

REVIEW

Open Access

Outbreaks of Ebola virus disease in Africa: the beginnings of a tragic saga

Jean-Philippe Chippaux^{1,2}

Abstract

The tremendous outbreak of Ebola virus disease occurring in West Africa since the end of 2013 surprises by its remoteness from previous epidemics and dramatic extent. This review aims to describe the 27 manifestations of Ebola virus that arose after its discovery in 1976. It provides an update on research on the ecology of Ebola viruses, modes of contamination and human transmission of the disease that are mainly linked to close contact with an infected animal or a patient suffering from the disease. The recommendations to contain the epidemic and challenges to achieve it are reminded.

Keywords: Ebola, Outbreak, Virus, Hemorrhagic fever, Africa

Introduction

The outbreak of Ebola virus disease (EVD) occurring in West Africa since December 2013 will mark the history – however brief – of the viruses. Occurring for the first time outside its original home – the Central African rainforest – this epidemic of EVD appears as the deadliest and longest of all those known so far. At this time (August 15th, 2014), more than 1,250 deaths have been reported, *i.e.* five times more than during the worst previous outbreak. In addition, its rapid propagation has led the World Health Organization (WHO) to declare on August 8th that EVD represent a “Public Health Emergency of International Concern” and urged the international community to take action to stop the spread. Finally, for the first time, in response to the severity of the situation, WHO agreed the use of experimental treatment against the EVD.

Review

Discovered in 1976 during two inaugural epidemics, both in Sudan and Democratic Republic of Congo (DRC), the Ebola viruses were responsible for 27 occurrences in Africa before the current outbreak of Guinea. This review aims to remind the characteristics of the different epidemics of EVD that were reported between 1976 and 2013.

Correspondence: jean-philippe.chippaux@ird.fr

¹UMR 216, Mother and Child Facing Tropical Diseases, Institut de Recherche pour le Développement (IRD), Cotonou, Bénin

²Université Paris Descartes, Sorbonne Paris Cité, Faculté de Pharmacie, Paris, France

Ebola viruses

The Ebola virus is, together with *Marburg marburgvirus*, native to East Africa, and belongs to the *Filoviridae* family (Table 1), whose newest member is *Lloviu cuevavirus*, recently isolated in Spain from bats [1-3].

Ebola viruses are RNA viruses whose genome encodes seven proteins [4,5]. They are of filamentary form, sometimes branched, with a diameter of 80 nm and a length of up to 14,000 nm (Figure 1). The protein shell encloses the tubular helical nucleocapsid. Surface transmembrane glycoproteins of the virion provide the binding and fusion with the cell membrane, and penetration into the cell. Glycoproteins are responsible for almost all the virulence, even though it does not explain all the pathogenicity [6]. Monocytes, particularly macrophages, are the first cells infected, triggering apoptosis in lymphocytes [7]. Within three days, the virions invade the endothelial system [4,5]. The inhibition of the immune response, including reduced production of interferon, favors the rapid spread of the virus in the body [8,9].

The genus Ebola comprises five species, including the four African species involved in human clinical cases [3,10]. Three of them were identified during Central African epidemics: first, *Sudan ebolavirus* and *Zaire ebolavirus*, both in 1976 respectively in Sudan and DRC, and more recently *Bundibugyo ebolavirus*, in 2007 in Uganda [11-14]. A fourth species, *Tai Forest ebolavirus*, was isolated from a primatologist who autopsied a chimpanzee in Côte d'Ivoire and survived the disease [15]. The last species, *Reston ebolavirus*, was isolated from monkeys native to Philippines

Table 1 Filoviridae virus occurrences in the world

Viral species	Year of discovery	Geographic origin	Number of outbreaks	Number of human cases	Number of deaths (CFR)	CFR (%)
<i>Marburg marburgvirus</i>	1967	Uganda	4	465	145	31
<i>Sudan ebolavirus</i>	1976	Sudan	6	792	426	54
<i>Zaire ebolavirus</i>	1976	DR Congo	12	1,388*	1,100*	79
<i>Reston ebolavirus</i>	1989	Philippines	0	0	0	–
<i>Tai Forest ebolavirus</i>	1994	Côte d'Ivoire	0	1	0	–
<i>Bundibugyo ebolavirus</i>	2007	Uganda	2	208	78	38
<i>Lloviu cuevavirus</i>	2010	Spain	0	0	0	–

CFR: case fatality rate. *Excluding the current West African outbreak.

farms [16]. The disease is deadly to monkeys. However, no human cases have been reported to date despite the presence of antibodies proving the infection in pigs (also without clinical illness) and humans [17].

According to Carroll *et al.* [10], the nearest common ancestor of Marburg and Ebola viruses dates back to 1,300 years. The separation of *Sudan ebolavirus* and other species of Ebola would have occurred soon after (about 1,200 years). The other species would have been separated much later, probably in the last hundred years or less (Figure 2).

Clinical presentation

Incubation ranges from 3 to 21 days [21]. Eichner *et al.* [22] determined that the incubation period was 12.7 ± 4.3 days, which fixes the average time between two generations of patients. The epidemic is considered complete after an interval of at least twice the maximum incubation period, *i.e.* 42 days after the death or recovery of the last confirmed case [23].

The disease lasts 5 to 15 days [24]. It begins suddenly with fever, headache, abdominal pain, arthralgia and myalgia, therefore the general picture resembles a flu syndrome [25-29]. About half of patients complain of cough and sore throat with dysphagia. Digestive disorders (diarrhea, nausea, vomiting) follow in a variable proportion of patients. Hemorrhages occur in 30-80% of patients, mostly

at the end of the illness, expressed by purpura, epistaxis, gingival bleeding, gastrointestinal bleeding or other, and appear to be associated with the severity of infection [24-30]. The variation in prevalence of symptoms, especially bleeding, is related to the viral species or clinical description that is based, according to the studies, on either suspected or confirmed cases.

Mortality is always high, although pathogenicity varies from one species to another (Table 1). In addition, some authors noted that mortality decreased during the epidemic. Several hypotheses have been advanced: loss of the virus virulence after successive generations, route of inoculation, viral load, improved management of cases and better enforcement of prophylaxis during the epidemic [24,26,30-33]. One cannot exclude that improving diagnosis in late epidemic reveals less severe cases.

Diagnosis

The clinical diagnosis is difficult at the beginning of the epidemics, because of the poor specificity of the symptoms [24,27,29,30,33]. When the virus responsible for the outbreak is identified, all suspected cases should be considered as high risk and meet the case definition and exposure risks (Tables 2 and 3) for better management of the epidemic. There is no carrier state.

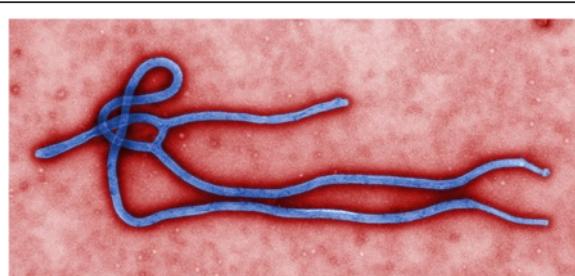


Figure 1 Ultrastructural morphology of Ebolavirus virion (image by US Centers for Disease Control and Prevention and Cynthia Goldsmith).

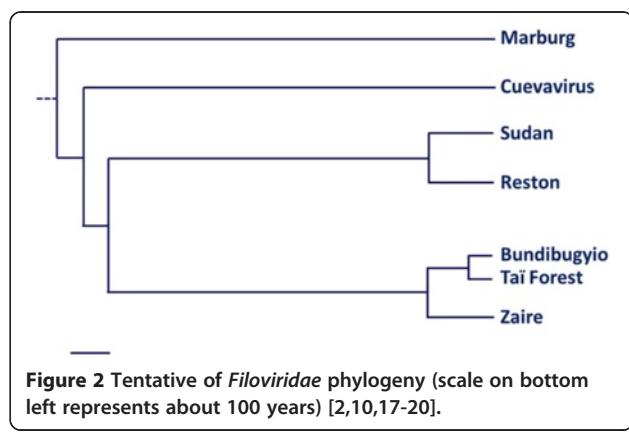


Figure 2 Tentative of Filoviridae phylogeny (scale on bottom left represents about 100 years) [2,10,17-20].

