

Nutritional Status and Body Composition in Patients With Hepatic Glycogen Storage Diseases Treated With Uncooked Cornstarch—A Controlled Study

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Abstract

Hepatic glycogen storage diseases (GSDs) are genetic diseases associated with fasting hypoglycemia. Periodic intake of uncooked cornstarch is one of the treatment strategies available for those disorders. For reasons that are still not clear, patients with hepatic GSDs may be overweight. **Aims:** To assess nutritional status and body composition in patients with hepatic GSDs receiving uncooked cornstarch. **Methods:** The sample included 25 patients with hepatic GSD (type Ia = 14; Ib = 6; III = 3; IX α = 1; IX β = 1), with a median age of 11.0 years (interquartile range [IQR] = 9.0–17.5), matched by age and gender with 25 healthy controls (median age = 12.0 years, IQR = 10.0–17.5). Clinical, biochemical, and treatment-related variables were obtained from medical records. Nutritional status and body composition were prospectively evaluated by bioelectrical impedance. **Results:** Patients and controls did not differ with regard to age and gender. Height was significantly reduced in patients (median = 1.43 m, IQR = 1.25–1.54) in comparison to controls (median = 1.54 m, IQR = 1.42–1.61; $P = .04$). Body mass index for age z-score and fat mass percentage were higher in patients (median = 1.84, IQR = 0.55–3.06; and 27.5%, IQR = 22.6–32.0, respectively) than in controls (median = 0.86, IQR = –0.55 to 1.82; $P = .04$ and 21.1%, IQR = 13.0–28.3; $P = .01$, respectively). When patients were stratified by type, those with GSD Ia had significantly higher adiposity (median fat mass = 28.7%, IQR = 25.3–32.9) than those with GSD III and GSD IX α / β (median fat mass = 20.9%, IQR = 14.9–22.6; $P = .02$). **Conclusions:** Our findings suggest that patients with hepatic GSD on treatment with cornstarch, especially those with GSD Ia, exhibit abnormalities in nutritional status and body composition, such as short stature and a trend toward overweight and obesity.

Keywords

hepatic glycogen storage diseases, nutritional status, body composition, adiposity, bioelectrical impedance

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Introduction

The glycogen storage diseases (GSDs), or glycogenosis, are a group of genetic diseases characterized by abnormalities in enzymes that regulate glycogen synthesis and degradation.¹ Glycogen, stored mainly in muscle and liver tissue, serves as a readily accessible source of energy² to maintain glucose homeostasis during fasting.³

Different GSD types exist and are classified accordingly to the organs affected, with a broad range of clinical manifestations, and gene/enzyme involved (Table 1). In hepatic GSDs, endogenous glucose production is impaired; thus, the main clinical consequence is fasting hypoglycemia.¹ Hepatomegaly is another common manifestation shared across nearly all GSD types. Symptoms vary depending on the specific form of GSD including hyperlactatemia, hyperlipidemia, hyperuricemia, cardiomyopathy, abnormal liver enzymes, hypotonia, and cirrhosis.^{1,2,10,11}

The primary treatment goal in hepatic GSDs is to prevent hypoglycemia, avoid long-term complications, and ensure adequate growth.¹² One of the treatment approaches is dietary and may involve frequent, periodic administration of uncooked cornstarch (UCCS) and continuous nocturnal gastric drip feeding (CNGDF).¹³ Liver transplantation can be considered in cases of severe hepatic cirrhosis, hepatic dysfunction, and/or hepatocellular carcinoma,⁷ and kidney transplantation when there is renal insufficiency.¹⁰

Considering the clinical heterogeneity of hepatic GSD, dietary recommendations need to be adapted to the needs of each type of diseases and the age of the patient. The nutritional recommendations for GSD I, III, and IX are presented in Table 2. Daily caloric intake should be prescribed and monitored closely, as insufficient energy provision will be unable to correct metabolic derangements and, consequently, leads to

Table 1. Classification of Hepatic Glycogen Storage Diseases and Summary of Their Characteristics.^a

