <https://doi.org/10.1093/jme/tjv112>.
45. Mayer M, Pifano C, Medina R. Aspectos epidemiológicos de la enfermedad de Chagas en Venezuela: bases para una campaña de saneamiento aplicable a zonas endémicas del medio rural venezolano. XII Conferencia Sanitaria Panamericana. 1946. Available from: http://iris.paho.org/xmlui/bitstream/handle/123456789/28807/CSP12_C30.pdf?sequence=1.
46. Leite G, dos Santos C, Falqueto A. Insecta, Hemiptera, Reduviidae, *Panstrongylus geniculatus*: geographic distribution map. Checklist. 2007; 3(2): 147-52.
47. Ceccarelli S, Balsalobre A, Medone P, Cano M, Gonçalves R, Gurgel R, et al. DataTri, a database of American triatomine species occurrence. Scientific Data. 2018; 5: 180071. <https://doi.org/10.1038/sdata.2018.71>.
48. Depickère S, Durán P, López R, Chávez T. Presence of intradomicile colonies of the triatomine bug *Panstrongylus rufotuberculatus* in Muñecas, La Paz, Bolivia. Acta Trop. 2011; 117(2): 97-100. <https://doi.org/10.1016/j.actatropica.2010.10.005>.
49. Depickère S, Durán P, López R, Martínez E, Chávez T. After five years of chemical control: colonies of the triatomine *Eratyrus mucronatus* are still present in Bolivia. Acta Trop. 2012; 123(3): 234-8. <https://doi.org/10.1016/j.actatropica.2012.05.005>.
50. Waleckx E, Depickère S, Salas R, Aliaga C, Monje M, Calle H, et al. New discoveries of sylvatic *Triatoma infestans* (Hemiptera: Reduviidae) throughout the Bolivian Chaco. Am J Trop Med Hyg. 2012; 86(3): 455-8. <https://doi.org/10.4269/ajtmh.2012.11-0205>.
51. Rojas M, Pinazo M, García L, Arteaga M, Uriona L, Gamboa S, et al. *Trypanosoma cruzi* infected *Panstrongylus geniculatus* and *Rhodnius robustus* adults invade households in the Tropics of Cochabamba region of Bolivia. Parasit Vectors. 2016; 9(158): 1-6. <https://doi.org/10.1186/s13071-016-1445-1>.
52. de Almeida F. Triatomíneos da Amazônia: encontro de três espécies naturalmente infectadas por *Trypanosoma* semelhante ao *cruzi*, no Estado do Amazonas (Hemiptera, reduviidae). Acta Amaz. 1971; 1(1): 89-93. <https://doi.org/10.1590/1809-43921971011089>.
53. Miles M, de Souza A, Póvoa M. Chagas disease in the Amazon basin. III. Ecotypes of ten triatomine bug species (Hemiptera: Reduviidae) from vicinity of Belém, Pará State, Brazil. J Med Entomol. 1981; 18(4): 266-78. <https://doi.org/10.1093/jmedent/18.4.266>.
54. Bento D, Farias L, Godoy M, Araújo J. Epidemiologia da doença de chagas na zona rural do município de Teresina - Piauí, Brasil. Rev Soc Bras Med Trop. 1992; 25(1): 51-8. <https://doi.org/10.1590/S0037-86821992000100008>.
55. de Souza K, Bavia M, Días A, Guimarães I, Soares É, Nascimento M, et al. Spatial distribution of triatomines (Reduviidae: Triatominae) in urban areas of the city of Salvador, Bahia, Brazil. Geospatial Health. 2011; 5(2): 199-203. <https://doi.org/10.4081/gh.2011.172>.
56. Santos F, Barreto W, de Macedo G, da Silva J, das Chagas S, Garcia C, et al. The reservoir system for *Trypanosoma* (Kinetoplastida, Trypanosomatidae) species in large neotropical wetland. Acta Trop. 2019; 199: 105098. <https://doi.org/10.1016/j.actatropica.2019.105098>.
57. Ucrós H. Distribución de los Triatominae en Colombia. Rev Fac Med. 1960; 28(10-12): 181-9. Available from: <https://revistas.unal.edu.co/index.php/revfacmed/article/viewFile/22596/23457>.
58. D'Alessandro A, Barreto P, Saravia N, Barreto M. Epidemiology of *Trypanosoma cruzi* in the Oriental Plains of Colombia. Am J Trop Med Hyg. 1984; 33(6): 1084-95. <https://doi.org/10.4269/ajtmh.1984.33.1084>.
59. Barreto M, Burbano M, Barreto P. Nuevos registros de flebotominos (Diptera: Psychodidae) y triatominos (Hemiptera: Reduviidae) para Risaralda, Cauca y Valle del Cauca, Colombia. Colombia Med. 1997; 28(3): 116-22. <https://colombiamedica.univalle.edu.co/index.php/comedica/article/view/62>.
60. Parra G, Flórez M, Angulo V. Vigilancia de Triatominae (Hemiptera: Reduviidae). Bogotá: Sic Editorial Ltda; 2015; 127 pp.