Table 2 Case definition of Ebola Virus Disease (EVD) [23,34]

Name	Definition
Index case	Very first case (probable or confirmed, see below) found to be the origin of the outbreak
Alert case	Any person with sudden onset of high fever or sudden death or bleeding or bloody diarrhea or blood in urine
Suspect case (person under investigation)	Any person, dead or alive, who present (or presented before the death): (i) fever (>38.5°C or 101.5 °F) with additional symptoms (severe headache, muscle pain, vomiting, diarrhea, abdominal pain, or unexplained hemorrhage) and (ii) epidemiologic risk factors within the past 21 days before the onset of symptoms (close contact with body fluids of a suspect or probable case of EVD, or direct handling of bush animals from disease-endemic areas)
Probable case	Person with symptoms compatible with EVD, as evaluated by a clinician, or a dead person with an epidemiological link with a confirmed case
Contacts	Person without suggestive symptom of the disease, but who has been in contact with a suspect or probable case of EVD (living in the same house, provided care during the illness, participated in the burial rites etc.). It should be important to assess the risk level (see Table 3).
Confirmed case	If laboratory samples are obtained at an appropriate time during the illness, the previous notification categories should be reclassified as "laboratory-confirmed" cases and "not a case"
"Not a case"	Case with positive laboratory response for either Ebola virus antigen or Ebola IgG antibody
	Person with no Ebola-specific detectable antibody or antigen

Laboratory diagnosis can only be performed in a specialized laboratory. First, there is no commercial reagent and, secondly, the samples represent an extreme biohazard that must be handled under containment conditions of the highest level (biosafety level 4 – BSL-4; Figure 3).

In patients, the diagnosis is carried out by the detection of viral antigens through ELISA, identification of nucleic acid by PCR, specific antibody titer, or virus isolation. Specific IgM and IgG antibodies appear during the second week following the first clinical signs (about 15 to 20 days after the infection). IgM titers persist about two months whereas IgG titers remain several years after the end of the disease [37].

Virus isolation can be achieved by inoculation in mice, guinea pigs or non-human primates in whom the disease

is very close to what is observed in humans. The virus grows on kidney cell lines from African green monkey *Cercopithecus aethiops*. The diagnosis is made by optical microscopy examination of cytopathic effects, visualization of the virions by electron microscopy, identification of specific proteins by ELISA, or RNA detection by PCR.

IgG titration allows retrospective diagnosis in convalescents or exposed persons, or epidemiological investigations even years after the epidemic.

Treatment

Antibodies acquired during the disease persist in survivors for more than ten years after the recovery. There is still no evidence that primary infection is protective in

Table 3 Definition and assessment of risk exposure [23,34-36]

Risk level	Definition
High-risk exposure	<ul style="list-style-type: none">Percutaneous injury, e.g. needlestick, or mucous membrane exposure to body fluids of an EVD patientDirect care or exposure to body fluids of an EVD patient without appropriate personal protective equipment (PPE)Laboratory worker processing body fluids of confirmed EVD patients without appropriate PPE or standard biosafety precautionsParticipation in funeral rites that include direct contact with human remains in the geographic area where an outbreak is occurring without appropriate PPE
Low-risk exposure	<ul style="list-style-type: none">Household member or other casual contact¹ with an EVD patientProviding patient care or casual contact¹ without high-risk exposure with EVD patients in health care facilities in EVD outbreak affected countries
No known exposure	Persons with no known exposure were present in an EVD outbreak affected country in the past 21 days with no low-risk or high-risk exposures

¹Casual contact is defined as (i) being within approximately 3 feet (1 meter) or within the room or care area for a prolonged period of time (e.g. healthcare personnel, household members) while not wearing recommended personal protective equipment; or (ii) having direct brief contact (e.g., shaking hands) with an EVD case while not wearing recommended personal protective equipment



Figure 3 Study of Ebola virus in a high-security laboratory BSL-4 (photo by IRD, ©IRD).

humans, apart from neutralization tests *in vitro* [38]. Moreover, up to this moment, there is no vaccine or effective treatment.

Studies on preventive vaccines are in progress [39,40]. Some of them showed very good efficacy and are produced under good manufacturing practices conditions. Phase I trials demonstrated that the drug is safe for humans [40]. However, vaccine development is hampered by limited commercial interests, even taking into account the risks of bioterrorism-related dispersion of such virus. The limited number of cases, despite the high mortality, adds to the complexity – and cost – of large-scale immunization of a scattered and often inaccessible population [41].

Post-exposure management should consider either passive immunotherapy, or administering drugs that block the action of the virus or its replication. Convalescent sera, thought to contain natural specific protective antibodies developed during the disease, have been used exceptionally, with some success [42,43]. It is noteworthy that these patients had also received better symptomatic treatments than regular patients, and were not representative.

However, despite this indisputable bias, several experimental studies have confirmed, particularly in non-human primates, the effectiveness of such approach [44]. The development of specific drugs is underway with some promising molecules, including monoclonal antibodies, which use is related to passive immunotherapy [45]. Recently, Warren *et al.* [46] showed that a new nucleoside analogue protects against infection with *Filoviridae* by inhibiting the viral polymerase in the *Macacus cynomolgus* model that, moreover, seems to tolerate well the treatment.

Symptomatic treatments are poorly documented because of the rarity of the disease (less than 2,500 known cases worldwide between 1976 and 2013) and the paucity of resources in endemic countries, which limits the choice of drugs [47]. Administration of antihemorrhagic drugs, substitution treatments, including transfusions, plasma-apheresis or dialysis, and resuscitation are anecdotal and concern very few patients [47]. Although one of the main virus targets, interferon has not been an effective treatment, which confirms that the impairment of the immune system is deep [4,5,47,48]. Most often, palliative treatments are limited to rehydration with sugar solutions, preferably orally to avoid injections, analgesic, antipyretic, antiemetic, anti-diarrheal and sedatives or antipsychotic drugs to ease agitated and anxious patients.

Natural history of ebola viruses

The emergence, less than 50 years ago, of Ebola virus remains an enigma. It is possible that sporadic cases or limited outbreaks occurred in the past and were unreported due to lack of epidemiological surveillance or appropriate diagnosis. Scattered and minor manifestations of EVD suggest that Ebola viruses circulate in a pathogen complex showing no (or few) contacts with human populations. Destruction of forests and human impact on broader areas could explain the increased frequency and severity of outbreaks [49]. However, we cannot exclude a recent emergence of the virus that would spread rapidly in a susceptible population [50]. According to Polonsky *et al.* [51], increasing frequency of epidemics may result from the combination of: improvement of monitoring and diagnostic capacities, increase of contact among humans and the natural reservoirs of the virus, and growth of the viral load and prevalence of the virus in reservoirs.

Several epidemiological investigations in Central and East Africa have shown circulation of Ebola virus in the human population at a significant rate, but that does not always entail the emergence of an epidemic [24,52,53].

The natural reservoir of the virus is not known with certainty. Extensive investigations made in small mammals, even sensitive to the Ebola viruses, were negative during the various epidemics in Central Africa [26,54-56].

Subsequent investigations continued outside epidemics. Although viral RNA and specific antibodies have been identified in small mammals, no potential natural host has been acknowledged until 2005 [57]. However, initially dismissed due to many negative samples, fruit bats (Figure 4) were found with specific viral DNA and antibodies. These animals seem resistant to *Filoviridae* pathogenicity [18,58-63].

The search for potential vectors, especially among arthropods, has always proved negative, including bedbugs (*Cimex hemipterus*) captured in the beds of infected persons [26,54,64,65].

Deadly outbreaks of Ebola virus have been observed in non-human primates with high mortality [66]. In addition to the contamination of the Swiss primatologist in Ivory Coast [15], the index case of several outbreaks have been more or less directly associated with hunting or consumption of bush meat, i.e. monkeys, antelopes, bats [26,58,65,67-71]. Natural infection of bats and sharing their narrow ecological niche with many species of non-human primates are strong arguments in favor of their role as a natural reservoir of the virus [58-61]. Some species, including *Eidolon helvum*, *Hypsognathus monstrosus*, *Myonycteris torquata* and *Epomops franqueti*, migrate long distances (>2,500 km), which could explain the multiple remote epidemic clusters [58,72,73].

Seasonal variation in mortality in chimpanzees of the Tai forest, Ivory Coast, and prevalence of specific antibodies against *Zaire ebolavirus* virus in febrile patients from East Africa suggests an influence of the climate in the occurrences of Ebola epidemics [66,74]. Pinzon *et al.* [75] found a close relationship between the onset of epidemics and particularly dry conditions at the end of the rainy season, leading to a change in the behavior of fruit-eating mammals, particularly sensitive to weather

changes, resulting in the increase of virus circulation or human contamination [76]. The seasons punctuate migration of bats, which could explain the emergence of epidemics [58,62].