Type (MIM)	Enzyme Involved	Gene	Inheritance Pattern	Clinical/Biochemical	Treatment
0, liver (240600)	Glycogen synthase (liver)	<i>GYS2</i>	AR	Fasting ketotic hypoglycemia; postprandial hyperglycemia, hyperlactatemia, and hyperlipidemia. No liver enlargement.	Protein-rich diet, low-glycemic index complex carbohydrates; bedtime uncooked cornstarch.
Ia (232200)	Glucose-6-phosphatase	<i>G6PC</i>	AR	Hypoglycemia, hepatomegaly, growth retardation, lactic acidosis, hyperuricemia, hyperlipidemia.	Uncooked cornstarch; galactose, fructose, lactose, and sucrose restriction.
Ib (232220)	Glucose-6-phosphate translocase	<i>SLC37A4</i>	AR	As in Ia, plus neutropenia, recurrent infections, inflammatory bowel disease.	Uncooked cornstarch; galactose, fructose, lactose, and sucrose restriction. Granulocyte colony-stimulating factor (filgrastim).
IIIa and IIIb (232400)	Glycogen debranching enzyme	<i>AGL</i>	AR	Hepatomegaly, hyperketotic hypoglycemia; growth retardation, hyperlipidemia, elevated AST, ALT, CPK. Muscle weakness and cardiomyopathy occur in subtype IIIa.	Uncooked cornstarch; protein-rich diet; sucrose restriction.
IV (232500)	Glycogen branching enzyme	<i>GBE1</i>	AR	Hepatomegaly, growth retardation, cirrhosis.	Liver transplantation in severe cases.
VI (232700)	Glycogen phosphorylase (liver)	<i>PYGL</i>	AR	Hepatomegaly, growth retardation; mild hypoglycemia, hyperlipidemia, hyperketosis.	If symptomatic: increased carbohydrate intake, frequent feedings, protein-rich diet.
IX α 1 and IX α 2 (306000)	Phosphorylase kinase	<i>PHKA2</i>	XLR	Hepatomegaly, fasting ketotic hypoglycemia, growth retardation, mild AST/ALT elevation, and hyperlipidemia.	Uncooked cornstarch; protein-rich diet; avoidance of large amounts of sucrose.
IX β (261750)	Phosphorylase kinase	<i>PHKB</i>	AR	As in IXa.	Uncooked cornstarch; protein-rich diet; avoidance of large amounts of sucrose.
IX γ (613027)	Phosphorylase kinase	<i>PHKG2</i>	AR	As in IXa, plus cirrhosis.	Uncooked cornstarch; protein-rich diet; avoidance of large amounts of sucrose.
XI (227810)	Glucose transporter 2	<i>GLUT2</i>	AR	Hypoglycemia, failure to thrive, rickets, protuberant abdomen due to enlarged liver and kidneys.	Restricted galactose intake; uncooked cornstarch; supplementation of water, electrolytes, and vitamin D.

Abbreviations: AR, autosomal recessive; XLR, X-linked recessive; GYS2, Glycogen synthase 2; G6PC, Glucose-6-phosphatase catalytic subunit; SLC37A4, Solute carrier family 37 member 4; AGL, Amylo-alpha-1, 6-glycosidase, 4-alpha-glucanotransferase; GBE1, 1,4-alpha-glucan branching enzyme I; PYGL, Glycogen phosphorylase L; PHKA2, Phosphorylase kinase regulatory subunit alpha 2; PHKB, Phosphorylase kinase regulatory subunit beta; PHKG2, Phosphorylase kinase catalytic subunit gamma 2; GLUT2, Solute carrier family 2 member 2; AR, autosomal recessive; XLR, X-linked recessive; AST, Aspartate Aminotransferase; ALT, Alanine Aminotransferase; CPK, Creatine Phosphokinase.

^aAdapted from Wolfsdorf and Weinstein,¹ Beauchamp et al,⁴ Kishnani et al,⁵ Hicks et al,⁶ Dagli et al,⁷ Chen et al,⁸ Kishnani et al,⁹ and Bali et al.¹⁰

Table 2. Dietary Recommendations for Hepatic GSD I, III, and IX.^a

Type	Age Group	UCCS (g/kg/dose)	Dose Interval of UCCS (h)	CARB (%)	PTN (%)	LIP (%)	Additional Recommendations
I	Infants	1.6	3-4	60-70	10-15	The remainder of %kcal ^b	Restriction of lactose and fructose
	Small children-puberty	1.7-2.5	4-6				
	Adults	1.7-2.5	Variable				
III	>12 months	1.0-2.5	4-6	35-55	20-30	20-35	Meal 3/3 h up to 12 months. High-protein diet (3 g/kg)
	Adults	Variable	Variable				
IX	All	0.6-2.5	Variable	^c	15-25	^c	High-protein diet

Abbreviations: CARB, carbohydrate; GSD, glycogen storage disorder; LIP, lipid; PTN, protein; UCCS, uncooked cornstarch.

^aAdapted from Kishnani et al,⁵ Hicks et al,⁶ Dagli et al,⁷ Kishnani et al,⁹ Bali et al,¹⁰ Froissart et al,¹² Goldberg and Slonim,¹⁴ and Sentner et al.¹⁵

^b<30% for children over 2 years.