61. Escalante M, Gómez D, Silvera L, Sánchez G, Vanegas J. Detection of high percentage of *Trypanosoma cruzi* infection, the etiologic agent of Chagas disease, in wild populations of Colombian Caribbean Triatomines. Acta Parasitol. 2015; 60(2): 315-21. <https://doi.org/10.1515/ap-2015-0044>.
62. Esteban L, Montes J, Angulo V. Diversidad de triatominae (Hemiptera: reduviidae) en Santander, Colombia: implicaciones epidemiológicas. Biomédica. 2017; 37(1): 42-52. <http://dx.doi.org/10.7705/biomedica.v37i1.3140>.
63. León C, Ortiz M, Tovar C, Negrete J, Arroyo E, González C. Detection of *Trypanosoma cruzi* strains circulating in Córdoba department (Colombia) isolated from triatomines (Hemiptera: Reduviidae) collected by the community. Biomédica. 2019; 39(2): 265-77. <https://doi.org/10.7705/biomedica.v39i2.3973>.
64. Ayala J. Los triatominos de Costa Rica. Archivos Entomológicos. 2017; 18: 189-215.
65. Espinoza L. Epidemiología de la enfermedad de Chagas en la República del Ecuador. Rev Ecuat Hig Med Trop. 1955; 12: 25-105.
66. Rodríguez J. Epidemiología de la enfermedad de Chagas en la República del Ecuador. Rev Goiana Medica. 1959; 5: 411-38.
67. Amunárriz M, Chico M, Guderian R. Chagas disease in Ecuador: a sylvatic focus in the Amazon region. Am J Trop Med Hyg. 1991; 44: 145-9.
68. Chico H, Sandoval C, Guevara E, Calvopiña H, Cooper P, Reed S, et al. Chagas disease in Ecuador: evidence for disease transmission in an indigenous population in the Amazon region. Mem Inst Oswaldo Cruz. 1997; 92(3): 317-20. <https://doi.org/10.1590/S0074-02761997000300002>.
69. Aguilar V, Abad-Franch F, Paucar C, Racines V. Epidemiology of Chagas disease in Ecuador. A brief review. Mem Inst Oswaldo Cruz. 1999; 94(1): 387-93. <https://doi.org/10.1590/S0074-02761999000700076>.
70. Chippaux J, Dedet J, Geoffroy G, Tavakilian G, Pajot F. La maladie de Chagas en Guyane Française. La nature et l'homme en Guyane. Cayenne: ORSTOM et Institut Pasteur; 1983; 32 pp. http://horizon.documentation.ird.fr/exl-doc/pleins_textes/divers11-11/15460.pdf.
71. Bérenger J, Pluot-Sigwalt D, Blanchet D, Aznar C. The Triatominae species of French Guiana (Heteroptera: Reduviidae). Mem Inst Oswaldo Cruz. 2009; 104(8): 1111-16. <https://doi.org/10.1590/S0074-02762009000800007>.
72. Maes J. Los triatominos (Heteroptera: Reduviidae) en Nicaragua. Rev Nica Ent. 1992; 21: 1-8.
73. Maes J, Ratcliffe B, Jameson M. Fauna entomológica de la reserva natural Bosawas, Nicaragua. XIII. *Panstrongylus rufotuberculatus*, un triatomino (Heteroptera-Reduviidae) nuevo para la fauna de Nicaragua. Rev Nica Ent. 1997; 41: 19-22.
74. Rodríguez I, Loaiza J. American trypanosomiasis, or Chagas disease, in Panama: a chronological synopsis of ecological and epidemiological research. Parasit Vectors. 2017; 10: 459. <https://doi.org/10.1186/s13071-017-2380-5>.

75. Guillén Z, Cáceres I, Elliot A, Ramírez J. Triatominos del norte peruano y su importancia como vectores de *Trypanosoma* spp. Rev Per Ent. 1988; 31: 25-30. <http://sisbib.unmsm.edu.pe/BVRevistas/entomologia/v31/pdf/a06v31.pdf>.
76. Cáceres A, Troyes L, González-Pérez A, Llontop E, Bonilla C, Murias E, et al. Enfermedad de Chagas en la región nororiental del Perú. I. Triatominos (Hemiptera Reduviidae) presentes en Cajamarca y Amazonas. Rev Peru Med Exp Salud Publica. 2002; 19(1): 17-23.
77. Náquira C, Cabrera R. Breve reseña histórica de la enfermedad de Chagas, a cien años de su descubrimiento y situación actual en el Perú. Rev Peru Med Exp Salud Publica. 2009; 26(4): 494-504.
78. Lent H. Triatomideos da Guiana holandesa. Redescrição de *Panstrongylus lignarius* (Walker, 1873). Mem Inst Oswaldo Cruz. 1943; 38(3): 485-96.
79. Hiwat H. Triatominae species of Suriname (Heteroptera: Reduviidae) and their role as vectors of Chagas disease. Mem Inst Oswaldo Cruz. 2014; 109(4): 452-8. <http://dx.doi.org/10.1590/0074-0276130408>.