Human transmission of Ebola viruses

The contamination of index cases is probably due to contact with an infected animal. Human transmission happens only through close contact with an ill or convalescent person, although at this stage the risk of infection is very small. Studies conducted during the various epidemics have shown that less than one fifth of the people (see Tables 2 and 3) living with a confirmed or probable primary patient have developed the disease [24-26,35]. All secondary cases were recorded among people with close contact with the patient and exposed to infected biological fluids. Conversely, people who had no contact with the patient were not sick. Such close contact with the patient throughout care occur mainly during the illness or burial preparation, including washing the body and funeral ritual that can be long and intimate [25,26,77-79]. The risk greatly increases due to the delay in diagnosis and appropriate management [24,27,29,30,33].

Ebola viruses have been detected in most patient secretions. They are present in the blood, saliva, feces, breast milk, tears and genital secretions. They have not been isolated from vomit, sputum, sweat or urine. However, the number of tested samples was low [80]. The virus persists in breast milk, genital secretions and glass during convalescence and up to 13 weeks after recovery [27,80,81]. Finally, the risk of transmission from fomites (towels, clothes and sheets from the patient), especially during convalescence, is low and basic protection measures are likely to be sufficient [80].



Figure 4 Group of bats *Hypsognathus monstrosus* in a mango tree in Lambaréne, Gabon (photo by Jean-Jacques Lemasson, ©IRD).

Nosocomial transmission is behind many hospital outbreaks [21,24,28]. Injection materials reused without precaution or inadequately sterilized have been repeatedly denounced and remain a major cause of epidemic spread. This also applies to traditional healers whose practices are often septic [67,82].

Finally, all the studies performed during the outbreaks of EVD showed that contamination occurs due to close contact with the blood or secretions of an infected patient through three ways [25,26,35,36]:

- Patient care – usually a family member takes care of the patient during the illness.
- Preparing the deceased for funeral before or during burial.
- Nosocomial transmission – reuse of medical equipment that has been previously used in a patient infected with Ebola virus.

There is no contamination by air or just handshake.

Manifestations of Ebola viruses in Africa (1976–2012)

Since their discovery in 1976, simultaneously in Sudan and DRC, and until 2012, i.e. the recent outbreaks observed in Uganda and again in the DRC, Ebola viruses were notified 27 times (Table 4, Figure 5), including in 22 epidemics. It is possible that sporadic cases or limited outbreaks have escaped any mention due to the remoteness and poverty of the concerned people.

Some individual infections, such as those observed in Tandala (DRC) in 1977, Taï (Côte d'Ivoire) in 1995, Luwero (Uganda) in 2011, indicate an occult but permanent circulation of Ebola viruses [15,83,95]. The confirmed case in Gozon (Côte d'Ivoire) in 1995, which may have come from Liberia, has not been completely documented and is not mentioned after the year 1999 [85,86]. The index case in Johannesburg in 1996 was a patient infected in Gabon, where he was staying during the Ebola outbreak in that country [82]. Finally, some epidemics may be the expression of successive waves of the same epidemic, as in Gabon in 1996, Gabon and Congo between 2000 and 2003, and Uganda in 2012, confirming the persistence of the Ebola virus.

Beyond the diversity of African outbreaks, particularly regarding the incidence and duration of the epidemics (Table 5), we can notice that most of them presented only one source of infection accounting for the spread of the virus. As a result, it can be assumed that either the opportunities for human contact with the virus are rare, or the risk of contamination is limited. The period between the index case and the alert averages about two months. In addition, an equivalent period occurs between the alert and the last confirmed case (Figure 6).

The origin of the disease and its management are not perceived the same way by different people [77-79,82]. At the beginning of epidemics, the disease is often confused with another endemic infection (such as malaria, dysentery, typhoid or yellow fever, influenza etc.), both by the general population and health personnel. When the incorrect causes are eliminated or diagnosis of EVD is confirmed, other explanations of cultural, religious, circumstantial or biomedical origins are proposed. It has been shown, in several epidemics, that affected people perceived the illness as divine punishment or evil spell, and even, sometimes, denied the disease itself [77-79]. Hesitations about diagnosis and the diversity of explanatory anthropological models result in the delay of the alert and difficulties in the implementation of measures against epidemics [79,82,92].

Finally, health authorities, for economic and political reasons, take late measures that only aim at minimizing the stress provoked by epidemics and avoiding panic. This results in inadequate or contradictory information, which adds to the general stress. Moreover, some measures taken to demonstrate the determination of the authorities are useless and counterproductive. The ban of gatherings and travel, border closures, police cordons and other initiatives only reinforce distrust of people *vis-à-vis* the authorities and health personnel.

Management of outbreaks

The delay in clinical diagnosis and epidemic alert represent a major issue because it exacerbates the spread of the virus.

In the absence of any vaccine or post-exposure treatments, it is essential to break the chain of transmission by acting directly on the causes and circumstances of outbreaks. All the precautions should be taken for patient care, management of dead bodies, burial and surveillance of contacts [106]. The Centers for Disease Control and the World Health Organization provide a manual in English, French and Portuguese (<http://www.cdc.gov/vhf/abroad/vhf-manual.html>) enabling health authorities to prepare for EVD outbreak management. After the description of the standard precautions regarding care of all patients to prevent infections (a brief summary of Good Medical Practice), the manual details the techniques used to control epidemics of EVD with many simple and practical hints:

- Clinical diagnosis of the disease.
- Isolation of the patient and planning the isolation area.
- Wearing appropriate clothing during treatment (Figure 7).
- Identification and monitoring of contacts.

Table 4 Characteristics of the African manifestations of Ebola virus (bolded names indicate the place of first case occurrence)

Year	Country	Districts	Ebola species	Length (weeks)	Number of cases			Ways of transmission	References
					Presumed*	Confirmed	Deaths		
1976	Sudan	Nzara , Maridi, Tembura, Juba	Sudan	22	227	57	151	284	Nursing patient Nosocomial**
1976	DRC	Yambuku , Abumombazi, Kinshasa	Zaïre	9	307	11	280	318	Nursing patient Funeral/burial ritual
1977	DRC	Tandala	Zaïre	-		1	1	1	[83]
1979	Sudan	Nzara , Yambio	Sudan	10	24	10	22	34	Nursing patient
1994	Gabon	Minkouka , Andock, Minkébé	Zaïre	13	32	19	31	51	Nursing patient
1994	Côte d'Ivoire	Taï	Taï Forest	-		1	0	1	[15]
1995	Côte d'Ivoire /Liberia	Gozon	?			1	0	1	[85,86]
1995	DRC	Kikwit , Mosango (\pm 30 villages)	Zaïre	27	233	82	255	315	Nursing patient Funeral/burial ritual
1996	Gabon	Mayibout	Zaïre	12	29	2	21	31	Eating bush meat Funeral/burial ritual
1996	Gabon	Booué , Balimba, Lastourville, Libreville	Zaïre	27	56	4	45	60	Eating bush meat Nursing patient
1996	South Africa	Johannesburg	Zaïre	-	0	2	2	2	[82]
2000	Uganda	Gulu , Masindi, Mbarara	Sudan	20	230	195	224	425	Nosocomial** Nursing patient Nosocomial**
2001-2002	Gabon	Mékambo , Makokou, Franceville	Zaïre	21	37	28	53	65	Nursing patient Funeral/burial ritual Nosocomial**
2001-2002	Congo	Mbomo, Kellé	Zaïre	20?	50	9	44	59	Nursing patient Funeral/burial ritual
2002	Congo	Mbomo	?	10	9		8	9	?
2002	Gabon	Ekata	?	10	2		2	2	?
2002-2003	Congo	Mbomo, Kellé	Zaïre	17	130	13	128	143	Nursing patient Funeral/burial ritual
2003	Congo	Mbomo , Mbandza	Zaïre	7	18	17	29	35	Nursing patient Funeral/burial ritual
2004	Sudan	Yambio	Sudan	10	4	13	7	17	Nursing patient

Table 4 Characteristics of the African manifestations of Ebola virus (bolded names indicate the place of first case occurrence) (Continued)

									Funeral/burial ritual	
									Nursing patient	[70]
2005	Congo	Etoumbi , Mbomo	Zaïre	6	11	1	10	12	Funeral/burial ritual	
									Nursing patient	
									Funeral/burial ritual	[70]
2007	DRC	Luebo	Zaïre	17	≤ 170	≥ 17	186	264	No data	[47,19]
2007	Uganda	Bundibugy , Kikyo	Bundibugyo	20	75	56	42	131	Nursing patient	
									Funeral/burial ritual	[29,30,33,94]
2008	DRC	Luebo , Mweka	Zaïre	5	≤ 29	≥ 3	15	32	No data	[19]
2011	Uganda	Luwero	Sudan			1	1	1		[95]
2012	Uganda	Kibale	Sudan	11	13	11	17	24	No data	[96,97]
2012	Uganda	Luwero , Kampala	Sudan	8	3	6	4	7	No data	[97-99]
									Nosocomial**	
2012	DRC	Isiro , Pawa, Dungu	Bundibugyo	29	41	36	36	77	Nursing patient	[79,100]
									Funeral/burial ritual	

*Alert, suspected or probable case, i.e. diagnosis based on clinical and/or epidemiological criteria but not biological evidence (see Table 3). In some outbreaks, case definition changed during the epidemics.

