

Characterization of *Rhodnius neglectus* from Two Regions of Brazil Using Isoenzymes, Genitalia Morphology and Morphometry

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Among the triatomines considered as secondary in the epidemiology of Chagas disease, *Rhodnius neglectus* is frequently captured in artificial ecotopes, especially peridomestic ones, rarely producing colonies indoors. Nevertheless, the presence of breeding colonies in houses was unquestionably demonstrated in some areas of the State of Goiás, Brazil. Previous isoenzyme comparisons of this species with morphologically close triatomines, such as *R. prolixus*, *R. robustus* or *R. nasutus*, did not produce definitive conclusions because of doubt about the geographical origin of the *R. neglectus*. We present here, for the first time, the isoenzyme profile of topotypes of *R. neglectus*. In addition, wild caught specimens from the type locality, Uberaba (Minas Gerais, Brazil), were compared to wild caught specimens from Jaraguá (Goiás, Brazil), where *R. neglectus* is more frequently reported invading houses. We used isoenzyme, morphology and morphometry analysis. Neither morphological nor enzymatic differences were found between areas, but metric, size-related divergence was evidenced between them.

Key words: *Rhodnius neglectus* - isoenzyme - morphometry - genitalia - Brazil

Rhodnius neglectus Lent, 1954 is a Brazilian species of Triatominae (Hemiptera, Reduviidae) considered of secondary importance as a vector of *Trypanosoma cruzi*, causative agent of Chagas disease (Silveira & Rezende 1994). It is commonly found in palm tree crowns in Central Brazil, although peridomestic colonies are increasingly reported, and domestic colonies have been found in the State of Goiás (Garcia-Zapata et al. 1985). The factors that favour domiciliation of Triatominae are poorly understood, although studies of *R. neglectus* in Goiás suggest that infestation of houses may follow environmental changes associated with changing patterns of land use (Diotaiuti et al. 1985).

Morphologically, *R. neglectus* is similar to other species of *Rhodnius*, especially *R. prolixus*, *R.*

robustus and *R. nasutus*, which are sometimes grouped with *R. neglectus* as the ‘*prolixus* group’ (Barrett 1988). *R. neglectus* was first separated by Lent (1954) mainly on the basis of minor differences in the male genitalia and in geographic distribution. *R. neglectus* seems confined to the savanna-like cerrado regions of Central Brazil, while *R. nasutus* occupies similar habitats in the drier caatinga of northeastern Brazil; in contrast, *R. prolixus* and *R. robustus* occur in the more humid areas of the Amazon basin northwards into Central America (Lent & Wygodzinsky 1979, Schofield 1994). However, study of these species and definition of their geographic distribution is made difficult by their morphological similarities, and by the poorly defined origin of some laboratory colonies used for comparative purposes (Dujardin et al. 1991, WHO 1991, Harry 1993, Solano et al. 1996).

As part of a wider study on the biosystematics of *Rhodnius*, we report here on the morphometric, morphological and isoenzyme characterization of *R. neglectus* collected from its type locality in Minas Gerais, and from a similar locality in the neighbouring Goiás. The aim of this study was to provide isoenzymatic characterization of the spe-

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cies as well as the metric properties of the topotype specimens, and to assess the level of interpopulation variability.

MATERIALS AND METHODS

The insects - *R. neglectus* adults and nymphs were collected from two species of palm, *Mauritia flexuosa* (known locally as 'buriti') and *Attalea speciosa* (= *Orbignya martiana*) (babaçu) from the type locality of Fazenda Água Limpa, near the town of Uberaba, Minas Gerais, where the insects are frequently found around the houses. They were also collected from buriti palms in the municipality of Jaraguá in the neighbouring Goiás, where intradomiciliation was already reported in a recent past. In both cases, collections were made with the help of local field staff of the Fundação Nacional de Saúde, following the techniques of Diotaiuti and Dias (1984). Laboratory colonies were established from these collections, with the bugs from Uberaba (UB) and Jaraguá (JA) maintained separately in the same insectary at 27+2°C and 60+5% relative humidity, and fed weekly on restrained chickens. The F1 adults from these colonies were used for morphological and morphometric analysis, and electrophoresis of thoracic muscle isoenzymes was carried out on the F1 adults from UB and F2 adults from JA. The number of UB and JA founders was 15 insects (adults and nymphs from three palm trees) and 35 (adults and nymphs from 10 palm trees), respectively.

Morphometry and morphology - Morphometric measurements were taken from 10 males and 10 females from each colony. The insects had the same nutritional conditions and were not fed before morphometry. Seven measurements were taken for each specimen: TL: total body length from anteclypeus to terminalia, PL: length of pronotum,

PS: length of pronotum plus scutellum, HL: total head length (excluding neck), PO: postocular distance, AO: anteoocular distance, and IO: synthlipsis. All measurements were taken by the same investigator using a monocular micrometer at 10 x 12.5 magnification.