80. Omah-Maharaj I. Studies on vectors of *Trypanosoma cruzi* in Trinidad, West Indies. Med Vet Entomol. 1992; 6(2): 115-20. <https://doi.org/10.1111/j.1365-2915.1992.tb00586.x>.
81. Noya-Alarcón O, Botto C, Cortez J, Ferrer E, Vietri M, Herrera L. Primer registro de *Panstrongylus geniculatus* (Latreille, 1811) en los municipios Alto Orinoco y Atures, estado Amazonas, Venezuela. Bol Malariol Salud Amb. 2011; 51(1): 81-5.
82. Carcavallo RU, Rodríguez M, Salvatella R, Curto de Casas S, Rocha D, Sherlock I, et al. Habitats and related fauna. In RU Carcavallo, I Galíndez Girón, J Jurberg, H Lent, editors. Atlas of Chagas disease vectors in the Americas. Rio de Janeiro: Fiocruz; 1998. p. 561-620.
83. Carcavallo RU, Rocha D, Galíndez Girón I, Sherlock I, Galvão C, Lent H, et al. Feeding sources and patterns. In RU Carcavallo, I Galíndez Girón, J Jurberg, H Lent, editors. Atlas of Chagas disease vectors in the Americas. Rio de Janeiro: Fiocruz; 1998. p. 537-60.
84. Reyes M. *Panstrongylus geniculatus* Latreille 1811 (Hemiptera: Reduviidae: Triatominae), vector de la enfermedad de Chagas en el ambiente domiciliario del centro-norte de Venezuela. Rev Biomed. 2009; 20: 180-205.
85. Abad-Franch F, Lima M, Sarquis O, Gurgel R, Sánchez M, Calzada J, et al. On palms, bugs, and Chagas disease in the Americas. Acta Trop. 2015; 151: 126-41. <http://dx.doi.org/10.1016/j.actatropica.2015.07.005>.
86. Garrouste R. La première observation in natura de l'entomophagie de *Panstrongylus geniculatus* (Latreille 1811) hématophage vecteur de la maladie de Chagas (Hemiptera: Reduviidae). Ann Soc Entomol Fr (NS). 2009; 45(3): 302-4. <https://doi.org/10.1080/0379271.2009.10697614>.
87. Bender A, Python A, Lindsay S, Golding N, Moyes C. Modeling geospatial distributions of the triatomine vectors of *Trypanosoma cruzi* in Latin America. PLoS Negl Trop Dis. 2020; 14(8): e0008411. <http://dx.doi.org/10.1371/journal.pntd.0008411>.
88. Barrett T. Advances in triatomine bug ecology in relation to Chagas disease. In K Harris, org. Advances in disease vector research. New York: Springer-Verlag; 1991. p. 143-76.
89. Jurberg J, Carcavallo R, Lent H. *Panstrongylus sherlocki* sp. n. do estado da Bahia, Brasil (Hemiptera, Reduviidae, Triatominae). Entomol Vect. 2001; 8(2): 261-74.
90. Nakad-Bechara C, Londoño J, Segovia M, León M, Martínez P, Rodríguez R, et al. Genetic variability of *Panstrongylus geniculatus* (Reduviidae: Triatominae) in the Metropolitan District of Caracas, Venezuela. Infect Genet Evol. 2018; 66: 236-44. <http://dx.doi.org/10.1016/j.meegid.2018.09.011>.
91. Caicedo-Garzón V, Salgado-Roa F, Sánchez-Herrera M, Hernández C, Arias-Giraldo L, García L, et al. Genetic diversification of *Panstrongylus geniculatus* (Reduviidae: Triatominae) in northern South America. PLoS One. 2019; 14(10): e0223963. <https://doi.org/10.1371/journal.pone.0223963>.
92. Tovar C, Negrete J, González C, León C, Ortiz M, Chacón J, et al. New scenarios of Chagas disease transmission in northern Colombia. J Parasitol Res. 2017; 3943215. <https://doi.org/10.1155/2017/3943215>.
93. Hernández C, Salazar C, Brochero H, Teherán A, Buitrago L, Vera M, et al. Untangling the transmission dynamics of primary and secondary vectors of *Trypanosoma cruzi* in Colombia: parasite infection, feeding sources and discrete typing units. Parasit Vectors. 2016; 9(1): 620. <https://doi.org/10.1186/s13071-016-1907-5>.
94. Brito R, Gorla D, Diotaiuti L, Gomes A, Souza R, Abad-Franch F. Drivers of house invasion by sylvatic Chagas disease vectors in the Amazon-Cerrado transition: A multi-year, state-wide assessment of municipality-aggregated surveillance data. PLoS Negl Trop Dis. 2017; 11(11): e0006035. <https://doi.org/10.1371/journal.pntd.0006035>.
95. Damborsky M, Bar M, Oscherov E. Detecção de triatominos (Hemiptera: Reduviidae) en ambientes domésticos y extradomésticos, Corrientes, Argentina. Cad Saude Publica. 2001; 17(4): 843-9. <https://doi.org/10.1590/S0102-311X2001000400018>.