**Hospital transmission due to needle and syringe contamination, contact with patient's blood, secretions or fomites.

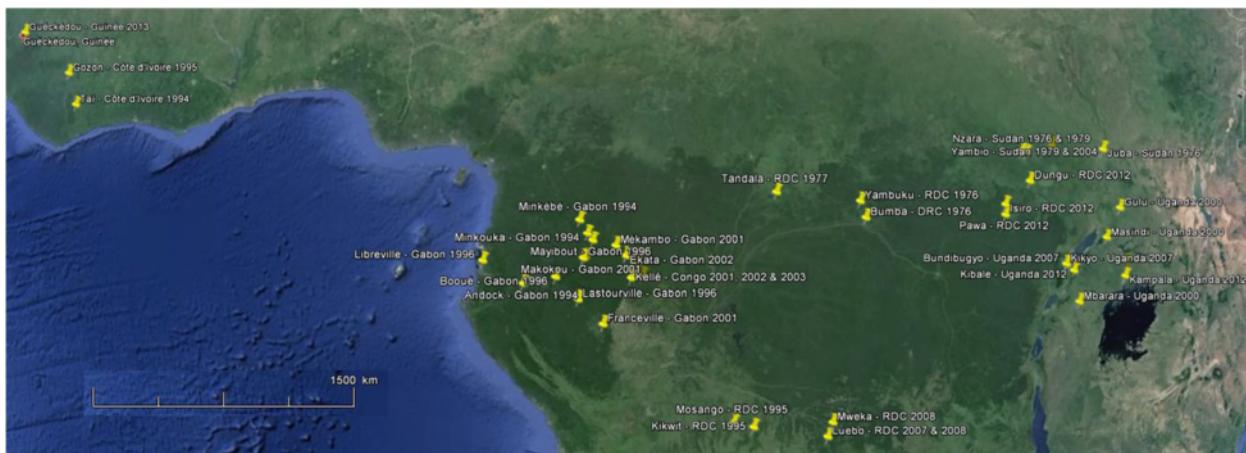


Figure 5 Geographical distribution of African manifestations of Ebola viruses (based on Google™ Earth map).

- Disinfection and sterilization of equipment and facilities.
- Waste disposal.
- Preparation of the body (Figure 8).
- Transportation to the place of burial (Figure 9).

However, the management of cases and now well-known procedures face many difficulties that arise quickly in most outbreaks. In addition to the virulence of Ebola viruses and usual customs, it should be taken into account the changes in behavior induced

Table 5 Length of African Ebola outbreaks

Outbreak	Index case	Alert	Last case	Length of outbreak*	Number of outbreak sources	References
Sudan 1976	Jun. 27, 1976	Sep. 15, 1976	Nov. 25, 1976	151 days	1	[25,101]
DRC 1976	Sep. 1, 1976	Sep. 21, 1976	Nov. 5, 1976	66 days	1	[26,102]
Sudan 1979	Jul. 31, 1979	Sep. 12, 1979	Oct. 6, 1979	67 days	1	[24,103]
Gabon 1994-5	Nov. 13, 1994	Dec. 18, 1994 ¹	Feb. 9, 1995	88 days	?	[67,84,104]
DRC 1995	Jan. 6, 1995	May 1, 1995	Jul. 16, 1995	191 days	1	[28,87]
Gabon 1996	Jan. 31, 1996	Feb. 13, 1996	Mar. 12, 1996	83 days	1	[67,88,89]
Gabon 1996-7	Jul. 13, 1996	Oct. 5, 1996	Jan. 18, 1997	189 days	?	[67,90,91]
Uganda 2000-1	Aug. 30, 2000	Oct. 8, 2000	Jan. 14, 2001	137 days	1	[21,105]
Gabon 2001-2	Oct. 25, 2001	Nov. 17, 2001	Mar. 22, 2002	148 days	5	[69,34]
Congo 2001-2	ND	ND	ND	ND	?	[34]
Congo 2002	May 17, 2002	June 6, 2002	Jul. 25, 2002	69 days	1	[34]
Gabon 2002	May 17, 2002	Jun. 21, 2002	Jul. 25, 2002?	69 days?	1	[34]
Congo 2002-3	Dec. 25, 2002	Jan. 28, 2003	Apr. 22, 2003	118 days	3	[71]
Congo 2003	Oct. 11, 2003	Oct. 24, 2003	Dec. 2, 2003	52 days	1	[92]
Sudan 2004	Apr. 15, 2004	May 6, 2004	June 26, 2004	72 days	1	[93]
Congo 2005	Apr. 18, 2005	Apr. 26, 2005	May 27, 2005	39 days	1	[70]
DRC 2007	Jun. 12, 2007	Aug. 22, 2007	Oct. 10, 2007	120 days	1	[58,19]
Uganda 2007-8	Aug. 20, 2007	Sep. 15, 2007	Jan. 8, 2008	141 days	1	[30,33,94]
DRC 2008-9	Nov. 27, 2008	Dec. 25, 2008	Jan. 1, 2009	34 days	1	[19]
Uganda 2012 ²	Jun. 11, 2012	Jul. 24, 2012	Aug. 24, 2012	74 days	1	[96,97]
Uganda 2012 ²	Oct. 13, 2012	Nov. 14, 2012	Dec. 5, 2012	53 days	1	[97-99]
DRC 2012 ²	Mar. 20, 2012	Aug. 17, 2012	Oct. 11, 2012	206 days	?	[79,100]

*Between the index case and recovery or death of the last case. ¹First reported as yellow fever [85]. ²Partial unconsolidated data. ND: not determined.

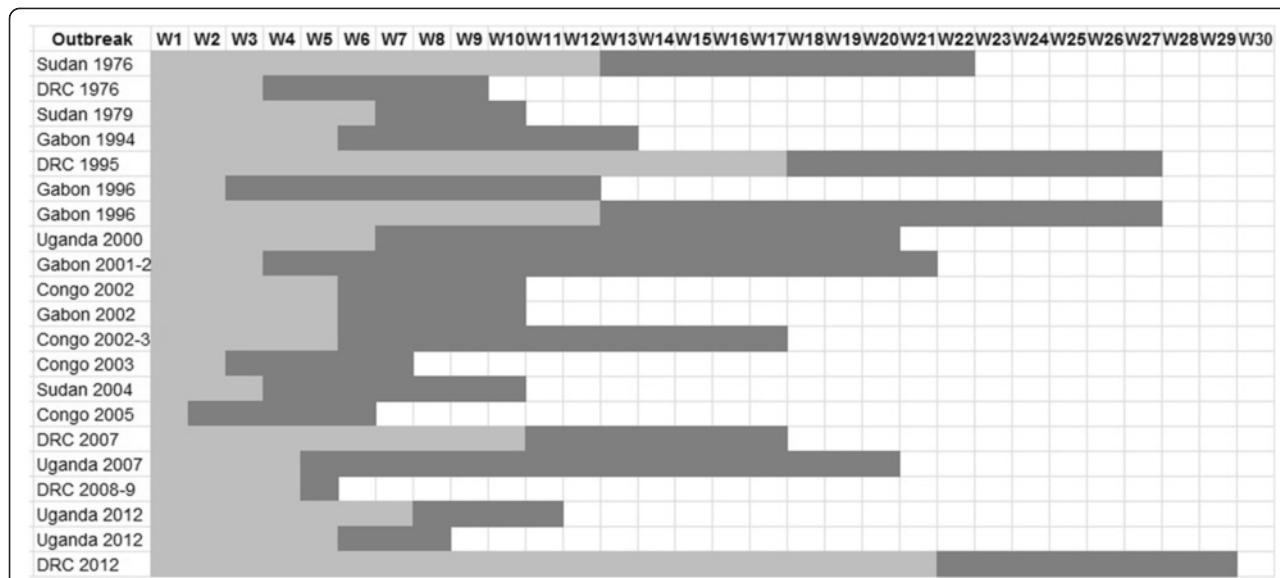


Figure 6 Alert delay and duration of the African Ebola outbreaks.