The same male specimens were also dissected for morphological examination of the genitalia, following the general procedure of Lent and Jurberg (1969) to examine the parameres, median process of the pygophore, phallus, phallosoma, phallosome support, phallosome process, endosome process and vesica.

Enzyme electrophoresis - Isoenzyme electrophoresis of thoracic muscle extracts from the above specimens was carried out on starch gels following the general procedures of Dujardin and Tibayrenc (1985), with enzyme staining according to Abderrazak et al. (1993). Fourteen enzyme systems were examined, as follows: alanine aminotransferase (EC 2.6.1.2., ALAT), aspartate aminotransferase (EC 2.6.1.1., ASAT), glucose-6-phosphate dehydrogenase (EC 1.1.1.49., G6PD), glucose phosphate isomerase (EC 5.3.1.9., GPI), hexokinase (EC 2.7.1.1., HK), isocitrate dehydrogenase (EC 1.1.1.42., IDH), malate dehydrogenase (EC 1.1.1.37., MDH), malic enzyme (EC 1.1.1.40., ME), mannose phosphate dehydrogenase (EC 5.3.1.8., MPI), peptidase 1 (EC 3.4.11., PEP-1, substrate leucyl-leucyl-leucine), peptidase 2 (EC 3.4.13., PEP-2, substrate leucyl-L-alanine), phosphoglucomutase (EC 5.4.2.2., PGM), 6-phosphogluconate dehydrogenase (EC 1.1.1.44., 6PGD) and pyruvate kinase (EC 2.7.1.40., PK).

Numerical analysis - Due to complete enzyme monomorphism and identity between samples, no detailed genetic analysis was proposed. Quantitative analysis was therefore restricted to the mor-

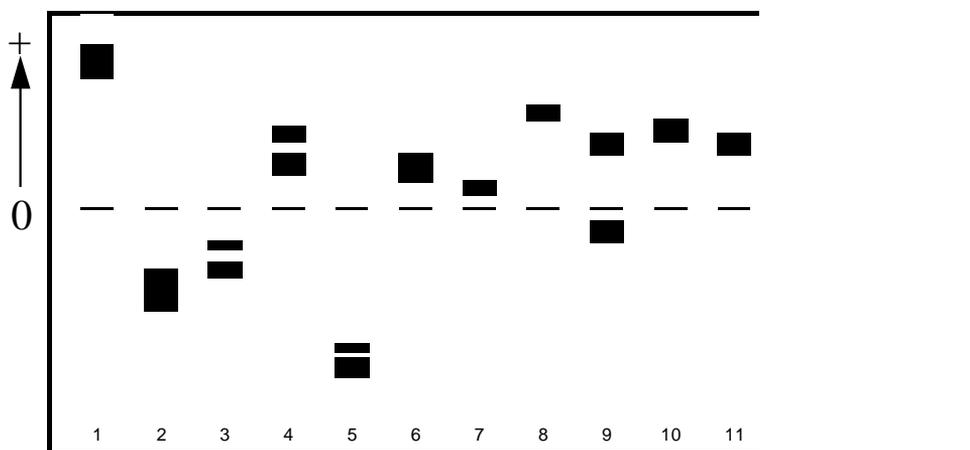


Fig. 1: electrophoretic patterns of *Rhodnius neglectus* from the type locality Uberaba, State of Minas Gerais, Brazil.