96. Barreto M. Estudos sobre reservatórios e vetores silvestres do *Trypanosoma cruzi*. XXI. Observações sobre a ecologia do *Panstrongylus geniculatus* (Latreille, 1811). (Hemiptera, Reduviidae). Rev Bras Biol. 1967; 27(4): 337-48.
97. de Oliveira A, da Silva I. Distribuição geográfica e indicadores entomológicos de triatomíneos sinantrópicos capturados no Estado de Goiás. Rev Soc Bras Med Trop. 2007; 40(2): 204-8. <https://doi.org/10.1590/S0037-86b2007000200011>.
98. Almeida P, Santos H, Barata J, Obara M, Cerretti W. Ocorrência de *Panstrongylus guentheri* Berg (Hemiptera: Reduviidae) no Mato Grosso do Sul. Neotrop Entomol. 2008; 37(1): 107-8. <https://doi.org/10.1590/S1519-566X2008000100019>.
99. Loruso E, Santos C, Jurberg J. Foco de doença de Chagas em São Félix, no estado do Rio de Janeiro. Rev Soc Bras Med Trop. 2008; 41(4): 419-20. <https://www.arca.fiocruz.br/handle/icict/27550>.
100. Peixoto R, Rocha D, Dale C, Galvão C. *Panstrongylus geniculatus* (Latreille, 1811) (Hemiptera, Reduviidae, Triatominae): first record on Ilha Grande, Rio de Janeiro, Brazil. CheckList. 2020; 16(2): 391-4. <https://doi.org/10.15560/16.2.391>.
101. Naiff M, Naiff R, Barret T. Vetores selváticos de doença de Chagas na área urbana de Manaus (AM): atividade de vôo nas estações secas e chuvosas. Rev Soc Bras Med Trop. 1998; 31(1): 103-5. <https://doi.org/10.1590/S0037-86821998000100014>.
102. Fé N, Magalhães L, Fé F, Arakian S, Monteiro W, Barbosa M. Ocorrência de triatomíneos em ambientes silvestres e domiciliares do município de Manaus, Estado do Amazonas. Rev Soc Bras Med Trop. 2009; 42(6): 642-6. <http://dx.doi.org/10.1590/S0037-86822009000600006>.
103. Batista D, Britto C, Monte G, Baccaro F. Occurrence of triatomines (Hemiptera: Reduviidae) in domestic and natural environments in Novo Remanso, Itacoatiara, Amazonas, Brazil. J Braz Soc Trop Med. 2019; 52: e20190063. <http://dx.doi.org/10.1590/0037-8682-0063-2019>.
104. de Assis G, Azeredo B, Gorla D, Diotaiuti L, Lana M. Entomological surveillance of Chagas disease in Berilo municipality, Jequitinhonha Valley, State of Minas Gerais, Brazil. Rev Soc Bras Med Trop. 2009; 42(6): 615-21. <https://doi.org/10.1590/S0037-86822009000600001>.
105. de Paula M, da Costa I, Freitas P, Limongi J, Neto A, Pinto R, et al. Occurrence of positivity for *Trypanosoma cruzi* in triato-

- mine from municipalities in Southeastern Brazil, from 2002 to 2004. Rev Soc Bras Med Trop. 2010; 43(1): 9-14. <http://dx.doi.org/10.1590/S0037-86822010000100003>.
106. Dias J, Queiroz D, Martins H, Gorla D, Pires H, Diotaiuti L. Spatial distribution of triatomines in domiciles of an urban area of the Brazilian Southeast Region. Mem Inst Oswaldo Cruz. 2016; 111(1): 43-50. <https://dx.doi.org/10.1590/0074-02760150352>.
107. Maeda M, Knox M, Gurgel-Gonçalves R. Occurrence of synanthropic triatomines (Hemiptera: Reduviidae) in the Federal District of Brazil. Rev Soc Bras Med Trop. 2012; 45(1): 71-6. <http://dx.doi.org/10.1590/S0037-86822012000100014>.
108. Minuzzi-Souza T, Nitz N, Cuba-Cuba C, Santalucia M, Knox M, Hagström L, et al. Synanthropic triatomines as potential vectors of *Trypanosoma cruzi* in central Brazil. Rev Soc Bras Med Trop. 2017; 50(6): 824-8. <https://doi.org/10.1590/0037-8682-0199-2017>.
109. Barreto M. Estudos sobre reservatórios e vetores silvestres do *Trypanosoma cruzi*. XXI. Observações sobre a ecología do *Panstrongylus geniculatus* (Latreille, 1811). (Hemiptera, Reduviidae). Rev Bras Biol. 1967; 27(4): 337-8.
110. Ceretti-Junior W, Vendrami D, de Matos-Junior M, Rimoldi-Ribeiro A, Alvarez J, Marques S, et al. Occurrences of triatomines (Hemiptera: Reduviidae) and first reports of *Panstrongylus geniculatus* in urban environments in the city of São Paulo, Brazil. Rev Inst Med Trop São Paulo. 2018; 60: e33. <http://dx.doi.org/10.1590/S1678-9946201860033>.