Figure 7 Protective clothing worn by healthcare personnel during the care and handling of a patient (photo by Jean-Paul Gonzalez, ©IRD).

by the fear generated by the epidemic and associated rumors.

Traditional practices regarding patient care and burial rituals often involve high risk conducts, such as washing and preparation of the body for exposure for several days, during which family and friends pay tribute by stroking or hugging the deceased [77-79,82,92]. Before appropriate measures are taken, which may take several weeks or months, the deceased persons are transported to their home community where people sometimes come from far away to attend the funeral and then go back home, which enables the spreading of the virus. The always frightening and often contradictory messages – and rumors – prompt patients to avoid going to the hospital due to fear of isolation and because of the lack of effective treatments. It becomes impossible to identify the cases, confirm diagnosis, protect and monitor contacts. Violent protests – with loss of life, involving sometimes the medical staff – have been reported in some outbreaks [71,82,92].

Other variables that hinder the adequate management of outbreaks include training of health workers who fail to immediately identify the disease, especially in areas where EVD has never been observed, either because of nonspecific clinical symptoms or lack of experience. Anyway, to date, there is no diagnostic test able to confirm an EVD suspicion. In addition, protective and disinfection equipment are usually absent and slow to become available in numerous health facilities [82]. Nosocomial transmission through contaminated and poorly (or not) sterilized medical instruments is a major multiplication factor in most outbreaks. Such picture is worsened by the fact that they hit in remote and poor regions.



Figure 8 Wrapping of the body in a mortuary sac (photo by Jean-Paul Gonzalez, ©IRD).

Social mobilization is a key component because all stakeholders should be involved to enable pooling resources and optimizing management of epidemics [71]. The ethical aspects should not be overlooked. Isolation of patients, required to avoid contamination, should not be seen as segregation. The family should be able to see and talk to patients, even if they are prevented from touching them. Authorities and medical staff should comply with, as far as possible, funeral rites by providing body bags and coffins for the families [92]. For instance, decontamination will be presented as ablutions that can be associated with the current ritual; deceased's clothes will be buried in the grave rather than burned to prevent stigmatization etc. [71,77-79].

After the epidemic, it is important to reseal the bonds of community through social, cultural and sporting activities.

Conclusion

Recently discovered in Central Africa, the Ebola viruses caused limited, but high-mortality epidemics. Fruit bats are probably the reservoir of the virus. The initial human infection results from contact with infected bush meat, and usually takes place in poor and inaccessible areas. Secondary transmission occurs during patient care at home, funeral rites or hospital dissemination due to the lack of preventive measures and because of inadequate training of health personnel with good medical practice.

The difficult diagnosis, the resistance of the population and the reluctance of authorities explain the slow and limited response to outbreaks, and the rapid spread of the epidemic that becomes hard to control.

In the absence of effective treatment, the discovery of an Ebola outbreak demands breaking the transmission



Figure 9 Boarding of a corpse in the vehicle for transportation to the burial site (photo by Jean-Paul Gonzalez, ©IRD).

chain at community and hospital levels, while respecting the traditions as much as possible, and the legitimate wish of relatives to accompany their sick or deceased relatives, in order to restore people confidence.