^cSimilar to recommendations for GSD III.

growth impairment. On the other hand, excessive UCCS intake may lead to obesity.¹²

There are several reports of overweight in patients with hepatic GSDs, particularly in GSD Ia,^{9,16,17} however, the etiology of this finding remains incompletely elucidated. On the one hand, dietary treatment with a complex carbohydrate can reduce the risk of short-term and long-term complications,¹⁸ on the other hand, it could be associated with overweight and, eventually, excess adiposity. Short stature has also been reported in GSD types I, III, and IX α/β .^{2,4,6,11,19}

Although changes in nutritional status appear to be frequent among patients with hepatic GSDs, knowledge gaps remain regarding these aspects as well as regarding the body composition of these patients. Within this context, the present study sought to evaluate nutritional status and body composition in patients with hepatic GSDs in comparison to healthy controls, through a review of clinical data and bioelectrical impedance analysis (BIA), in an attempt to improve the current understanding of the association of these aspects with GSD.

Methods

Study Design

This cross-sectional study used a convenience sampling strategy to enroll patients with hepatic GSDs and healthy controls matched for age and gender. All patients were recruited from the Medical Genetics Service at Hospital de Clínicas de Porto Alegre (SGM-HCPA), Brazil. The 25 healthy controls had a median age of 12.0 years (interquartile range [IQR] = 10.0-17.5) and were recruited from the community of the same hospital. This study was approved by the HCPA Research Ethics Committee (protocol number: 14-0120) and conducted in accordance with the provisions of the Declaration of Helsinki. All participants or their legal guardians signed an informed consent term.

Sample

The sample included 25 patients receiving UCCS treatment, with a median age of 11.0 years (IQR = 9.0-17.5), of whom

Table 3. Nutritional Status and Body Composition in Patients With Hepatic Glycogenosis Receiving Uncooked Cornstarch Therapy and Controls—Summary of the Main Findings.^a

	GSD (n = 25)	Controls (n = 25)	P Value
Gender (F/M)	13/12	13/12	–
Age (years)	11.0 (9.0 to 17.5)	12.0 (10.0 to 17.5)	.45
Weight (kg)	49.0 (29.8 to 66.9)	50 (36.2 to 62.5)	.95
Height (m)	1.43 (1.25 to 1.54)	1.54 (1.42 to 1.61)	.04 ^b
BMI (kg/m ²)			
≤ 19 years	22.5 (18.3 to 26.0)	20.3 (16.3 to 25.2)	.22
> 19 years	28.2 (25.0 to 38.9)	21.9 (19.9 to 23.5)	.01 ^b
Phase angle (°)	6.2 (5.4 to 7.0)	6.5 (5.6 to 7.1)	.40
Fat-free mass (%)	72.5 (68.0 to 77.3)	78.9 (71.7 to 87.0)	.01 ^b
Fat mass (%)	27.5 (22.6 to 32.0)	21.1 (13.0 to 28.3)	.01 ^b
Height for age (z)	–1.31 (–1.92 to 0.13)	–0.06 (–0.63 to 0.58)	<.01 ^b
Weight for age (z)	0.07 (–0.78 to 2.02)	0.22 (–0.56 to 0.86)	.86
BMI-for-age (z)	1.84 (0.55 to 3.06)	0.86 (–0.55 to 1.82)	.04 ^b

Abbreviations: BMI, body mass index; F/M, female/male; GSD, glycogen storage disease.

^aFat mass expressed as percentage of total body weight. Data expressed as median (interquartile range).

^bStatistically significant.

14 had GSD Ia, 6 had GSD Ib, 3 had GSD III, and 2 had GSD IX α/β (Tables 3 and 4). All patients included in this study had a diagnosis of hepatic GSD confirmed by enzymatic and/or genetic analysis (median age at diagnosis = 8.5 months; IQR = 6-21.5) and are seen at the Outpatient Metabolic Disorders Clinic at SGM-HCPA (median age at treatment onset = 12 months; IQR = 6-35.2) every 3 to 6 months. Clinical variables (GSD type and comorbidities), biochemical parameters (serum lactate, glucose, triglycerides, and cholesterol levels), and treatment parameters (dose of UCCS administered) were obtained through a review of the medical records for the patients' latest visits (up to 3 months before inclusion in the study).

