

Research Article

The FMR1 premutation as a cause of premature ovarian failure in Brazilian women

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Abstract

The loss-of-function mutation of the *FMR1* gene due to expansion of the 5' UTR CGG repeat causes the fragile X syndrome, the most frequent form of inherited mental retardation. On the other hand, the *FMR1* premutation, which is transcriptionally active and produces the protein, confers an increased risk for premature ovarian failure (POF) to carrier females. Among 41 unrelated Brazilian women with idiopathic POF, we found three carriers of premutations (CGG expansionse \geq 59 repeats) and two carriers of high-intermediate alleles (50-55 repeats). Two premutations and two intermediate alleles were detected among the 16 familial POF cases, and one premutated woman, among the 25 sporadic cases. The premutation frequency among the familial cases (12.5%) differed significantly from that found in a control group of 96 unrelated Brazilian women aged \geq 47 years, who had not experience POF and in which no premutations or high-intermediate alleles were detected. In the search for factors influencing the probability of a premutation carrier presenting POF, another 20 unrelated premutated women with POF, from fragile X families, were included in the study. The analysis of the *FMR1*-linked loci DXS548 and FRAXAC1 did not indicate any association of a particular haplotype with the occurrence of POF. An effect of X-inactivation skewing was not apparent in blood cells, and POF-associated premutations showed a wide range of repeat sizes, from 59, the smallest known to expand to full mutations upon transmission to offspring, to approximately 200.

Key words: premature ovarian failure, menopause, FMR1 premutation, fragile X syndrome.

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Introduction

The fragile X syndrome is the most frequent single cause of inherited mental retardation, with an estimated incidence of 1 in 4,000 males and 1 in 8,000 females (Crawford *et al.*, 2001). In most cases, the loss-of-function mutation, which causes this syndrome, consists of an expansion of the polymorphic CGG trinucleotide repeat in the 5' untranslated region of the *FMR1* gene at Xq27.3. This fully mutated allele is abnormally hypermethylated and is not transcribed (Fu *et al.*, 1991; Oberlé *et al.*, 1991; Pieretti *et al.*, 1991; Verkerk *et al.*, 1991; Nolin *et al.*, 2003). Alleles with 6 to ~55 triplets are considered to be normal, those with 29-30 triplets being the most common in the general population. Alleles with repeats in the ~55-200 range, the premutations, are transcribed and the protein (FMRP) is

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produced, but these alleles are unstable and may expand to full mutations upon maternal transmission. The smallest allele known to have expanded to a full mutation had a 59-triplet repeat (Nolin *et al.*, 2003). However, the boundary between normal and premutation alleles is not well defined, and constitutes a "gray zone" containing highnormal and low-premutation alleles (Eichler *et al.*, 1994). These intermediate alleles are defined as those varying between 41~60 repeats, which may or may not be transmitted in an unstable manner (Murray *et al.*, 1997). Their tendency to expand is directly related to the size of the repeat (Nolin *et al.*, 2003).

Premature ovarian failure (POF), defined as the cessation of ovarian function before the age of 40, affects approximately 1% of women in the general population (Coulam *et al.*, 1986). It has been described both in patients with X chromosome abnormalities, mostly deletions, and with a normal karyotype. A familial pattern of premature ovarian failure, suggesting autosomal or X-linked inheritance, has been reported, but few genes causing POF have

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424 Costa et al.