Received: 26 August 2014 Accepted: 19 September 2014

Published: 3 October 2014

References

1. Gordon Smith CE, Simpson DI, Bowen ET, Zlotnik I: **Fatal human disease from vervet monkeys.** *Lancet* 1967, **2**(7526):1119–1121.
2. Negredo A, Palacios G, Vázquez-Morón S, González F, Dopazo H, Molero F, Juste J, Quetglas J, Savji N, De La Cruz Martínez M, Herrera JE, Pizarro M, Hutchison SK, Echevarría JE, Lipkin WI, Tenorio A: **Discovery of an ebolavirus-like filovirus in Europe.** *PLoS Pathog* 2011, **7**(10):e1002304.
3. Kuhn JH, Bao Y, Bavari S, Becker S, Bradfute S, Brauburger K, Rodney Brister J, Bukreyev AA, Cai Y, Chandran K, Davey RA, Dolnik O, Dye JM, Enterlein S, Gonzalez JP, Formenty P, Freiberg AN, Hensley LE, Hoenen T, Honko AN, Ignat'yev GM, Jahrling PB, Johnson KM, Klenk HD, Kobinger G, Lackemeyer MG, Leroy EM, Lever MS, Mühlberger E, Netesov SV, et al: **Virus nomenclature below the species level: a standardized nomenclature for filovirus strains and variants rescued from cDNA.** *Arch Virol* 2014, **159**(5):1229–1237.
4. Ascenzi P, Bocedi A, Heptonstall J, Capobianchi MR, Di Caro A, Mastrangelo E, Bolognesi M, Ippolito G: **Ebolavirus and Marburgvirus: insight the Filoviridae family.** *Mol Aspects Med* 2008, **29**(3):151–185.
5. Feldmann H, Geisbert TW: **Ebola haemorrhagic fever.** *Lancet* 2011, **377**(9768):849–862.
6. Grosset A, Marzi A, Hoenen T, Herwig A, Gardner D, Becker S, Ebihara H, Feldmann H: **The Ebola virus glycoprotein contributes to but is not sufficient for virulence in vivo.** *PLoS Pathog* 2012, **8**(8):e1002847.
7. Ströher U, West E, Bugany H, Klenk HD, Schnittler HJ, Feldmann H: **Infection and activation of monocytes by Marburg and Ebola viruses.** *J Virol* 2001, **75**(22):11025–11033.
8. Basler CF, Wang X, Mühlberger E, Volchkov V, Paragas J, Klenk HD, Garcia-Sastre A, Palese P: **The Ebola virus VP35 protein functions as a type I IFN antagonist.** *Proc Natl Acad Sci U S A* 2000, **97**(22):12289–12294.
9. Kühl A, Pöhlmann S: **How Ebola virus counters the interferon system.** *Zoonoses Public Health* 2012, **59**(Suppl 2):116–131.
10. Carroll SA, Towner JS, Sealy TK, McMullan LK, Khristova ML, Burt FJ, Swanepoel R, Rollin PE, Nichol ST: **Molecular evolution of viruses of the family Filoviridae based on 97 whole-genome sequences.** *J Virol* 2013, **87**(5):2608–2616.
11. Johnson KM, Lange JV, Webb PA, Murphy FA: **Isolation and partial characterisation of a new virus causing acute haemorrhagic fever in Zaire.** *Lancet* 1977, **1**(8011):569–571.
12. Bowen ET, Lloyd G, Harris WJ, Platt GS, Baskerville A, Vella EE: **Viral haemorrhagic fever in southern Sudan and northern Zaire. Preliminary studies on the aetiological agent.** *Lancet* 1977, **1**(8011):571–573.
13. Pattyn S, van der Groen G, Courteille G, Jacob W, Piot P: **Isolation of Marburg-like virus from a case of haemorrhagic fever in Zaire.** *Lancet* 1977, **1**(8011):573–574.
14. Towner JS, Sealy TK, Khristova ML, Albariño CG, Conlan S, Reeder SA, Quan PL, Lipkin WI, Downing R, Tappero JW, Okware S, Lutwama J, Bakamutumaho B, Kayiwa J, Comer JA, Rollin PE, Ksiazek TG, Nichol ST: **Newly discovered Ebola virus associated with hemorrhagic fever outbreak in Uganda.** *PLoS Pathog* 2008, **4**(11):e1000212.
15. Le Guenno B, Formenty P, Wyers M, Gounon P, Walker F, Boesch C: **Isolation and partial characterisation of a new strain of Ebola virus.** *Lancet* 1995, **345**(8960):1271–1274.
16. Jahrling PB, Geisbert TW, Dalgard DW, Johnson ED, Ksiazek TG, Hall WC, Peters CJ: **Preliminary report: isolation of Ebola virus from monkeys imported to USA.** *Lancet* 1990, **335**(8688):502–505.
17. Barrette RW, Metwally SA, Rowland JM, Xu L, Zaki SR, Nichol ST, Rollin PE, Towner JS, Shieh WJ, Batten B, Sealy TK, Carrillo C, Moran KE, Bracht AJ, Mayr GA, Sirios-Cruz M, Catbagan DP, Lautner EA, Ksiazek TG, White WR, McIntosh MT: **Discovery of swine as a host for the Reston ebolavirus.** *Science* 2009, **325**(5937):204–206.
18. Leroy EM, Kumulungui B, Pourrut X, Rouquet P, Hassanin A, Yaba P, Délicat A, Paweska JT, Gonzalez JP, Swanepoel R: **Fruit bats as reservoirs of Ebola virus.** *Nature* 2005, **438**(7068):575–576.
19. Grard G, Biek R, Tamfum JJ, Fair J, Wolfe N, Formenty P, Paweska J, Leroy E: **Emergence of divergent Zaire ebola virus strains in Democratic Republic of the Congo in 2007 and 2008.** *J Infect Dis* 2011, **204**(Suppl 3):776–784.
20. Barrette RW, Xu L, Rowland JM, McIntosh MT: **Current perspectives on the phylogeny of Filoviridae.** *Infect Genet Evol* 2011, **11**(7):1514–1519.
21. Okware SI, Omaswa FG, Zaramba S, Opio A, Lutwama JJ, Kamugisha J, Rwaguma EB, Kagwa P, Lamunu M: **An outbreak of Ebola in Uganda.** *Trop Med Int Health* 2002, **7**(12):1068–1075.
22. Eichner M, Dowell SF, Firese N: **Incubation period of Ebola hemorrhagic virus subtype Zaire.** *Osong Public Health Res Perspect* 2011, **2**(1):3–7.
23. World Health Organization: **Ebola haemorrhagic fever: fact sheet revised in May 2004.** *Wkly Epidemiol Rec* 2004, **79**(49):435–439.
24. Baron RC, McCormick JB, Zubeir OA: **Ebola virus disease in southern Sudan: hospital dissemination and intrafamilial spread.** *Bull World Health Organ* 1983, **61**(6):997–1003.
25. World Health Organization: **Ebola haemorrhagic fever in Sudan, 1976. Report of a WHO/International Study Team.** *Bull World Health Organ* 1978, **56**(2):247–270.
26. Report of an International Commission: **Ebola haemorrhagic fever in Zaire, 1976.** *Bull World Health Organ* 1978, **56**(2):271–293.
27. Bwaka MA, Bonnet MJ, Calain P, Colebunders R, De Roo A, Guimard Y, Katwili KR, Kibadi K, Kipasa MA, Kuvula KJ, Mapanda BB, Massamba M, Mupapa KD, Muyembe-Tamfum JJ, Ndaberey E, Peters CJ, Rollin PE, Van den Enden E, Van den Enden E: **Ebola hemorrhagic fever in Kikwit, Democratic Republic of the Congo: clinical observations in 103 patients.** *J Infect Dis* 1999, **179**(Suppl 1):1–7.
28. Khan AS, Tshioko FK, Heymann DL, Le Guenno B, Nabeth P, Kerstiens B, Fleerackers Y, Kilmarx PH, Rodier GR, Nkuo O, Rollin PE, Sanchez A, Zaki SR, Swanepoel R, Tomori O, Nichol ST, Peters CJ, Muyembe-Tamfum JJ, Ksiazek TG: **The reemergence of Ebola hemorrhagic fever, Democratic Republic of the Congo, 1995.** *Commission de Lutte contre les Épidémies à Kikwit.* *J Infect Dis* 1999, **179**(Suppl 1):76–86.
29. MacNeil A, Farnon EC, Wamala J, Okware S, Cannon DL, Reed Z, Towner JS, Tappero JW, Lutwama J, Downing R, Nichol ST, Ksiazek TG, Rollin PE: **Proportion of deaths and clinical features in Bundibugyo Ebola virus infection.** *Uganda Emerg Infect Dis* 2010, **16**(12):1969–1972.
30. Roddy P, Howard N, Van Kerkhove MD, Lutwama J, Wamala J, Yoti Z, Colebunders R, Palma PP, Sterk E, Jeffs B, Van Herp M, Borchert M: **Clinical manifestations and case management of Ebola haemorrhagic fever caused by a newly identified virus strain, Bundibugyo, Uganda, 2007–2008.** *PLoS One* 2012, **7**(12):e52986.
31. Borchert M, Mutyaba I, Van Kerkhove MD, Lutwama J, Luwaga H, Bisoborwa G, Turyagaruka J, Pirard P, Ndayimirije N, Roddy P, Van Der Stuyft P: **Ebola haemorrhagic fever outbreak in Masindi District, Uganda: outbreak description and lessons learned.** *BMC Infect Dis* 2011, **11**:357.
32. Towner JS, Rollin PE, Bausch DG, Sanchez A, Crary S, Vincent M, Lee WF, Spiropoulou CF, Ksiazek TG, Lukwya M, Kaducu F, Downing R, Nichol ST: **Rapid diagnosis of Ebola haemorrhagic fever by reverse transcription PCR in an outbreak setting and assessment of patient viral load as a predictor of outcome.** *J Virol* 2004, **78**(8):4330–4341.
33. MacNeil A, Farnon EC, Morgan OW, Gould P, Boehmer TK, Blaney DD, Wiersma P, Tappero JW, Nichol ST, Ksiazek TG, Rollin PE: **Filovirus outbreak detection and surveillance: lessons from Bundibugyo.** *J Infect Dis* 2011, **204**(Suppl 3):761–767.
34. World Health Organization: **Outbreak(s) of Ebola haemorrhagic fever, Congo and Gabon, October 2001–July 2002.** *Wkly Epidemiol Rec* 2003, **78**(26):223–228.
35. Dowell SF, Mukunu R, Ksiazek TG, Khan AS, Rollin PE, Peters CJ: **Transmission of Ebola hemorrhagic fever: a study of risk factors in family members, Kikwit, Democratic Republic of the Congo, 1995.** *Commission de Lutte contre les Épidémies à Kikwit.* *J Infect Dis* 1999, **179**(Suppl 1):87–91.
36. Centers for Disease Control and Prevention [CDC]: **Ebola hemorrhagic fever.** *CDC* 2014, <http://www.cdc.gov/vhf/ebola/hcp/case-definition.html>.
37. Ksiazek TG, Rollin PE, Williams AJ, Bressler DS, Martin ML, Swanepoel R, Burt FJ, Leman PA, Khan AS, Rowe AK, Mukunu R, Sanchez A, Peters CJ: **Clinical virology of Ebola hemorrhagic fever (EHF): virus, virus antigen, and IgG and IgM antibody findings among EHF patients in Kikwit, Democratic Republic of the Congo, 1995.** *J Infect Dis* 1999, **179**(Suppl 1):177–187.