Table 4. Nutritional Status and Body Composition According to the Type of Hepatic Glycogenosis—Summary of the Main Findings (N = 25).^a

	GSD Ia (n = 14)	GSD Ib (n = 6)	GSD III or IX α/β (n = 5)	P Value
Gender (F/M)	9/5	3/3	1/4	–
Age (years)	13.5 (9.7 to 18.2)	11.0 (5.2 to 18.5)	9.0 (8.5 to 12.0)	.24
UCCS/day (g)	383.0 (287.5 to 510.0)	332.5 (228.0 to 438.7)	288.0 (207.0 to 376.0)	.36
UCCS/weight (g/kg)	5.98 (4.82 to 8.12)	7.61 (6.38 to 11.29)	9.46 (4.88 to 11.44)	.20
Weight (kg)	56.0 (41.3 to 87.1)	43.8 (18.1 to 68.9)	30.0 (27.0 to 50.3)	.17
Height (m)	1.49 (1.34 to 1.56)	1.37 (1.04 to 1.52)	1.29 (1.24 to 1.52)	.37
BMI (kg/m ²)	24.9 (22.0 to 33.1)	23.2 (18.1 to 28.0)	19.0 (16.7 to 21.1)	.06
Fat-free mass (%)	71.3 (67.0 to 74.6) ^b	69.8 (63.0 to 77.6)	79.1 (77.3 to 85.0) ^b	.02 ^c
Fat mass (%)	28.7 (25.3 to 32.9) ^b	30.1 (22.4 to 36.9)	20.9 (14.9 to 22.6) ^b	.02 ^c
Phase angle (°)	6.65 (5.32 to 7.27)	5.40 (4.42 to 6.55)	6.20 (5.95 to 6.90)	.21
Height for age (z)	–1.17 (–1.82 to 0.11)	–1.78 (–2.05 to –0.46)	–0.91 (–1.93 to 0.94)	.53
Weight for age (z)	0.62 (–1.92 to 2.10)	0.55 (–0.78 to 1.52)	–0.18 (–1.07 to 1.85)	.99
BMI for age (z)	2.54 (0.44 to 3.59)	2.50 (1.01 to 3.01)	0.97 (0.22 to 1.50)	.33

Abbreviations: BMI, body mass index; F/M, female/male; GSD, glycogen storage disease; UCCS, uncooked cornstarch.

^aFat mass expressed as percentage of total body weight. Data expressed as median (interquartile range).

^bDenotes significant between-group difference.

^cStatistically significant.

Anthropometric Measurements

Weight was measured using digital scales (resolution 0.1 kg; Model 2096PP/2; Toledo, São Paulo, Brazil), while height was measured with a wall-mounted stadiometer (precision 0.1 cm; Harpenden; Holtain, Crymych, Wales, United Kingdom). These measurements were obtained while participants were in standing position and wearing minimal clothing. Body mass index (BMI) was calculated as weight (kg) divided by height in meter square (m²) and classified as underweight, normal weight, overweight, or obese, per the World Health Organization (WHO) criteria.^{20,21} In patients aged <19 years, nutritional status was calculated using BMI for age z scores calculated in WHO Anthro version 3.2.2 and WHO Anthroplus version 1.0.4.^{22,23}

Bioelectrical Impedance Analysis of Body Composition

To estimate body composition (fat mass and fat-free mass), a BIA system was used (Biodynamics 450, version 5.1; Biodynamics Corporation, Seattle, Washington), with resting-tab electrocardiogram electrodes (Conmed Corporation, Utica, New York), as described elsewhere in the literature. Participants were instructed to refrain from physical activity or intake of caffeine-enriched foods and beverages in the 8 hours preceding assessment as well as to empty their bladders and remove all metal objects before starting the test.²⁴ Due to the need to maintain normal glucose levels, patients could not be asked to fast for 8 hours. Therefore, a 3-hour fast was defined as standard for all patients. Controls underwent the same preparation and were given the same instructions but with an 8-hour fast. During the test, participants remained in the supine position, with the limbs outstretched away from the body, while 4 electrodes were applied in the following distribution: 1 on the wrist, 1 on the hand, 1 on the ankle, and 1 on the foot (all on the right side of the body). The adiposity (percentage fat mass) was

calculated according to the Obesity Medicine Association²⁵ reference values for adults and McCarthy et al²⁶ reference values for children. Bioelectrical impedance analysis also provided data on phase angle (PA), a variable derived from the relationship between resistance (R), reactance (Xc), and impedance (Z) ($PA = \arctan(Xc/R)$) to the passage of a low-amplitude, high-frequency (50 kHz) electrical current during the procedure.²⁷ The cutoff of Bony-Westphal et al²⁸ was used to evaluate PA.

Statistical Analysis

All analyses were carried out using SPSS Statistics, version 21.0 (IBM Corp, Armonk, New York). Categorical variables were expressed as absolute and relative frequencies, and continuous variables, as medians and IQRs. The Mann-Whitney *U* test was used for comparison between the case and control groups. The Kruskal-Wallis test, followed by a post hoc analysis, was used for comparison between subgroups of patients with different GSD types. Spearman coefficients were used to test for correlation. The significance level was set at 5%.