TABLE I
Metric properties of the *Rhodnius neglectus* specimens

Organ	Var	Loc	Sex	Obs	Mean	SD	Min	Max	CV
Head	HL	Jaraguá	F	10	0.59	0.03	0.52	0.63	5.63
			M	10	0.54	0.03	0.48	0.55	4.77
		Uberaba	F	10	0.54	0.02	0.50	0.57	3.80
			M	10	0.49	0.03	0.45	0.53	5.72
	AO	Jaraguá	F	10	0.36	0.02	0.31	0.38	6.40
			M	10	0.32	0.02	0.30	0.35	5.03
		Uberaba	F	10	0.32	0.01	0.30	0.35	4.63
			M	10	0.29	0.02	0.25	0.31	6.41
	PO	Jaraguá	F	10	0.13	0.01	0.12	0.15	8.02
			M	10	0.13	0.01	0.11	0.15	8.53
		Uberaba	F	10	0.13	0.01	0.11	0.14	6.48
			M	10	0.12	0.01	0.10	0.13	9.39
IO	Jaraguá	F	10	0.07	0.01	0.06	0.07	7.82	
		M	10	0.06	0.00	0.06	0.07	7.67	
	Uberaba	F	10	0.06	0.01	0.05	0.07	10.20	
		M	10	0.06	0.01	0.05	0.07	9.62	
Thorax	PL	Jaraguá	F	10	0.37	0.03	0.31	0.41	8.42
			M	10	0.36	0.02	0.33	0.39	5.23
		Uberaba	F	10	0.33	0.02	0.30	0.35	6.18
			M	10	0.30	0.02	0.27	0.32	5.10
	PS	Jaraguá	F	10	0.60	0.03	0.55	0.63	4.82
			M	10	0.59	0.02	0.55	0.61	3.04
		Uberaba	F	10	0.55	0.03	0.50	0.60	5.21
			M	10	0.50	0.02	0.47	0.55	4.99
Body	TL	Jaraguá	F	10	2.43	0.12	2.19	2.60	4.87
			M	10	2.29	0.09	2.14	2.45	3.94
		Uberaba	F	10	2.12	0.10	2.00	2.30	4.68
			M	10	2.00	0.10	1.85	2.15	5.04

HL: total head length (excluding neck); AO: anteoocular distance; PO: postocular distance; IO: synthlipis; PL: length of pronotum; PS: length of pronotum plus scutellum; TL: total body length from anteclypeus to terminalia; Var: variable; Loc: locality; Obs: observations; SD: standard deviation; CV: coefficient of variation.

phometric data (Table I). Data were log-transformed for canonical variate analysis (Pimentel 1992) and the results have illustrated by plotting the centroids of each group on the first two canonical variables (Fig. 2) (Albrecht 1980). Table II summarises the corresponding discriminant classifications, with significance estimated according to Wilks (1932). All computations used JMP software (SAS Institute 1995).

RESULTS

Morphometry and morphology - Male genitalia showed no qualitative differences between samples, and presented all the features characteristic of *R. neglectus* as described by Lent and Jurberg (1969) and Lent and Wygodzinsky (1979). Only *R. neglectus* was captured in our studied areas.

By morphometry, all lengths were smaller in the Uberaba specimens, although there was greater

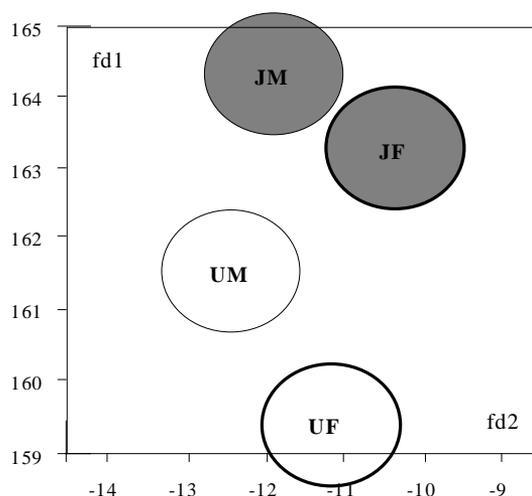


Fig. 2: canonical variate analysis of the two *Rhodnius neglectus* populations from Jaraguá, State of Goiás (J) and Uberaba, State of Minas Gerais (U) according to sex (M=males and F=females).

variability among the JA specimens. Females were also generally larger than males in both localities, and the discriminant analysis was highly significant (Pwilks < 0.0001) giving a substantial differentiation (Kappa = 0.77) between both sexes and localities (Table II, Fig. 2).

Isoenzymes - No polymorphism, nor difference in electrophoretic migration at any locus was found in comparisons of the specimens from UB and JA. In this study, we did not succeed in visualising the G6PD, 6PGD and PEP-1 enzyme systems, although other authors have shown these to be monomorphic in other samples of *R. neglectus* (Dujardin et al. 1991, Harry 1992, Solano et al. 1996). The remaining 11 enzymes each revealed a single zone of activity, except for GPI, IDH, MDH and PEP-2 which each revealed two bands, so that we could score a total of 15 loci controlling the synthesis of these systems (Fig. 1).

DISCUSSION

R. neglectus was first separated from other *Rhodnius* species by Lent (1954). Prior to that date (and in some subsequent studies) the prevailing *Rhodnius* species found in palm tree crowns and peridomestic habitats throughout the cerrados of central Brazil had been reported as *R. prolixus*, and the confusion has been repeated in many uncritically published distribution maps. But clarification of the distribution of these species is important because of their differing epidemiological significance. *R. prolixus* is the main domestic vector of *T. cruzi* in Venezuela, Colombia and parts of Central America, where it is subject to increasing attention as a primary target for vector control interventions (Schofield & Dujardin 1997). In contrast, *R. neglectus* is considered a secondary vec-

tor in Central Brazil, which has not achieved great epidemiological significance because of its mainly silvatic and peridomestic habits (Diotaiuti et al. 1995). Moreover, the distributional question is further confounded by the similarities with two other *Rhodnius* species - *R. nasutus* in northeastern Brazil, and *R. robustus* in the Amazon region (Schofield 1994, WHO 1991). Although in our experience the genital characteristics alone do not permit us readily to separate *R. neglectus* from other species of the 'prolixus group', they do readily distinguish *R. neglectus* from other species that might conceivably have occurred in our sample areas. In spite of it, only *R. neglectus* was captured in our studied areas.