38. Sobarzo A, Ochayon DE, Lutwama JJ, Balinandi S, Guttman O, Marks RS, Kuehne AI, Dye JM, Yavelsky V, Lewis EC: **Lobel: Persistent immune responses after Ebola virus infection.** *N Engl J Med* 2013, **369**(5):492–493.
39. Fausther-Bovendo H, Mulangu S, Sullivan NJ: **Ebolavirus vaccines for humans and apes.** *Curr Opin Virol* 2012, **2**(3):324–329.
40. Hoenen T, Groseth A, Feldmann H: **Current ebola vaccines.** *Expert Opin Biol Ther* 2012, **12**(7):859–872.
41. MacNeil A, Rollin PE: **Ebola and Marburg hemorrhagic fevers: neglected tropical diseases?** *PLoS Negl Trop Dis* 2012, **6**(6):e1546.
42. Emond RT, Evans B, Bowen ET, Lloyd G: **A case of Ebola virus infection.** *Br Med J* 1977, **2**(6086):541–544.
43. Mupapa K, Massamba M, Kibadi K, Kuvula K, Bwaka A, Kipasa M, Colebunders R, Muyembe-Tamfum JJ: **Treatment of Ebola hemorrhagic fever with blood transfusions from convalescent patients.** International Scientific and Technical Committee. *J Infect Dis* 1999, **179**(Suppl 1):18–23.
44. Dye JM, Herbert AS, Kuehne AI, Barth JF, Muhammad MA, Zak SE, Ortiz RA, Prugar LI, Pratt WD: **Postexposure antibody prophylaxis protects nonhuman primates from filovirus disease.** *Proc Natl Acad Sci U S A* 2012, **109**(13):5034–5039.
45. Wong G, Qiu X, Olinger GG, Kobinger GP: **Post-exposure therapy of filovirus infections.** *Trends Microbiol* 2014, **22**(8):456–463.
46. Warren TK, Wells J, Panchal RG, Stuthman KS, Garza NL, Van Tongeren SA, Dong L, Retterer CJ, Eaton BP, Pegoraro G, Honnold S, Bantia S, Kotian P, Chen X, Taubenheim BR, Welch LS, Minning DM, Babu YS, Sheridan WP, Bavari S: **Protection against filovirus diseases by a novel broad-spectrum nucleoside analogue BCX4430.** *Nature* 2014, **508**:402–405.
47. Clark DV, Jahrling PB, Lawler JV: **Clinical management of filovirus-infected patients.** *Viruses* 2012, **4**:1668–1686.
48. Jahrling PB, Geisbert TW, Geisbert JB, Swearengen JR, Bray M, Jaax NK, Huggins JW, LeDuc JW, Peters CJ: **Evaluation of immune globulin and recombinant interferon-alpha2b for treatment of experimental Ebola virus infections.** *J Infect Dis* 1999, **179**(Suppl 1):224–234.
49. Changula K, Kajihara M, Mweene AS, Takada A: **Ebola and Marburg virus diseases in Africa: increased risk of outbreaks in previously unaffected areas?** *Microbial Immunol* 2014, **58**(9):483–491.
50. Groseth A, Feldmann H, Strong JE: **The ecology of Ebola virus.** *Trends Microbiol* 2007, **15**(9):408–416.
51. Polonsky JA, Wamala JF, de Clerck H, Van Herp M, Sprecher A, Porten K, Shoemaker T: **Emerging filoviral disease in Uganda: proposed explanations and research directions.** *Am J Trop Med Hyg* 2014, **90**(5):790–793.
52. Ivanoff B, Duquesnoy P, Languijat G, Saluzzo JF, Georges A, Gonzalez JP, McCormick J: **Haemorrhagic fever in Gabon. I. incidence of Lassa, Ebola and Marburg viruses in Haut-Ogooué.** *Trans R Soc Trop Med Hyg* 1982, **76**(6):719–720.
53. Johnson ED, Gonzalez JP, Georges AJ: **Filovirus activity among selected ethnic groups inhabiting the tropical forest of equatorial Africa.** *Trans R Soc Trop Med Hyg* 1993, **87**(5):536–538.
54. Germain M: **Collection of Mammals and Arthropods During the Epidemic of Haemorrhagic Fever in Zaire.** In *Ebola Virus Haemorrhagic Fever*. Edited by Pattyn SR. Amsterdam: Elsevier / North-Holland Biomedical Press; 1978:131–133.
55. Arata AA, Johnson B: **Approaches Towards Studies on Potential Reservoirs of Viral Haemorrhagic Fever in Southern Sudan (1977).** In *Ebola Virus Haemorrhagic Fever*. Edited by Pattyn SR. Amsterdam: Elsevier / North-Holland Biomedical Press; 1978:134–139.
56. Leirs H, Mills JN, Krebs JW, Childs JE, Akaibe D, Woollen N, Ludwig G, Peters CJ, Ksiazek TG: **Search for the Ebola virus reservoir in Kikwit, Democratic Republic of the Congo: reflections on a vertebrate collection.** *J Infect Dis* 1999, **179**(Suppl 1):S155–S163.
57. Morvan JM, Deubel V, Gounon P, Nakouné E, Barrière P, Murri S, Perpète O, Selekon B, Coudrier D, Gautier-Hion A, Colyn M, Volekho V: **Identification of Ebola virus sequences present as RNA or DNA in organs of terrestrial small mammals of the Central African Republic.** *Microbes Infect* 1999, **1**(14):1193–1201.
58. Leroy EM, Epelboin A, Mondonge V, Pourrut X, Gonzalez JP, Muyembe-Tamfum JJ, Formenty P: **Human Ebola outbreak resulting from direct exposure to fruit bats in Luebo, Democratic Republic of Congo, 2007.** *Vector Borne Zoonotic Dis* 2009, **9**(6):723–728.
59. Pourrut X, Kumulungui B, Wittmann T, Moussavou G, Délicat A, Yaba P, Nkoghe D, Gonzalez JP, Leroy EM: **The natural history of Ebola virus in Africa.** *Microbes Infect* 2005, **7**(7–8):1005–1014.
60. Pourrut X, Délicat A, Rollin PE, Ksiazek TG, Gonzalez JP, Leroy EM: **Spatial and temporal patterns of Zaire ebolavirus antibody prevalence in the possible reservoir bat species.** *J Infect Dis* 2007, **196**(Suppl 2):176–183.
61. Pourrut X, Souris M, Towner JS, Rollin PE, Nichol ST, Gonzalez JP, Leroy E: **Large serological survey showing cocirculation of Ebola and Marburg viruses in Gabonese bat populations, and a high seroprevalence of both viruses in *Rousettus aegyptiacus*.** *BMC Infect Dis* 2009, **9**:159.
62. Hayman DTS, Emmerich P, Yu M, Wang L-F, Suu-Ire R, Fooks AR, Cunningham AA, Wood JL: **Long-term survival of an urban fruit bat seropositive for Ebola and Lagos bat viruses.** *PLoS One* 2010, **5**(8):e11978.
63. Hayman DT, Yu M, Crameri G, Wang LF, Suu-Ire R, Wood JL, Cunningham AA: **Ebola virus antibodies in fruit bats, Ghana.** *West Africa Emerg Infect Dis* 2012, **18**(7):1207–1209.
64. Reiter P, Turell M, Coleman R, Miller B, Maupin G, Liz J, Kuehne A, Barth J, Geisbert J, Dohn D, Glick J, Pecor J, Robbins R, Jahrling P, Peters C, Ksiazek T: **Field investigations of an outbreak of Ebola hemorrhagic fever, Kikwit, Democratic Republic of the Congo, 1995: arthropod studies.** *J Infect Dis* 1999, **179**(Suppl 1):S148–S154.
65. Olson SH, Reed P, Cameron KN, Ssebide BJ, Johnson CK, Morse SS, Karesh WB, Mazet JA, Joly DO: **Dead or alive: animal sampling during Ebola hemorrhagic fever outbreaks in humans.** *Emerg Health Threats J* 2012, **5**: doi: 10.3402/ehtj.v50.9134.
66. Formenty P, Boesch C, Wyers M, Steiner C, Donati F, Dind F, Walker F, Le Guenno B: **Ebola virus outbreak among wild chimpanzees living in a rain forest of Côte d'Ivoire.** *J Infect Dis* 1999, **179**(Suppl 1):S120–S126.
67. Georges AJ, Leroy EM, Renaut AA, Benissan CT, Nabias RJ, Ngoc MT, Obiang PI, Lepage JP, Bertherat EJ, Bénoni DD, Wickings EJ, Amblard JP, Lansoud-Soukate JM, Milleliri JM, Baize S, Georges-Courbot MC: **Ebola hemorrhagic fever outbreaks in Gabon, 1994–1997: epidemiologic and health control issues.** *J Infect Dis* 1999, **179**(Suppl 1):S65–S75.
68. Karesh W, Reed P: **Ebola and great apes in Central Africa: current status and future needs.** *Bull Soc Pathol Exot* 2005, **98**(3):237–238.
69. Nkoghe D, Formenty P, Leroy EM, Nngue S, Edou SY, Ba JI, Allarangar Y, Cabore J, Bachy C, Andraghetti R, de Benoit AC, Galanis E, Rose A, Bausch D, Reynolds M, Rollin P, Choueibou C, Shongo R, Geronne B, Koné LM, Yada A, Roth C, Mve MT: **Plusieurs épidémies de fièvre hémorragique à virus Ebola au Gabon, octobre 2001 à avril 2002.** *Bull Soc Pathol Exot* 2005, **98**(3):224–229.
70. Nkoghe D, Kone ML, Yada A, Leroy E: **A limited outbreak of Ebola haemorrhagic fever in Etoumbi, Republic of Congo, 2005.** *Trans R Soc Trop Med Hyg* 2011, **105**(8):466–472.
71. Formenty P, Libama F, Epelboin A, Allarangar Y, Leroy E, Moudzeo H, Tarangonia P, Molamou A, Lenzi M, Ait-Ikhlef K, Hewlett B, Roth C, Grein T: **La riposte à l'épidémie de fièvre hémorragique à virus Ebola en République du Congo, 2003: une nouvelle stratégie?** *Med Trop* 2003, **63**:291–295.
72. Thomas DW: **The annual migrations of three species of West African fruit bats (Chiroptera: Pteropodidae).** *Can J Zoology* 1983, **61**(10):2266–2272.
73. Richter HV, Cumming GS: **First application of satellite telemetry to track African straw-coloured fruit bat migration.** *J Zool* 2008, **275**(2):172–176.
74. Johnson BK, Wambui C, Ocheng D, Gichogo A, Oogo S, Libondo D, Gitau LG, Tuwei PM, Johnson ED: **Seasonal variation in antibodies against Ebola virus in Kenyan fever patients.** *Lancet* 1986, **1**(8490):1160.
75. Pinzon JE, Wilson JM, Tucker CJ, Arthur R, Jahrling PB, Formenty P: **Trigger events: enviroclimatic coupling of Ebola hemorrhagic fever outbreaks.** *Am J Trop Med Hyg* 2004, **71**(5):664–674.
76. Gautier-Hion A, Michaloud G: **Are figs always keystone resources for tropical frugivorous vertebrates? A test in Gabon.** *Ecology* 1989, **70**(6):1826–1833.
77. Hewlett BS, Amola RP: **Cultural contexts of Ebola in northern Uganda.** *Emerg Infect Dis* 2003, **9**(10):1242–1248.
78. Hewlett BS, Epelboin A, Hewlett BL, Formenty P: **Medical anthropology and Ebola in Congo: cultural models and humanistic care.** *Bull Soc Pathol Exot* 2005, **98**(3):230–236.
79. Epelboin A: **Rapport de mission anthropologique sur l'épidémie d'Ebola Isiro, R. D. Congo, 4 au 30 septembre 2012.** France: CNRS-MNHN Paris & OMS; 2012 [download2.cerimes.fr/canalu/documents/smm/marburg.en.angola.uige.april.2005.fun.railles.de.crise.le.tailleur.et.les.siens_13719/2012rdc.ebola.rapport.epelboinww.pdf].
80. Bausch DG, Towner JS, Dowell SF, Kaducu F, Lukwya M, Sanchez A, Nichol ST, Ksiazek TG, Rollin PE: **Assessment of the risk of Ebola virus transmission from bodily fluids and fomites.** *J Infect Dis* 2007, **196**(Suppl 2):S142–S147.