Results

The 25 patients included belonged to 24 unrelated families. The rate of parental consanguinity was 20.8%. Four patients (16%) had clinical diagnosis of inflammatory bowel disease; all had a diagnosis of GSD type Ib and were overweight or obese at the time of assessment. Three patients (12%), all with GSD Ia, were being treated for hypothyroidism; of these, 2 (67%) had excess weight and 1 had normal weight.

Median daily UCCS intake was 6.48 g/kg body weight (IQR = 5.11–9.97), for a median total of 360 g/day (IQR = 265–447). The UCCS doses were administered every 3 to 6 hours. The median total cholesterol and triglyceride levels in plasma, available for 24 patients, were 167.5 mg/dL (IQR =

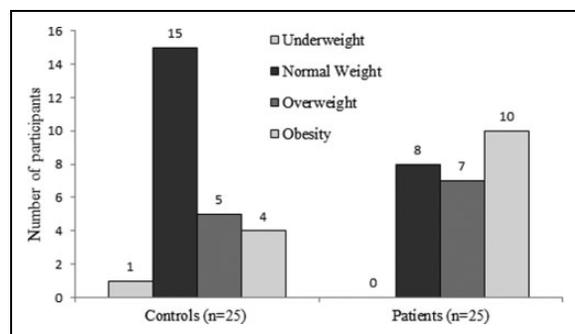


Figure 1. Classification of the nutritional status of patients with hepatic glycogen storage disease and controls, according to body mass index.^{20,21}

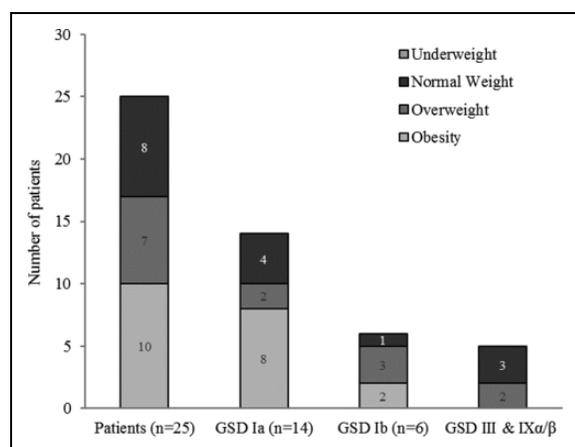


Figure 2. Classification of the nutritional status of patients with hepatic glycogen storage disease, according to body mass index.^{20,21}

146.2-206.0) and 197.0 mg/dL (IQR = 119.7-407.5), respectively. The median serum glucose level was 86.5 mg/dL (IQR = 80.5-95.2, $n = 22$), while the median lactate level was 1.78 mmol/dL (IQR = 1.22-2.69, $n = 20$).

Table 3 reports anthropometric parameters and body composition data for patients and controls. Bioelectrical impedance analysis revealed that 16 (64%) patients (3 with normal weight, 3 overweight, and 10 obese) and 7 (28%) controls (1 with normal weight, 2 overweight, and 4 obese) had excess adiposity. Eight (32%) patients (6 with GSD Ia and 2 with GSD Ib) and 5 controls (20%) had PA values below the cutoff. Phase angle correlated strongly with fat-free mass in controls ($r_s = 0.7$; $P < .01$) and moderately in patients ($r_s = 0.4$; $P = .03$). Figure 1 shows the distribution of nutritional status in the sample, stratified by BMI classification.^{20,21} High rates of excess weight (overweight = 28%; obesity = 40%) were found among patients. Conversely, most controls had normal weight (60%), while 9 had excess weight (overweight = 20%; obesity = 16%).

Stratification of patients by GSD type revealed a significant difference only for fat mass percentage; in this sample, patients with GSD Ia had greater adiposity than those with GSD III or IXα/β (Table 4).

Regarding stratification of nutritional status by GSD type (Figure 2), we found a heterogeneous distribution of BMI values: 2 of 5 patients with GSD III and IXα/β were overweight, while 5 out of 6 with GSD Ib and 10 out of 14 with GSD Ia had excess weight (overweight or obesity).