been identified so far. DIAPH2 at Xg22, one of the human homologues of the Drosophila melanogaster diaphanous gene, was found disrupted by a balanced translocation in a mother and a daughter with POF (Bione et al., 1998). Mutations of FSHR, the follicle-stimulating hormone receptor gene at 2p21, which led to partial loss of function of the protein, were described in secondary amenorrhoea (Beau et al., 1998). Mutations of this receptor gene were previously reported as the cause of primary ovarian failure (Aittomaki et al., 1996). FOXL2, the forkhead transcription factor gene at 3q23, which is mutated in blepharophimosis, ptosis and epicanthus inversus syndrome with or without ovarian failure, had novel mutations described that caused only POF (Harris et al., 2002). Ovarian failure was also associated to mutations in E1F2B, the eukaryotic translation initiation factor 2B, in patients with leukodystrophy, a condition termed ovarioleukodystrophy (Fogli et al., 2003). However, given that more than 30% of all POF cases are familial, there must be many more POF-causing alleles awaiting discovery. Accordingly, it was a breakthrough when a significant association between the fragile X premutations and POF was demonstrated both by the analysis of women carrying premutations, and by the screening of women affected by POF (Cronister et al., 1991; Schwartz et al., 1994; Conway et al., 1995, Vianna-Morgante et al., 1996; Murray et al., 1998; Allingham-Hawkins et al., 1999; Vianna-Morgante et al., 1999; Marozzi et al., 2000).

Although no doubt remains that the fragile X premutation is associated with POF, it has been evident since the first studies that the presence of the premutation is not deterministic for POF to occur and, according to The International Collaborative POF in Fragile X Study (Allingham-Hawkins et al., 1999), about 24% of premutation carriers in fragile X families experience POF. However, some data suggest that POF is at the extreme of the spectrum of the premutation effects on ovarian function. The age at menopause of premutated women who did not experience POF was shown to be significantly lower than the menopausal age of their non-carrier relatives (Vianna-Morgante et al., 1999). Pointing in the same direction are the increased serum follicle-stimulating hormone concentrations, a marker of late ovarian failure, observed in premutation carriers (Braat et al., 1999; Murray et al., 1999), even in those which are on oral contraceptive (Hundscheid et al., 2001). Factor(s) causing a woman to exceed a threshold and manifest POF are presently unknown. In fact, the nature of the POF/premutation association remains elusive. The absence of FMRP does not make a woman more likely to present POF, since fully mutated women do not tend to experience POF (Allingham-Hawkins et al., 1999; Vianna-Morgante et al., 1999). Premutation carriers produce an excess of FMR1 mRNA, probably as a response to the impairment of FMRP synthesis that seems to be correlated with the size of the repeat expansion (Tassone et al., 2000). It is speculated that this mRNA excess could be toxic to the cells and interfere with normal ovary function. A direct effect of the expansion itself abnormally recruiting RNAbinding proteins, thus generally impeding the availability of these factors, has been considered as an explanation for the neurological impairment (FXTAS syndrome) presented by some male carriers of the FMR1 premutation (Hagerman and Hagerman, 2004). The same mechanism is apparently at the basis of another trinucleotide-expansion disease, myotonic dystrophy (Day and Ranum, 2005). Another possibility is that the premutation expansion affects the expression of neighboring genes involved in ovary function (Vianna-Morgante et al., 1996). Under these hypotheses, the size of the expanded repeat could be a factor influencing the manifestation of POF. Indeed, a significant positive association of the FMR1 repeat size with ovarian dysfunction was detected in a recent study of a large sample of women with repeats ranging from common to premutation sizes (Sullivan et al., 2005). Another possibility was raised some years ago, in a study of Dutch fragile-X families showing that POF occurred predominantly in carrier women who inherited the premutation from their fathers (Hundscheid et al., 2000). However, this parent-of-origin effect was not confirmed, either in the United Kingdom (Murray et al., 2000), or by us in Brazil (Vianna-Morgante and Costa, 2000). It has been suggested that reduced fitness of premutated women with POF could have influenced these studies differently, selection being more effective against mother-daughter pairs in a population where women reproduce later (Sherman, 2000), and actually at the time Hundscheid et al. (2000) conducted their study, the Netherlands had the oldest maternal age of any other coun-

Herein we report the investigation of the premutation/POF association in 41 unrelated Brazilian women ascertained because they presented with POF, and in a further 20 premutated women belonging to Brazilian fragile X families, who had experienced POF. We determined the frequency of the premutation among women ascertained by POF, and showed that the *FMR1* premutation is an important single cause of POF in our population, especially in POF familial cases. We failed to identify predisposing factors influencing POF manifestation in *FMR1* premutation carriers.