111. Ribeiro M, Castro G, de Souza J, da Rosa J, Camargo L, Meneguetti D. Occurrence of triatomines in an urban residential complex in the municipality of Rio Branco, Acre, South-Western Amazon. Rev Soc Bras Med Trop. 2019; 52: e20180177 <http://dx.doi.org/10.1590/0037-8682-0177-2018>.
112. Cortés L, Suárez H. Triatominos (Reduviidae: Triatominae) en un foco de enfermedad de Chagas en Talaigua Nuevo (Bolívar, Colombia). Biomédica. 2005; 25(4): 568-74. <https://doi.org/10.7705/biomedica.v25i4.1383>.
113. Hernández C, Vera M, Cucunubá Z, Flórez C, Cantillo O, Buitrago L, et al. High-Resolution molecular typing of *Trypanosoma cruzi* in 2 large outbreaks of acute Chagas disease in Colombia. J Infect Dis. 2016; 214(8): 1252-5. <https://doi.org/10.1093/infdis/jiw360>.
114. Dib J, Barnabe C, Tibayrenc M, Triana O. Incrimination of *Eratyrs cuspidatus* (Stål) in the transmission of Chagas disease by molecular epidemiology analysis of *Trypanosoma cruzi* isolates from a geographically restricted area in the north of Colombia. Acta Trop. 2009; 111(3): 237-42. <https://doi.org/10.1016/j.actatropica.2009.05.004>.
115. Angulo V, Esteban L, Luna K. *Attalea butyracea* próximas a las viviendas como posible fuente de infestación domiciliaria por *Rhodnius prolixus* (Hemiptera: Reduviidae) en los Llanos Orientales de Colombia. Biomédica. 2012; 32(2): 1-26. <http://doi.org/10.7705/biomedica.v32i2.430>.
116. León C, Ortiz M, Tovar C, Negrete J, Arroyo E, González C. Detection of *Trypanosoma cruzi* strains circulating in Cordoba department (Colombia) isolated from triatomines (Hemiptera: Reduviidae) collected by the community. Biomédica. 2019; 39(2): 265-77. <https://doi.org/10.7705/biomedica.v39i2.3973>.
117. Reyes M, Torres A, Esteban L, Flórez M, Angulo V. Riesgo de transmisión de la enfermedad de Chagas por intrusión de triatominos y mamíferos silvestres en Bucaramanga, Santander, Colombia. Biomédica. 2017; 37(1): 68-78. <http://dx.doi.org/10.7705/biomedica.v37i1.3051>.
118. Ayala C, Hernández C, Eyes M, Romero L, Álvarez R, Blanco P. Detección de infección natural por *Trypanosoma cruzi* (Trypanosomatidae) en triatominos del municipio de Colosó, Colombia. Acta Biol Col. 2019; 24(1): 180-4. <http://dx.doi.org/10.15446/abc.v24n1.72306>.
119. Vega S, Mendoza A, Cabrera R, Cáceres A, Campos E, Ancca J, et al. Primer caso de enfermedad de Chagas aguda en la Selva Central del Perú: investigación de colaterales, vectores y reservorios. Rev Peru Med Exp Salud Publica. 2006; 23(4): 288-92.
120. Padilla C, Alvarado U, Ventura G, Luna-Caipo D, Suárez M, Tuñóque J, et al. Detección de unidades discretas de tipificación de *Trypanosoma cruzi* en triatominos recolectados en diferentes regiones naturales de Perú. Biomédica. 2017; 37(2): 167-79. <https://doi.org/10.7705/biomedica.v37i0.3559>.
121. Torres D, Cabrera R. Geographical distribution and intra-domiciliary capture of sylvatic triatomines in La Convención province, Cusco, Peru. Rev Inst Med Trop São Paulo. 2010; 52(3): 157-60. <https://doi.org/10.1590/S0036-46652010000300008>.
122. Cabrera R, Vega S, Cáceres A, Ramal C, Álvarez C, Ladera P, et al. Epidemiological investigation of an acute case of Chagas disease in an area of active transmission in Peruvian Amazon region. Rev Inst Med Trop São Paulo. 2010; 52(5): 269-72. <http://dx.doi.org/10.1590/S0036-46652010000500009>.
123. Cáceres A, Vega S, Ancca J, Pinto J, Vela G, Cárdenas V, et al. Aspectos entomológicos de la enfermedad de Chagas en Huallaga y Picota, San Martín, Perú. An Fac Med. 2010; 71(1): 28-36.
124. Hiwat H. Triatominae species of Suriname (Heteroptera: Reduviidae) and their role as vectors of Chagas disease. Mem Inst Oswaldo Cruz. 2014; 109(4): 452-8. <http://dx.doi.org/10.1590/0074-0276130408>.
125. Pifano F. El potencial enzooótico silvestre del complejo ecológico *Schizotrypanum cruzi* - *Didelphis marsupialis* - *Panstrongylus geniculatus* y sus incursiones a la vivienda humana del valle de Caracas - Venezuela. Bol Acad Cienc Fis Mat Nat. 1986; 46: 143-4.