Barrett (1988) suggested that the genus *Rhodnius* could be considered as a complex of species without clear-cut interspecific boundaries, and the morphological similarities between many of the species - especially of the 'prolixus group' - have certainly led to confusion in defining their geographical distribution (Dujardin et al. 1991, WHO 1991). This problem has also been exacerbated by the use of laboratory colonies, often of uncertain origin and always with the risk of contamination between different *Rhodnius* strains. For this reason, previous isoenzyme studies of the group have often given results that are difficult to interpret. Dujardin et al. (1988, 1991) could find no enzymatic difference between various samples of *R. prolixus* and samples of *R. neglectus* thought to originate from chicken coops in Goiás. Subsequently however, various *R. prolixus* strains were found to differ at four or five loci from the Fiocruz reference strain of *R. neglectus* (maintained for many years in the laboratory of the Instituto Oswaldo Cruz in Rio de Janeiro) (Dujardin 1990,

TABLE II
Canonical variate analysis of the morphometric measures
CV1 81% - CV2 14% - P Wilks < 0.0001

Count		Reclassification				Predicted
		Jaraguá		Uberaba		
		Males	Females	Males	Females	
Jaraguá	Females	7	0	0	0	7
	Males	3	8	0	0	11
Uberaba	Females	0	0	9	1	12
	Males	0	2	1	9	12
Observed		10	10	10	10	40
Kappa = 0.77						

Between localities, the observed agreement was 95.5% with only two individuals wrongly classified. Between sexes, the observed agreement was 87.5%. The Kappa statistics measure the agreement between observed and predicted groups. It is scaled to be 0 when the agreement is as expected by chance and 1 when agreement is perfect. For intermediate values, Landis and Koch (1977) suggest the following interpretation: 0.00-0.20, slight; 0.21-0.40, fair; 0.41-0.60, moderate; 0.61-0.80, substantial.

Dujardin et al. 1991, Solano et al. 1996) and marked isoenzyme differentiation was found by Harry (1993) between reproductively isolated strains of *R. prolixus* and *R. neglectus* from various localities.

To help resolve this problem, we present here a clearly defined isoenzyme pattern derived from topotype specimens of *R. neglectus* which may be considered as the type profile of the species. Comparison with other results (Dujardin et al. 1991, Harry 1993, Solano et al. 1996) shows that this profile will readily distinguish *R. neglectus* from other species of *Rhodnius*. Moreover, our study has shown complete enzyme monomorphism both in the topotype specimens and in specimens from another locality ($P=0$, with P as the proportion of polymorphic loci). We doubt that this would be due to small sample size or genetic drift because of the number of founders collected from the field, and the small number of generations (one or two) that elapsed between our original collections and the isoenzyme analysis. Very low levels of enzyme polymorphism have been found by other authors in laboratory strains of *R. neglectus* ($P=0.07$, Dujardin et al. 1991), also in wild caught *R. pictipes* ($P=0.10$ to 0.19 , Dujardin et al. 1988, Harry et al. 1992a), in *R. prolixus* from various sources ($P=0.07$ to 0.23 , Dujardin et al. 1988, Harry 1992, Harry et al. 1992a, b) and in *R. robustus* ($P=0$ to 0.16 , Harry 1992, Harry et al. 1992a, b). These and our results thus lend support to the idea of low genetic variability within the genus *Rhodnius* (Dujardin et al. 1998) and also suggest that the type profiles will provide a good representation of *R. neglectus* throughout its range.

In contrast to the isoenzyme results however, the morphometric analysis revealed clear differences between the two populations studied here. The UB (topotype) specimens were smaller than those from JA, although the latter showed greater variability. The two localities represent isolated populations, subsequently reared in identical insectary conditions, so that we could attribute these morphometric differences to genetic drift between the two, environmental changes, climatic and others. Reports from the National Health Foundation of Brazil have indicated a higher likelihood of finding domestic *R. neglectus* in the JA region compared to the UB region, so that it is tempting to suggest that the greater metric variability of the JA specimens may indicate greater behavioural plasticity, but this must await the results of further study.

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