Subjects and Methods

Patients and controls

Premature ovarian failure (POF) was considered as the cessation of menstruation for at least one year before the age of 40. A total of 41 unrelated women ascertained by idiopathic POF were studied to determine the frequency of the *FMR1* premutation among them. Sixteen of them were familial cases of POF and 25 occurred sporadically. Thirty-two of the women were attending the Outpatient Climacteric Clinic, at the Department of Obstetrics and Gy-

necology of the University of São Paulo Medical School. The nine other women with menopause were referred from different medical services in the State of São Paulo. The control group included 96 normal unrelated women who had not experienced POF before age 47, who attended the same clinic at the Medical School, on a routine basis. In the search for factors related to the manifestation of POF, we also included in the analysis 20 unrelated premutation carriers with POF, ascertained in fragile X families referred to the Genetic Counseling Service of the Department of Genetics and Evolutionary Biology at the University of São Paulo. For X-inactivation studies, a control group formed by 53 premutated women without POF belonging to the same fragile X families was used. This study was approved by the Ethics Committee of the Hospital das Clínicas, University of São Paulo Medical School (269/99).

Methods

DNA was extracted from peripheral blood lymphocytes. Screening for FRAXA premutations was performed by PCR, with primers c and f (Fu et al., 1991), followed by electrophoresis on sequencing acrylamide gels, as previously described by Kenneson et al. (1996), with slight modifications (Mingroni-Netto et al., 2002). When only one allele was detected, in order to distinguish between homozygosis and heterozygosis for a non-amplified expanded allele, Southern blotting was carried out using doubly digested EcoR I/Eag I fragments probed with StB12.3, as previously described (Mingroni-Netto et al., 1994). The X-inactivation patterns were studied by determining the activation ratios (the proportion of active normal alleles) through densitometry comparison of the allele bands on the auto-radiographs (BIORAD GS 700 Image Densitometer, Molecular Analyst/BC). In order to investigate if a particular subgroup of premutations was associated with POF, the microsatellite loci DXS548 and FRAXAC1, which are tightly linked to the FMR1 gene (Richards et al., 1991; Riggins et al., 1992), were genotyped by PCR, according to previously described procedures (Mingroni-Netto et al., 1994). The allele nomenclature followed Macpherson et al. (1994).

Results

No premutations or intermediate alleles in the upper range of the distribution (\geq 50 repeats) were found in the

control group of 96 women. Among the 41 unrelated women referred because of POF, three (7.32%) were diagnosed as premutation carriers (p = 0.03; Fisher's exact test). Among the 16 familial cases of POF, two women had the premutation (12.5%; p = 0.02), and one premutated woman was detected among the 25 isolated cases (4%; p = 0.21) (Table 1). Two high-intermediate alleles, with 50 and 52 repeats, were found in the familial cases.

In the study of factors influencing POF manifestation, we included 20 unrelated women with a premutation and POF, who were detected in fragile X families (Table 2). So, a total of 23 women carrying the premutation were genotyped for the *FMR1*-linked microsatellite loci DXS548 and FRAXAC1. The haplotype linked to the premutation could be identified in women from the fragile X families by genotyping affected males in the families: the haplotypes 2-1 and 6-4, the most frequently linked to fragile X chromosomes, were found in eight and four women, respectively (Table 2). In the three carriers ascertained through POF, the DXS548 and FRAXAC1 alleles were determined, but haplotypes could not be inferred (Table 1).

We determined the activation ratios - the proportion of active normal alleles - in blood cells in all 23 premutated women with POF, and in 53 premutated women who did not experience POF. The mean activation ratios were 48.59 and 45.60, respectively, not differing statistically (p = 0.56; Mann-Whitney test).

Discussion

Among the 41 women ascertained by POF, the frequency of the FMR1 premutation (7.3%) was significantly higher than in the control group, in which premutations were not detected. The difference was also significant when we analyzed the 16 familial cases separately (12.5%). On the other hand, the 4% frequency of premutations among the 25 sporadic cases did not reach significance relative to the controls. These results confirm the FMR1 premutation as an important causative factor of POF. Two previous studies on series of women with POF point in the same direction. In the United Kingdom, the screening of 147 women with idiopathic POF for the fragile X premutation, revealed six premutation carriers (4.1%), four among 25 women with familial POF (16%), and two others among 122 women with sporadic POF (1.6%) (Murray et al., 1998). In Italy, six carriers (6%) were found among 106

Table 1 - Premutation carriers ascertained by POF.