81. Kibadi K, Mupapa K, Kuvula K, Massamba M, Ndalberey D, Muyembe-Tamfum JJ, Bwaka MA, De Roo A, Colebunders R: Late ophthalmologic manifestations in survivors of the 1995 Ebola virus epidemic in Kikwit, Democratic Republic of the Congo. *J Infect Dis* 1999, **179**(Suppl 1):S13–S14.
82. Milleliri JM, Tevi-Benissan C, Baize S, Leroy E, Georges-Courbot MC: Épidémies de fièvre hémorragique à virus Ebola au Gabon (1994–2002): aspect épidémiologiques et considérations sur les mesures de contrôle. *Bull Soc Pathol Exot* 2004, **97**:199–205.
83. Heymann DL, Weisfeld JS, Webb PA, Johnson KM, Cairns T, Berquist H: Ebola hemorrhagic fever: Tandala, Zaire, 1977–1978. *J Infect Dis* 1980, **142**(3):372–376.
84. Amblard J, Obiang P, Edzang S, Prehaud C, Bouloy M, Le Guenno B: Identification of the Ebola virus in Gabon in 1994. *Lancet* 1997, **349**(9046):181–182.
85. World Health Organization: Ebola haemorrhagic fever: confirmed case in Côte d'Ivoire and suspect cases in Liberia. *Wkly Epidemiol Rec* 1995, **70**(50):359.
86. World Health Organization: Ebola haemorrhagic fever in Côte d'Ivoire. *Wkly Epidemiol Rec* 1995, **70**(21–52):367.
87. World Health Organization: Ebola haemorrhagic fever, Zaire. *Wkly Epidemiol Rec* 1995, **70**(34):241–242.
88. World Health Organization: Ebola haemorrhagic fever, Gabon. *Wkly Epidemiol Rec* 1996, **71**(9):71.
89. World Health Organization: Outbreak of Ebola haemorrhagic fever in Gabon officially declared over. *Wkly Epidemiol Rec* 1996, **71**(17):125–126.
90. World Health Organization: Ebola haemorrhagic fever: a summary of the outbreak in Gabon. *Wkly Epidemiol Rec* 1997, **72**(1–2):342.
91. Georges-Courbot MC, Lu CY, Lansoud-Soukate J, Leroy E, Baize S: Isolation and partial molecular characterisation of a strain of Ebola virus during a recent epidemic of viral haemorrhagic fever in Gabon. *Lancet* 1997, **349**(9046):181.
92. Boumambouki P, Formenty P, Epelboin A, Campbell P, Atsangandoko C, Allarangar Y, Leroy EM, Kone ML, Molamou A, Dinga-Longa O, Salemo A, Kounkou RY, Mombouli V, Ibara JR, Gaturuku P, Nkunku S: Prise en charge des malades et des défunts lors de l'épidémie de fièvre hémorragique à virus Ebola à Mbandza et Mbomo d'octobre à décembre 2003 au Congo. *Bull Soc Pathol Exot* 2005, **98**(3):218–223.
93. World Health Organization: Outbreak of Ebola haemorrhagic fever in Yambio, south Sudan, April - June 2004. *Wkly Epidemiol Rec* 2005, **80**(43):370–375.
94. World Health Organization: Ebola haemorrhagic fever, Uganda - end of the outbreak. *Wkly Epidemiol Rec* 2008, **83**(10):89–90.
95. World Health Organization: Ebola, Uganda. *Wkly Epidemiol Rec* 2011, **86**(22):221.
96. World Health Organization: Uganda: Ebola (situation as of 27 August 2012) 2012. <http://www.afro.who.int/en/clusters-a-programmes/dpc/epidemic-a-pandemic-alert-and-response/outbreak-news/3674-uganda-ebola-situation-as-of-27-august-2012.html>.
97. Parkes-Ratanshi R, Elbireer A, Mbambu B, Mayanja F, Coutinho A, Merry C: Ebola outbreak response; experience and development of screening tools for viral haemorrhagic fever (VHF) in a HIV center of excellence near to VHF epicentres. *PLoS One* 2014, **9**(7):e100333.
98. World Health Organization: Ebola haemorrhagic fever, Uganda - update. *Wkly Epidemiol Rec* 2012, **87**(49–50):493.
99. World Health Organization Regional Office for Africa: DR Congo: Ebola. *Bull World Health Organ* 2012, **2**:8.
100. Mukwamba FKN: Déclaration de la fin de l'épidémie de fièvre hémorragique à virus Ebola survenue dans le District Sanitaire du Haut-Uélé Ouest, Province Orientale, RD Congo.: Government of the Democratic Republic of the Congo; 2012. <http://reliefweb.int/report/democratic-republic-congo/d%C3%A9clarer-de-la-fin-de-l%20E2%80%99%C3%A9pid%C3%A9se-de-fi%C3%A8re-h%C3%A9morragique-%C3%A0-virus>.
101. World Health Organization: Suspected viral haemorrhagic fever outbreaks in Sudan and Zaire. *Wkly Epidemiol Rec* 1976, **51**(41):321.
102. World Health Organization: Viral haemorrhagic fever. Zaire. *Wkly Epidemiol Rec* 1977, **52**(1):2.
103. World Health Organization: Viral haemorrhagic fever. Sudan. *Wkly Epidemiol Rec* 1979, **54**(41):319.
104. World Health Organization: Yellow fever. Gabon. *Wkly Epidemiol Rec* 1995, **70**(9):64.
105. Centers for Disease Control and Prevention: Outbreak of Ebola hemorrhagic fever – Uganda, August 2000–January 2001. *Morb Mortal Wkly Rep* 2001, **50**(5):73–77.
106. Heymann DL, Barakamitiye D, Szczepanowski M, Muyembe-Tamfum JJ, Bele O, Rodier G: Ebola hemorrhagic fever: lessons from Kikwit, Democratic Republic of the Congo. *J Infect Dis* 1999, **179**(Suppl 1):S283–S286.

doi:10.1186/1678-9199-20-44

Cite this article as: Chippaux: Outbreaks of Ebola virus disease in Africa: the beginnings of a tragic saga. *Journal of Venomous Animals and Toxins including Tropical Diseases* 2014 **20**:44.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