Discussion and Conclusion

This was the first study to evaluate body composition by BIA in patients with hepatic GSD on treatment with cornstarch. We chose BIA for evaluation of body composition because it is a noninvasive, low-cost, user-friendly method widely employed in clinical practice. Bioelectrical impedance analysis is a predictive technique for assessment of body composition whereby the passage of a low-amplitude, high-frequency electrical current through the body, particularly through the lowest-resistance compartment, allows direct measurement of resistance and reactance values, which are used to calculate impedance and PA; finally, it estimates total body water content, extracellular and intracellular water, fat-free mass, and fat mass.²⁹⁻³¹

In a review, Andreoli et al³² suggested that, compared to other techniques, BIA is a good tool for estimation of body composition in isovolemic patients. The estimated error for prediction of total body water content and fat-free mass by BIA is 2% to 4%. To achieve optimal precision or even improve the precision of this method, it is essential that measurement conditions be standardized.³³

It is well-known that nonfasting state interferes with BIA findings. However, patients with GSD cannot be deprived of food for long periods without risking hypoglycemia. According to Kyle et al,²⁴ food and drink intake 2 to 4 hours before the test can reduce body impedance by 4 to 15 Ω, which represents an error of less than 3%. Thus, although an 8-hour fast is recommended, shorter periods can be acceptable both in clinical practice and in research settings.

A high frequency of obesity in patients with GSD was found by Chen et al¹⁶ in a study of 13 GSD I patients in treatment (UCCS or CNGDF) for more than 5 years. The results of this evaluation showed that 10 of 13 patients were classified as obese according to percentage of ideal body weight for length percentile. Similar findings were described by Santos et al¹⁷ in a cross-sectional study that evaluated patients with GSD I treated with UCCS, z scores of BMI for age showed that 16 of 21 patients were overweight. Short stature has also been reported by several authors as common problems in the hepatic GSD.^{2,4,6,11,19}

In agreement with the literature, our findings showed that most patients had excess weight and adiposity, with higher BMI, BMI for age z scores, and body fat percentage than controls as well as lower median height than controls. Although anthropometric parameters revealed a stature deficit in the patient group, as in the Weinstein and Wolfsdorf¹⁸ study, the median z score for height (-1.31 ; IQR = -1.92 to 0.13) was within the expected range according to WHO standards.²¹ The pathophysiology of short stature in hepatic GSDs has yet to be

elucidated. However, studies suggest that the carbohydrate overload for treatment can contribute not only to excess weight^{5,10,12,34} but also to better height scores in these patients.^{17,18} Although our treated patients had a short stature, suggesting that the use of UCCS does not heal the low height.

Comparison of body composition revealed that fat mass was significantly greater in patients than in controls overall and was significantly greater in patients with GSD Ia than in patients with GSD III and IX α / β ($P = .02$). Patients with GSD III and IX α / β were those who most resembled controls in terms of fat mass percentage.

Body mass index is widely used for assessment of nutritional status, but the underestimation of obesity is a major limitation of this method.³⁵ In 1 study, BIA was used to evaluate body composition in 1244 European children, from 2004 to 2006. The authors concluded that, in the pediatric population, fat-free mass and fat mass measurements provide a better understanding of growth and changes in body composition than BMI.³⁶ This discrepancy was also observed in our sample, in which excess adiposity was identified even in some patients classified as having normal weight by BMI.

Patients with GSD Ia, Ib, and III/IX were not found to differ regarding the daily amount of UCCS received (in g or in g/kg), but patients with GSD Ia were found to have higher fat mass than patients with GSD III/IX. So, although many authors suggest that excess weight and body fat in patients with GSD are associated with UCCS excessive intake,^{5,10,12,34} our findings did not confirm this hypothesis. Another factor that may have been associated with excess weight, although not evaluated in this study, is the low engagement in physical activity. Uncertainty to whether physical activity is allowed and what dietary changes would be required may contribute to higher rates of sedentary behavior in this population.

Another BIA parameter that plays an important role in assessment of nutritional status is PA.^{37–39} A positive correlation between PA and fat-free mass has been described both in healthy individuals and in pathological conditions.^{40–42} We observed similar correlation in our control sample but not in patients with hepatic GSD, despite statistical significance ($P = .03$). Therefore, we suggest that PA is not a good predictor of fat-free mass in hepatic GSDs.

This parameter has also been associated with inflammation⁴³ and may be useful in the assessment of patients with hepatic GSDs, particularly GSD Ib, because of their greater susceptibility to inflammatory conditions.⁴⁴ There was no difference in PA between patients and controls, however, the median PA was lower in patients with GSD Ib than in patients with other types of GSD, suggesting possible predictive value in this subpopulation.

In conclusion, our findings suggest that Brazilian patients with hepatic GSDs have a shorter median height than healthy controls but still within normal range, and a tendency toward overweight and obesity, as demonstrated both by BMI (especially in patients with GSD type I) and by BIA-measured adiposity. Prospective studies with larger samples, taking into account measurement of anthropometric parameters as well

as adiposity, may contribute to the further elucidation of the impact of these conditions on nutritional status and height. Furthermore, we believe greater research attention should be given to achieving a better understanding of the heterogeneous impacts of different GSD types on nutritional status, which could help prevent comorbidities associated with excess weight and adiposity and, thus, improve patient's quality of life.