Patients	Menopause age (years)	CGG-rep	eat sizes	DXS548	FRAXAC1
		Premutated alleles	Normal alleles	Alleles	
Familial POF	35	86	23	2-7	1-3
Familial POF	30	87	30	6-7	3-4
Sporadic POF	39	59	29	2-7	1-3

426 Costa et al.

Table 2 - Premutation carriers with POF ascertained in fragile X families.

Patients	Age at menopause (years)	Recurrence of POF	$(CGG)_n$		DXS548/FRAXAC1 haplotypes	
			Normal alleles	Premutated alleles*	Fra(X) chromosome	Normal chromosome
F1	31	Familial	23	*	8-3	8-4
F2	28	Sporadic	30	*	7-4	7-3
F3	35	Sporadic	20	*	7-4	7-3
F4	30	Sporadic	20	*	5-3	7-3
F5	29	Sporadic	29	*	6-4	9-3
F6	30	Sporadic	30	*	6-4	9-3
F7	13	Sporadic	20	*	6-4	7-3
F8	37	Sporadic	26	75	2-1	7-3
F9	34	Sporadic	20	*	6-4	6-3
F10	17	Familial	42	99	2-3	2-1
F11	35	Familial	21	*	2-1	6-3
F12	38	Sporadic	30	*	2-3	7-3
F13	14	Familial	30	*	2-1	7-3
F14	35	Familial	30	*	2-1	2-3
F15	28	Sporadic	29	*	2-1	2-4
F16	37	Familial	30	*	2-1	7-3
F17	38	Familial	30	*	?-3	7-3
F18	34	Sporadic	32	*	2-1	7-3
F19	25	Familial	20	*	2-1	7-3
F20	36	Sporadic	23	110	7-4	7-3

^{*}Premutation diagnosed by Southern blotting.

women with POF, four among 33 familial cases (12%), and two among 73 sporadic cases (3%) (Marozzi *et al.*, 2000). On the other hand, in a study conducted in the USA (Kenneson *et al.*, 1997), none of 33 women who experienced ovarian failure under the age of 40 years (17 familial and 16 sporadic POF cases) carried the premutation. Taken together, these studies point to the significance of the *FMR1* premutation as a cause of POF, especially when this condition is familial. Indeed, the overall frequency of the premutation in these series of POF women is clearly dependent on the proportion of familial cases.

We also investigated some factors that could influence the manifestation of POF associated with the *FMR1* premutation. The pattern of silencing of the premutation by X-inactivation could influence the occurrence of POF in carriers, the amount of active premutations having to reach a threshold for the effect to appear. We did not observe a significant difference between the activation ratios in blood cells of premutated women who experienced POF and of those who did not. However, this possibility cannot be ruled out, since the pattern of X-inactivation in the ovary might not be the same as in blood. The size of the premutation could also influence the manifestation of POF. In our sample, the measure of the expansion was tentative, from Southern blots, since the size of the repeat could not be pre-

cisely determined in every premutation carrier, due to the inherent difficulties in amplifying these GC-rich segments. This prevented the performing of a correlation analysis between premutation sizes and ages at menopause. However, it is noteworthy that the sizes of the premutations in women experiencing POF encompass a wide range, from as small as 59 up to near 200 trinucleotides.

POF associated with the *FMR1* premutation appears to cluster in some families (Vianna-Morgante et al., 1996), while in others none of the carrier women have POF. Considering that peculiarities of certain premutations could explain these clusters, we genotyped the DXS548 and FRAXAC1 loci in the search for a linkage disequilibrium involving certain haplotypes and the POF-associated premutations. No trend in this direction was observed, since the distribution of the haplotypes in premutation women with POF did not characterize their families as a subset of fragile X families. Indeed, 60% of the premutation carriers with POF had the haplotypes most commonly observed on the fragile X chromosomes in Brazilian families (Mingroni-Netto et al., 1999) as well as in other populations (Chiurazzi et al., 1996). Therefore, no indication was found that the POF-associated premutations had a specific origin or resided on a particular haplotype background.