126. Londoño J, Briceño L, Carrasco H, Rodríguez M. Riesgo de transmisión urbana de *Trypanosoma cruzi* por *Panstrongylus geniculatus*. XV Congreso Colombiano de Parasitología y Medicina Tropical, Colombia. Biomédica. 2011; 31(3): 51-2.
127. Añez N, Saavedra C, Crisante G, Rojas A, Lizano E. Infección natural por *Trypanosoma cruzi* en *Panstrongylus geniculatus* (Latreille, 1811) de la región montaña de Mérida, Venezuela. Bol Malaria Salud Amb. 2005; 45(2): 139-41. <http://www.saber.ulav.edu/handle/123456789/29808>.
128. Carrasco H, Torrellas A, García C, Segovia M, Feliciangeli M. Risk of *Trypanosoma cruzi* I (Kinetoplastida: Trypanosomatidae) transmission by *Panstrongylus geniculatus* (Hemiptera: Reduviidae) in Caracas (Metropolitan District) and neighboring States, Venezuela. Int J Parasitol. 2005; 35(13): 1379-84. <http://doi.org/10.1016/j.ijpara.2005.05.003>.
129. Rojas M, Várquez P, Villarreal M, Velandia C, Vergara L, Morán-Borges Y, et al. Estudio seroepidemiológico y entomológico sobre la enfermedad de Chagas en un área infestada por *Triatoma maculata* (Erichson 1848) en el centro-occidente de Venezuela. Cad Saude Pública. 2008; 24(10): 2323-33. <https://doi.org/10.1590/S0102-311X2008001000013>.
130. Serrano O, Mendoza F, Suárez B, Soto A. Seroepidemiología de la enfermedad de Chagas en dos localidades del municipio Costa de Oro, estado Aragua, Venezuela. Biomédica. 2008; 28(1): 108-15. <https://doi.org/10.7705/biomedica.v28i1.113>.
131. García-Jordán N, Berrizbeitia M, Concepción J, Aldana E, Cáceres A, Quiñones W. Estudio entomológico de vectores transmisores de la infección por *Trypanosoma cruzi* en la población rural del estado Sucre, Venezuela. Biomédica. 2015; 35(2): 247-57. <http://dx.doi.org/10.7705/biomedica.v35i2.2390>.
132. Castro M, Barrett T, Santos W, Abad-Franch F, Rafael J. Attraction of Chagas disease vectors (Triatominae) to artificial light sources in the canopy of primary Amazon rainforest. Mem Inst Oswaldo Cruz. 2010; 105(8): 1061-4. <http://dx.doi.org/10.1590/S0074-02762010000800019>.

133. Jácome-Pinilla D, Hincapié-Peñaloza E, Ortiz M, Ramírez J, Guhl F, Molina J. Risks associated with dispersive nocturnal flights of sylvatic Triatominae to artificial lights in a model house in the northeastern plains of Colombia. *Parasit Vectors*. 2015; 8: 600. <https://dx.doi.org/10.1186/s13071-015-1209-3>.
134. Valente V, Valente S, Noireau F, Carrasco H, Miles M. Chagas disease in the Amazon Basin: Association of *Panstrongylus geniculatus* (Hemiptera: Reduviidae) with domestic pigs. *J Med Entomol*. 1998; 35(2): 99-103. <https://doi.org/10.1093/jmedent/35.2.99>.
135. Maestre-Serrano R, Eyes-Escalante M. Actualización de la presencia y distribución de triatominos en el departamento del Atlántico-Colombia: 2003-2010. *Bol Malariol Salud Amb*. 2012; 52(1): 125-8.
136. Montenegro D, Vera M, Zuleta L, Llanos I, Junqueira A. Estrategia para determinar la línea base en áreas de interrupción vectorial de la enfermedad de Chagas. *Rev Panam Salud Publica*. 2016; 39(6): 341-51.
137. Reyes M, Rodríguez A. Domiciliation of selvatic Chagas disease vector *Panstrongylus geniculatus* (Latreille, 1811) (Triatominae: Reduviidae) in Venezuela. *Trans R Soc Trop Med Hyg*. 2000; 94(5): 508. [https://doi.org/10.1016/s0035-9203\(00\)90068-3](https://doi.org/10.1016/s0035-9203(00)90068-3).
138. Feliciangeli M, Carrasco H, Patterson J, Suárez B, Martínez C, Medina M. Mixed domestic infestation by *Rhodnius prolixus* Stål, 1859 and *Panstrongylus geniculatus* (Latreille, 1811), vector incrimination, and seroprevalence for *Trypanosoma cruzi* among inhabitants in El Guamito, Lara state, Venezuela. *Am J Trop Med Hyg*. 2004; 71(4): 501-5. <https://doi.org/10.4269/ajtmh.2004.71.501>.