Declaration of Conflicting Interests

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References

1. Wolfsdorf J, Weinstein DA. Glycogen storage diseases. *Rev Endocr Metab Disord*. 2003;4(1):95-102.
2. Gazzero E, Andreu AL, Bruno C. Neuromuscular disorders of glycogen metabolism. *Curr Neurol Neurosci Rep*. 2013;13(3):333.
3. Bhattacharya K. Investigation and management of the hepatic glycogen storage diseases. *Transl Pediatr*. 2015;4(2):240-248.
4. Beauchamp NJ, Dalton A, Ramaswami U, et al. Glycogen storage disease type IX: high variability in clinical phenotype. *Mol Genet Metab*. 2007;92(1-2):88-99.
5. Kishnani PS, Austin SL, Arn P, et al. Glycogen storage disease type III diagnosis and management guidelines. *Genet Med*. 2010;12(7):446-463.
6. Hicks J, Wartchow E, Mierau G. Glycogen storage diseases: a brief review and update on clinical features, genetic abnormalities, pathologic features, and treatment. *Ultrastruct Pathol*. 2011;35(5):183-196.
7. Dagli A, Sentner CP, Weinstein DA. Glycogen storage disease type III. 2016. <http://www.ncbi.nlm.nih.gov/books/NBK26372/>. Accessed September 12, 2017.
8. Chen Y, Kishnani PS, Koeberl D. Glycogen storage diseases In: Valle D, Beaudet AL, Vogelstein B, et al. eds. *The Online Metabolic and Molecular Bases of Inherited Disease*. 2014; chapter 71.
9. Kishnani PS, Austin SL, Abdenur JE, et al; American College of Medical Genetics and Genomics. Diagnosis and management of glycogen storage disease type I: a practice guideline of the American college of medical genetics and genomics. *Genet Med*. 2014;16(11):e1.
10. Bali DS, Chen Y, Goldstein JL. Glycogen storage disease type I. 2016. <http://www.ncbi.nlm.nih.gov/books/NBK1312/>. Accessed September 12, 2017.
11. Özen H. Glycogen storage diseases: new perspectives. *World J Gastroenterol*. 2007;13(18):2541-2553.
12. Froissart R, Piraud M, Boudjemline AM, et al. Glucose-6-phosphatase deficiency. *Orphanet J Rare Dis*. 2011;6:27.