In conclusion, this work confirms in Brazilian women the *FMR1* premutation as the most frequent single cause of POF identified to date and should be considered in the diagnosis of women experiencing POF. It should be noted that all the premutations that we ascertained through POF were detected in families without any history of mental retardation. Genetic counseling and diagnostic tests for possible carriers could be offered, aiming at the prevention of mental retardation.

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References

- Aittomaki K, Herva R, Stenman U-H, Juntunen K, Ylostalo P, Hovatta O and de la Chapelle A (1996) Clinical features of primary ovarian failure caused by a point mutation in the follicle-stimulating hormone receptor gene. J Clin Endocr Metab 81:3722-3726.
- Allingham-Hawkins DJ, Babul-Hirji R, Chitayat D, Holden JJA, Yang KT, Lee C, Hudson R, Gorwill H, Sarah L, Nolin SL, Glicksman A, Jenkins EC, Brown WT, Howard-Peebles PN, Becchi C, Cummings E, Fallon L, Seitz S, Black SH, Vianna-Morgante AM, Costa SS, Otto PA, Mingroni-Netto RC, Murray A, Webb J, MacSwinney F, Dennis N, Jacobs PA, Syrrou M, Georgiou I, Patsalis PC, Uzielli MLG, Guarducci S, Lapi E, Cecconi A, Ricci U, Ricotti G, Biondi C, Scarselli B and Vieri F (1999) Fragile X premutation is a significant risk factor for premature ovarian failure The International Collaborative POF in Fragile X Study Preliminary data. Am J Med Genet 83:322-325.
- Beau I, Touraine P, Meduri G, Gougeon A, Desroches A, Matuchansky C, Milgrom E, Kuttenn F and Misrahi M (1998) A novel phenotype related to partial loss of function mutations of the follicle stimulating hormone receptor. J Clin Invest 102:1352-1359.
- Bione S, Sala C, Manzini C, Arrigo G, Zuffardi O, Banfi S, Borsani G, Jonveaux P, Philippe C, Zuccotti M, Ballabio A and Toniolo D (1998) A human homologue of the *Drosophila melanogaster* diaphanous gene is disrupted in a patient with premature ovarian failure: Evidence for conserved function in oogenesis and implications for human sterility. Am J Hum Genet 62:533-541.
- Braat DD, Smits AP and Thomas CM (1999) Menstrual disorders and endocrine profiles in fragile X carriers prior to 40 years of age: A pilot study Am J Med Genet 83:327-328.
- Chiurazzi P, Genuardi M, Kozak L, Giovannucci-Uzielli ML, Bussani C, Dagna-Bricarelli F, Grasso M, Perroni L, Sebastio G, Sperandeo MP, Oostra BA and Neri G (1996) Fragile X founder chromosome in Italy: A few initial events and possible explanation for their heterogeneity. Am J Med Genet 64:209-215.
- Conway GS, Hettiarachchi S, Murray A and Jacobs PA (1995) Fragile X premutations in familial premature ovarian failure. Lancet 346:309-310.
- Coulam CB, Adamson SC and Annegers JF (1986) Incidence of premature ovarian failure. Obstet Gynecol 67:604-606.