139. Rodríguez-Bonfante C, Amaro A, García M, Wohlert L, Guillén P, García R, et al. Epidemiología de la enfermedad de Chagas en el municipio Andrés Eloy Blanco, Lara, Venezuela: infestación triatomínica y seroprevalencia en humanos. *Cad Salud Pública*. 2007; 23(5): 1133-40. <https://doi.org/10.1590/S0102-311X2007000500015>.
140. Carrasco H, Segovia M, Londoño J, Ortegoza J, Rodríguez M, Martínez C. *Panstrongylus geniculatus* and four other species of triatomine bug involved in the *Trypanosoma cruzi* enzootic cycle: high risk factors for Chagas disease transmission in the Metropolitan District of Caracas, Venezuela. *Parasit Vectors*. 2014; 7(602): 1-5. <https://doi.org/10.1186/s13071-014-0602-7>.
141. Díaz-Bello Z, Thomas M, López M, Zavala-Jasper R, Noya O, Alarcón de Noya B, et al. *Trypanosoma cruzi* genotyping supports a common source of infection in a school-related oral outbreak of acute Chagas disease in Venezuela. *Epidemiol Infect*. 2014; 142(1): 156-62. <https://doi.org/10.1017/S0950268813000757>.
142. Nicholls R, Cucunubá Z, Knudson A, Flórez A, Montilla M, Puerta C, et al. Enfermedad de Chagas aguda en Colombia, una entidad poco sospechada. Informe de 10 casos presentados en el periodo 2002 a 2005. *Biomédica*. 2007; 27(1): 8-17. <https://doi.org/10.7705/biomedica.v27i1.244>.
143. Rueda K, Trujillo J, Carranza J, Vallejo G. Transmisión oral de *Trypanosoma cruzi*: una nueva situación epidemiológica de la enfermedad de Chagas en Colombia y otros países suramericanos. *Biomédica*. 2014; 34(4): 631-41. <https://doi.org/10.7705/biomedica.v34i4.2204>.
144. Ríos J, Arboleda M, Montoya A, Alarcón E, Parra-Henao G. Probable brote de transmisión oral de enfermedad de Chagas en Turbo, Antioquia. *Biomédica*. 2011; 31(2): 185-195. <https://doi.org/10.7705/biomedica.v31i2.302>.
145. Hernández L, Ramírez A, Cucunubá Z, Zambrano P. Brote de Chagas agudo en Lebrija, Santander, 2008. *Revista del Observatorio de Salud Pública de Santander*. 2009; 4(1): 28-36.
146. Zambrano P, Cucunubá Z, Montilla M, Flórez C, Parra E, Cortés L. Brotes de síndrome febril asociado a miocarditis aguda chagásica de posible transmisión oral en el departamento de Santander, diciembre de 2008 a mayo de 2009. *Inf Quinc Epidemiol Nac*. 2008; 15: 17-32.
147. Soto H, Tibaduiza T, Montilla M, Triana O, Suárez D, Torres M, et al. Investigación de vectores y reservorios en brote de Chagas agudo por posible transmisión oral en Aguachica, Cesar, Colombia. *Cad Saude Pública*. 2014; 30(4): 746-56. <https://doi.org/10.1590/0102-311X00024013>.
148. Santalla J, Oporto P, Espinoza E, Ríos T, Brutus L. Primer brote reportado de la enfermedad de Chagas en la Amazonía Boliviana: reporte de 14 casos agudos por transmisión oral de *Trypanosoma cruzi* en Guayaramerín, Beni-Bolivia. *Biofarbo*. 2011; 19(1): 52-8.
149. Alarcón de Noya B, Díaz-Bello Z, Colmenares C, Ruiz-Guevara R, Mauriello L, Zavala-Jaspe R, et al. Large urban outbreak of orally acquired acute Chagas disease at a school in Caracas, Venezuela. *J Infect Dis*. 2010; 201(9): 1308-15. <http://dx.doi.org/10.1086/651608>.
150. Suárez J, de Suárez C, Alarcón de Noya B, Espinosa R, Chiurillo M, Villaroel A, et al. Enfermedad de Chagas sistémico en fase aguda por transmisión oral: diagnóstico integral de un caso autopsiado. *Gac Med Caracas*. 2010; 118(3): 212-22.
151. Añez N, Crisante G, Rojas A, Dávila D. Brote de enfermedad de Chagas agudo de posible transmisión oral en Mérida, Venezuela. *Bol Malariol Salud Amb*. 2013; 53(1): 1-10.
152. Alarcón de Noya B, Díaz-Bello Z, Colmenares C, Ruiz-Guevara R, Mauriello L, Muñoz A, et al. Update on oral chagas disease outbreaks in Venezuela: Epidemiological, clinical and diagnostic approaches. *Mem Inst Oswaldo Cruz*. 2015; 110(3): 377-86. <http://dx.doi.org/10.1590/0074-02760140285>.
153. Ramírez J, Montilla M, Cucunubá Z, Floréz A, Zambrano P, Guhl F. Molecular epidemiology of human oral Chagas disease outbreaks in Colombia. *PLoS Negl Trop Dis*. 2013; 7(2): e2041. <http://dx.doi.org/10.1371/journal.pntd.0002041>.