13. Chou JY, Jun HS, Mansfield BC. Glycogen storage disease type I and G6Pase- β deficiency: etiology and therapy. *Nat Rev Endocrinol*. 2010;6(12):676-688.
14. Goldberg T, Slonim AE. Nutrition therapy for hepatic glycogen storage diseases. *J Am Diet Assoc*. 1993;93(12):1423-1430.
15. Sentner CP, Hoogeveen IJ, Weinstein DA, et al. Glycogen storage disease type III: diagnosis, genotype, management, clinical course and outcome. *J Inherit Metab Dis*. 2016;39(5):697-704.
16. Chen YT, Bazzarre CH, Lee MM, Sidbury JB, Coleman RA. Type I glycogen storage disease: nine years of management with cornstarch. *Eur J Pediatr*. 1993;152(suppl 1):S56-S59.
17. Santos BL, Souza CF, Schuler-Faccini L, et al. Glycogen storage disease type I: clinical and laboratory profile. *J Pediatr*. 2014;90(6):572-579.
18. Weinstein DA, Wolfsdorf JI. Effect of continuous glucose therapy with uncooked cornstarch on the long-term clinical course of type Ia glycogen storage disease. *Eur J Pediatr*. 2002;161(suppl 1):S35-S39.
19. Rake JP, Visser G, Labrune P, Leonard JV, Ullrich K, Smit GP. Glycogen storage disease type I: diagnosis, management, clinical course and outcome. Results of the European Study on Glycogen Storage Disease type I (ESGSD I). *Eur J Pediatr*. 2002;161(suppl 1):S20-S34.
20. World Health Organization. *Physical Status: The Use and Interpretation of Anthropometry*. Report of a WHO Expert Committee. Geneva, Switzerland: WHO; 1995.
21. World Health Organization. *WHO Child Growth Standards: Methods and Development: Length/Height-for-Age, Weight-for-Age, Weight-for-Length, Weight-for-Height and Body Mass Index-for-Age*. Geneva, Switzerland: WHO; 2006.
22. World Health Organization. *WHO AnthroPlus for Personal Computers Manual: Software for Assessing Growth of the World's Children and Adolescents*. Geneva, Switzerland: WHO; 2009.
23. World Health Organizations. *WHO Anthro for Personal Computers Manual: Software for Assessing Growth and Development of the World's Children*. Geneva, Switzerland: WHO; 2010.
24. Kyle UG, Bosaeus I, De Lorenzo AD, et al. Bioelectrical impedance analysis—part II: utilization in clinical practice. *Clin Nutr*. 2004;23:1430-1453.
25. Obesity Medicine Association. Obesity Algorithm®: Clinical Guidelines for Obesity Treatment. 2016. <https://obesitymedicine.org/obesity-algorithm/>. Accessed July 01, 2017.
26. McCarthy HD, Cole TJ, Fry T, Jebb SA, Prentice AM. Body fat reference curves for children. *Int J Obes*. 2006;30:598-602.
27. Kyle UG, Bosaeus I, De Lorenzo AD, et al. Bioelectrical impedance analysis—part I: review of principles and methods. *Clin Nutr*. 2004;23(5):1226-1243.
28. Bosy-Westphal A, Danielzik S, Dörhöfer RP, Later W, Wiese S, Müller MJ. Phase angle from bioelectrical impedance analysis: population reference values by age, sex, and body mass index. *J Parenter Enteral Nutr*. 2006;30(4):309-316.
29. Wells JCK, Fewtrell MS. Measuring body composition. *Arch Dis Child*. 2006;91(7):612-617.
30. Associação Médica Brasileira, Conselho Federal de Medicina. *Utilização da Bioimpedância para Avaliação da Massa Corporórea*. Projeto Diretrizes. 2009.
31. Fosbøl MØ, Zerahn B. Contemporary methods of body composition measurement. *Clin Physiol Funct Imaging*. 2015;35(2):81-97.
32. Andreoli A, Garaci F, Cafarelli FP, Guglielmi G. Body composition in clinical practice. *Eur J Radiol*. 2006;85(8):1461-1468.
33. Kushner RF, Gudivaka R, Schoeller DA. Clinical characteristics influencing bioelectrical impedance analysis measurements. *Am J Clin Nutr*. 1996;64(suppl 3):423S-427S.
34. Derks TGJ, Martens DH, Sentner CP. Dietary treatment of glycogen storage disease type Ia: uncooked cornstarch and/or continuous nocturnal gastric drip-feeding? *Mol Genet Metab*. 2013;109(1):1-2.
35. Peltz G, Aguirre MT, Sanderson M, Fadden MK. The role of fat mass index in determining obesity. *Am J Hum Biol*. 2010;22(5):639-647.
36. Rush E, Reed PW, McLennan S, Coppinger T, Simmons D, Graham D. Tracking of body mass indices over 2 years in Māori and European children. *Eur J Clin Nutr*. 2012;66(2):143-149.
37. Nagano M, Suita S, Yamanouchi T. The validity of bioelectrical impedance phase angle for nutritional assessment in children. *J Pediatr Surg*. 2000;35(7):1035-1039.
38. Barbosa-Silva MC, Barros AJ. Bioelectrical impedance analysis in clinical practice: a new perspective on its use beyond body composition equations. *Curr Opin Clin Nutr Metab Care*. 2005;8(3):311-317.
39. Kyle UG. Can phase angle determined by bioelectrical impedance analysis assess nutritional risk? a comparison between healthy and hospitalized subjects. *Clin Nutr*. 2012;31(6):875-881.
40. Selberg O, Selberg D. Norms and correlates of bioimpedance phase angle in healthy human subjects, hospitalized patients, and patients with liver cirrhosis. *Eur J Appl Physiol*. 2002;86(6):509-516.
41. Visser M, van Venrooij LM, Wanders DC, et al. The bioelectrical impedance phase angle as an indicator of undernutrition and adverse clinical outcome in cardiac surgical patients. *Clin Nutr*. 2012;31(6):981-986.
42. Gonzalez MC, Barbosa-Silva TG, Bielemann RM, Gallagher D, Heymsfield SB. Phase angle and its determinants in healthy subjects: influence of body composition. *Am J Clin Nutr*. 2016;103(3):712-716.
43. Stobäus N, Pirlich M, Valentini L, Schulzke JD, Norman K. Determinants of bioelectrical phase angle in disease. *Br J Nutr*. 2012;107(8):1217-1220.
44. Chou JY, Jun HS, Mansfield BC. Neutropenia in type Ib glycogen storage disease. *Curr Opin Hematol*. 2010;17(1):36-42.