- Crawford DC, Acuna JM and Sherman SL (2001) *FMR1* and the fragile X syndrome: Human genome epidemiology review. Genet Med 3:359-371.
- Cronister A, Schreiner R, Wittenberger M, Amiri K, Harris K and Hagerman RJ (1991) Heterozygous fragile X female: Historical, physical, cognitive, and cytogenetic features. Am J Med Genet 38:269-274.
- Day JW and Ranum LPW (2005) RNA pathogenesis of the myotonic dystrophies. Neurom Disord 15:5-16.
- Eichler EE, Holden JJ, Popovich BW, Reiss AL, Snow K, Thibodeau SN, Richards CS, Ward PA and Nelson DL (1994) Length of uninterrupted CGG repeats determines instability in the *FMR1* gene. Nat Genet 8:88-94.
- Fogli A, Rodriguez D, Eymard-Pierre E, Bouhour F, Labauge P, Meaney BF, Zeesman S, Kaneski CR, Schiffmann R and Boespflug-Tanguy O (2003) Ovarian failure related to eukaryotic initiation factor 2B mutations. Am J Hum Genet 72:1544-1550.
- Fu YH, Kuhl DPA, Pizzuti A, Pieretti M, Sutcliffe JS, Richards S, Verkerk AJMH, Holden JJA, Fenwick Jr RG, Warren ST, Ostra BA, Nelson DL and Caskey CT (1991) Variation of the CGG repeat at the fragile X site results in genetic instability: Resolution of the Sherman paradox. Cell 67:1047-1058
- Hagerman PJ and Hagerman RJ (2004) The fragile-X premutation: A maturing perspective. Am J Hum Genet 74:805-816
- Harris SE, Chand AL, Winship IM, Gersak K, Aittomaki K and Shelling AN (2002) Identification of novel mutations in *FOXL2* associated with premature ovarian failure. Molec Hum Reprod 8:729-733, 2002.
- Hundscheid RDL, Braat DDM, Kiemeney LALM, Oostra BA, Smits APT and Thomas CMG (2001) Increased serum FSH in female fragile X premutation carriers with either regular menstrual cycles or on oral contraceptives. Hum Reprod 16:457-462.
- Hundscheid RDL, Sistermans EA, Thomas CMG, Braat DDM, Straatman H, Kiemeney LALM, Oostra BA and Smits APT (2000) Imprinting effect in premature ovarian failure confined to paternally inherited fragile X premutations. Am J Hum Genet 66:413-418.
- Kenneson A, Cramer DW and Warren ST (1997) Fragile X premutations are not a major cause of early menopause. Am J Hum Genet 61:1362-1369.
- Macpherson JN, Bullman H, Yuings SA and Jacobs PA (1994) Insert size and flanking haplotype in fragile X and normal populations: Possible multiple origins for the fragile X mutation Hum Mol Genet 3:399-405.
- Marozzi A, Vegetti W, Manfredini E, Tibiletti G, Testa G, Crosignani PG, Ginelli E, Meneveri R and Dalprà L (2000) Association between idiopathic premature ovarian failure and fragile X premutation. Hum Reprod 15:197-202.
- Mingroni-Netto RC, Angeli CB, Auricchio MT, Leal-Mesquita ER, Ribeiro-Dos-Santos AK, Ferrari I, Hutz MH, Salzano FM, Hill K, Hurtado AM and Vianna-Morgante AM (2002) Distribution of CGG repeats and FRAXAC1/DXS548 alleles in South American populations. Am J Med Genet 111:243-52.
- Mingroni-Netto RC, Costa SS, Angeli CB and Vianna-Morgante AM (1999) DXS548/FRAXAC1 haplotypes in fragile X

428 Costa et al.

chromosomes in the Brazilian population. Am J Med Genet 84:204-207.