154. Valente V. Potential for domestication of *Panstrongylus geniculatus* (Latreille, 1811) (Lepidoptera, Reduviidae, Triatominae) in the municipality of Muaná, Marajó Island, State of Pará, Brazil. *Mem Inst Oswaldo Cruz*. 1999; 94(1): 399-400. <http://dx.doi.org/10.1590/S0074-02761999000700078>.
155. Dario M, Lisboa C, Costa L, Moratelli R, Nascimento M, Costa L, et al. High *Trypanosoma spp.* diversity is maintained by bats and triatomines in Espírito Santo state, Brazil. *PLoS One*. 2017; 12(11): e0188412. <https://doi.org/10.1371/journal.pone.0188412>.
156. Ferro A, Sobral-Souza T, Vancine M, Muylaert R, de Abreu A, Pelloso S, et al. Spatial prediction of risk areas for vector transmission of *Trypanosoma cruzi* in the State of Paraná, southern Brazil. *PLoS Negl Trop Dis*. 2018; 12(10): e0006907. <https://doi.org/10.1371/journal.pntd.0006907>.
157. Dario M, Andrade T, dos Santos C, Fux B, Brandão A, Falqueto A. Molecular characterization of *Trypanosoma cruzi* samples derived from *Triatoma vitticeps* and *Panstrongylus geniculatus* of the Atlantic rainforest, southeast Brazil. *Parasite*. 2018; 25: 59. <https://doi.org/10.1051/parasite/2018060>.
158. Zingales B, Andrade S, Briones M, Campbell D, Chiari E, Fernandes O, et al. A new consensus for *Trypanosoma cruzi* intraspecific nomenclature: second revision meeting recommends TcI to TcVI. *Mem Inst Oswaldo Cruz*. 2009; 104(7): 1051-4. <http://dx.doi.org/10.1590/s0074-02762009000700021>.
159. Marcili A, Lima L, Cavazzana Jr, Junqueira A, Veludo H, Da Silva F, et al. A new genotype of *Trypanosoma cruzi* associated with bats evidenced by phylogenetic analyses using SSU rDNA, cytochrome b and Histone H2B genes and genotyping based on ITS1 rDNA. *Parasitology*. 2009; 136(6): 641-55. <https://doi.org/10.1017/S0031182009005861>.
160. Messenger LA, Ramírez JD, Llewellyn MS, Guhl F, Miles MA. Importation of hybrid human-associated *Trypanosoma cruzi* strains of southern South American origin, Colombia. *Emerg Infect Dis*. 2016; 22(8): 1452-5.

161. Barnabé C, Brisse S, Tibayrenc M. Population structure and genetic typing of *Trypanosoma cruzi*, the agent of Chagas disease: a multilocus enzyme electrophoresis approach. Parasitology. 2000; 120(5): 513-26. <https://doi.org/10.1017/s0031182099005661>.
162. Liang L, Gong P. Climate change and human infectious diseases: a synthesis of research findings from global and spatio-temporal perspectives. Environ Int. 2017; 103: 99-108. <https://doi.org/10.1016/j.envint.2017.03.011>.
163. Laferty K. The ecology of climate change and infectious diseases. Ecology. 2009; 90(4): 888-900. <https://doi.org/10.1890/08-0079.1>.
164. Gage K, Burkot T, Eisen R, Hayes E. Climate and vector-borne diseases. Am J Prev Med. 2008; 35(5): 436-50. <https://doi.org/10.1016/j.amepre.2008.08.030>.
165. Roca M, Lazzari C. Effects of relative humidity on the hematophagous bug *Triatoma infestans*: hygropreference and eclosion success. J Insect Phys. 1994; 40(10): 901-7. [https://doi.org/10.1016/0022-1910\(94\)90024-8](https://doi.org/10.1016/0022-1910(94)90024-8).
166. Luz C, Fargues J, Grunewald J. Development of *Rhodnius prolixus* (Hemiptera: Reduviidae) under constant and cyclic conditions of temperature and humidity. Mem Inst Oswaldo Cruz. 1999; 94(3): 403-9. <http://dx.doi.org/10.1590/S0074-02761999000300022>.
167. Tamayo L, Guhl F, Vallejo G, Ramírez J. The effect of temperature increase on the development of *Rhodnius prolixus* and the course of *Trypanosoma cruzi* metacyclogenesis. PLoS Negl Trop Dis. 2018; 12(8): e0006735. <https://doi.org/10.1371/journal.pntd.0006735>.
168. Pereira J, Almeida P, Sousa A, Paula A, Machado R, Gurgel-Gonçalves R. Climatic factors influencing triatomine occurrence in Central-West Brazil. Mem Inst Oswaldo Cruz. 2013; 108(3): 335-41. <http://dx.doi.org/10.1590/S0074-02762013000300012>.
169. Ceccarelli S, Rabinovich J. Global climate change effects on Venezuela's vulnerability to Chagas disease is linked to the geographic distribution of five Triatomine species. J Med Entomol. 2015; 52(6): 1333-43. <https://doi.org/10.1093/jme/tjv119>.