- Mingroni-Netto RC, Fernandes JG and Vianna-Morgante AM (1994) Relationship of expansion of CGG repeats and X-inactivation with expression of fra(X)(q27.3) in heterozygotes Am J Med Genet 5:443-446.
- Murray A, Ennis S and Morton N (2000) No evidence for parent of origin influencing premature ovarian failure in fragile X premutation carriers. Am J Hum Genet 67:253-254.
- Murray A, Macpherson JN, Pound MC, Sharrock A, Youings SA, Dennis NR, Mckechnie N, linehan P, Morton NE and Jacobs PA (1997) The role of size, sequence and haplotype in the stability of FRAXA and FRAXE alleles during transmission. Hum Mol Genet 6:173-84.
- Murray A, Webb J, Grimley S, Conway GS and Jacobs PA (1998) Studies of FRAXA and FRAXE in women with premature ovarian failure. J Med Genet 35:637-640.
- Murray A, Webb J, MacSwiney F, Shipley EL, Morton NE and Conway GS (1999) Serum concentrations of follicle stimulating hormone may predict premature ovarian failure in FRAXA premutation women. Hum Reprod 14:1217-1218.
- Nolin SL, Brown WT, Glicksman A, Houck Jr GE, Gargano AD, Sullivan A, Biancalana V, Brondum-Nielsen K, Hjalgrim H, Holinski-Feder E, Kooy F, Longshore J, Mandel JL, Matthijs G, Rosseau F, Steinbach P, Vaisanen ML, Von Koskull H and Sherman SL (2003) Expansion of the fragile X CGG repeat in females with premutation or intermediate alleles. Am J Hum Genet 72:454-464.
- Oberlé I, Rousseau F, Heitz D, Kretz C, Devys D, Hanauer A, Boué J, Bertheas MF and Mandel JL (1991) Instability of a 550-base pair DNA segment and abnormal methylation in fragile X syndrome. Science 252:1097-1102.
- Pieretti M, Zhang F, Fu YH, Warren ST, Oostra BA, Caskey CT and Nelson DL (1991) Absence of expression of the *FMR1* gene in fragile X syndrome. Cell 66:817-822.
- Richards RI, Holman K, Kozman H, Kremer E, Lynch M, Pritchard M, Yu S, Mulley J and Sutherland GR (1991) Fragile X syndrome: Genetic localisation by linkage mapping of two microsatellite repeats FRAXAC1 and FRAXAC2

- which immediately flank the fragile site. J Med Genet 28:818-823.
- Riggins GJ, Sherman SL, Oostra BA, Sutcliffe JS, Feitell D, Nelson DL, Van Oost BA, Smits APT, Ramos FJ, Pfendner E, Kuhl DPA, Caskey CT and Warren ST (1992) Characterization of a highly polymorphic dinucleotide repeat 150 kb proximal to the fragile X site. Am J Med Genet, 43:237-243.
- Schwartz CE, Dean J, Howard-Peebles PN, Bugge M, Mikkelsen M, Tommerup N, Hull C, Hagerman R, Holden JJA and Stevenson RE (1994) Obstetrical and gynecological complications in fragile X carriers: A multicenter study. Am J Med Genet 51:400-402.
- Sherman SL (2000) Premature ovarian failure among fragile X premutation carriers: Parent-of-origin effect? Am J Hum Genet 67:11-13.
- Sullivan AK, Markus M, Epstein MP, Allen EG, Anido AE, Paquin JJ, Yadav-Shah M and Sherman SL (2005) Association of *FMR1* repeat size with ovarian dysfunction. Hum Reprod 20:402-412.
- Tassone F, Hagerman RJ, Chamberlain WD and Hagerman PJ (2000) Transcription of the *FMR1* gene in individuals with fragile X syndrome. Am J Med Genet 97:195-203.
- Verkerk AJ, Pieretti M, Sutcliffe JS, Fu YH, Kuhl DP, Pizzuti A, Reiner O, Richards S, Victoria MF, Zhang F, Eussen BE, Van Ommen G-JB, Blonden LAJ, Riggins GJ, Chastain JL, Kunst CB, Galjaard H, Caskey CT, Nelson DL, Oostra BA and Warren ST (1991) Identification of a gene (*FMR-1*) containing a CGG repeat coincident with a breakpoint cluster region exhibiting length variation in fragile X syndrome. Cell 65:905-914.
- Vianna-Morgante AM and Costa SS (2000) Premature ovarian failure is associated with maternally and paternally inherited premutations in Brazilian families with fragile X. Am J Hum Genet 67:254-255.
- Vianna-Morgante AM, Costa SS, Pares AS and Verreschi ITN (1996) FRAXA premutation associated with premature ovarian failure. Am J Med Genet 64:373-375.
- Vianna-Morgante AM, Costa SS, Pavanello RCM, Otto PA and Mingroni-Netto RC (1999) Premature ovarian failure (POF) in Brazilian fragile X carriers. Genet Mol Biol, 22:471-474.

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