

In vitro and *in vivo* antioxidant activity of the pulp of Jatobá-do-cerrado

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Abstract

Oxygen metabolism in cells causes the production of free radicals, which produce damage, including changes in cell structure and function. Antioxidants are substances that, at low concentrations, slow down or prevent oxidation. Fruits and vegetables contribute to the dietary supply of these compounds. The flora of the Cerrado in Brazil has shown to have high levels of bioactive compounds. This study aimed to characterize the antioxidant activity of the pulp of jatobá-do-cerrado *in vitro* and *in vivo*. *In vitro* antioxidant activity of the aqueous, ethanol and aqueous acetone extracts was evaluated by the DPPH method. We determined total phenols by the Folin-Ciocalteu assay and tannins by the Folin-Denis method. *In vivo* antioxidant potential of the aqueous acetone extract was evaluated by the TBARS technique. The aqueous acetone extract had the highest antioxidant capacity, followed by the aqueous and ethanol extracts. The same pattern occurred in the extraction of phenols and in the extraction of tannins. *In vivo* administration of the aqueous acetone extract inhibited lipid peroxidation compared to the control group. The inhibition of peroxidation has increased by elevating the dosage concentration of the extracts, demonstrating a significant antioxidant potential *in vivo* as well as *in vitro*.

Keywords: bioactive compounds; rats; fruits of cerrado.

Practical Application: The bioactives found in jatobá demonstrated to be antioxidant *in vitro* and *in vivo*.

1 Introduction

The oxidative process is used for the production of energy required for essential cell activities; however, oxygen metabolism in the cells also causes the production of free radicals. Oxidizing agents are compounds produced by metabolism, and if not controlled, can cause damage (Alves et al., 2010).

Antioxidants can be defined as substances that at low concentrations retard or prevent the oxidation of a substrate (Sucupira et al., 2012). They are responsible for the inhibition and reduction of lesions caused by free radicals in cells (Abrahão et al., 2012). Among the foods that contain natural antioxidants, fruits and vegetables are the main contributors to the dietary supply of these compounds associated with beneficial health effects (Sucupira et al., 2012).

Jatobá-do-cerrado (jatobá, *Hymenaea stigonocarpa*) belongs to the family Fabaceae and is a fruit found naturally in regions of the Brazilian Cerrado and Cerradão (Empresa Brasileira de Pesquisa Agropecuária, 2010). This fruit has potential for cultivation and marketing its by-products, for example, the flour extracted from jatobá (Ávila et al., 2010).

Popularly, the tea made with jatobá bark is used for kidney and liver diseases, intestinal infections, and even for healing and as an expectorant, and its pulp is used as a laxative (Empresa Brasileira de Pesquisa Agropecuária, 2010). A study of the methanol extract of jatobá bark in rats has demonstrated that jatobá has the capacity for medicinal treatment of gastric and

duodenal ulcers and diarrhea as well, confirming popular findings (Orsi et al., 2012).

The same findings were reported in animal models of trinitrobenzenesulfonic acid-induced ulcer (Orsi et al., 2014), justifying the need to determine the antioxidant activity of this fruit. Moreover, jatobá is already commonly used in the treatment of certain diseases related to oxidative stress, and its potential in the prevention of such diseases has not yet been explored.

2 Materials and methods

2.1 Raw material

The fruits of jatobá were purchased commercially in the city of Campo Grande, MS, Brazil in February 2014, during the fruit harvest period.

In the Food Technology Unit and Public Health Laboratories of the Biological Sciences and Health Center of the Federal University of Mato Grosso do Sul (UTA-CCBS-UFMS), the fruits were washed with water and scrubbed, sanitized with bleach (150 ppm), air-dried, weighed, peeled and pulped in the dark to avoid the effect of light on the antioxidant compounds in the fruits.

The separated pulp of the fruit was homogenized and sieved (40 mesh Tamil), obtaining a mash that was packaged in plastic

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bags, stored in the dark in a biochemical oxygen demand (BOD) refrigerator at 2 °C until analysis.

Upon completion of the *in vitro* analysis, the extract with the highest antioxidant potential was selected for use in *in vivo* studies.

2.2 Evaluation of antioxidant activity

In vitro antioxidant activity was determined in aqueous, ethanol (with absolute ethanol) and aqueous acetone (water:acetone, 20:80) extracts. The extractions were performed with the fruit pulp. Approximately 2.5 g of pulp was ground in a beaker and 15 ml of solvent were added (distilled water, ethanol or 80% acetone). The mixture of sample and solvent was homogenized using a mechanical stirrer for 20 minutes and then filtered through gauze in a glass funnel. The residue retained was recovered and extracted with 15 mL of fresh solvent, and the mixture was homogenized in a blender for about 20 minutes and filtered through new gauze. The second filtrate was added to the first, and finally, extraction solvent was added up to 50 mL, providing the extracts for analysis, which were diluted 1:1 with extraction solvent.

Total *in vitro* antioxidant activity was determined by the DPPH method, and total phenols and tannins by the Folin Ciocalteu and Folin Denis assays, respectively.

A new aqueous acetone extract was prepared for use in animals, since this extract of jatobá pulp showed the highest antioxidant potential. Since acetone in the extract is toxic to animals, this was evaporated in a rotary evaporator and then dissolved in distilled water. Three doses of the extract were tested, 150, 250 and 500 µL, diluted with distilled water for administration of a total of 1 mL.

2.3 Experimental assay

Male Wistar rats weaned at 21 days were used in the experiment. The animals were allowed to adapt for a week, where the animals were placed in polyethylene cages and in groups, receiving only water and food. Afterwards, the experiment was started, using a 30-day treatment period.

The distribution of animals was performed randomly as follows: eight animals in the control group (C) and 8 animals in each group treated with jatobá extract at doses of 150 (T1), 250 (T2) and 500 µL/day (T3), totaling 32 animals, kept with four animals per cage. The animals were given water and commercial feed *ad libitum*, under a 12-hour light/ dark cycle and controlled temperature. The rats were given distilled water or extract by gavage once daily for 30 days. The control group received only 1 mL of distilled water, and the treated groups were

given the different doses of jatobá extract in 1 mL of distilled water. After the treatment periods, *in vivo* antioxidant activity was determined by the TBARS technique, using brain tissue.

The procedures involving laboratory animals were approved by the Ethics Committee for the Use of Animals (CEUA) of the Federal University of Mato Grosso do Sul (UFMS) (Protocol No. 526/2013).

2.4 Statistical analysis

In vitro data showed a normal distribution, and therefore, one-way ANOVA was applied; and with significant results, a normality test was used with Tukey's post-test ($p < 0.05$) for comparison of means. *In vivo* data showed a non-normal distribution, so the Kruskal-Wallis test was applied with Dunn's post-test.

3 Results and discussion

The levels of antioxidant activity, total phenols and tannins found in the pulp of jatobá are given in Table 1.

The aqueous acetone, aqueous and ethanol extracts differed significantly from each other ($p < 0.001$) in relation to antioxidant activity, and the aqueous acetone extract showed the greatest inhibition of DPPH radical oxidation, followed by the aqueous and ethanol extracts.

Dias et al. (2013) found an IC₅₀ of 5.84 ± 0.10 µL/mg in the ethyl acetate fraction of the fruit pulp, lower than that found in this study, for all extractions. Compared to other fruits of the Cerrado, the IC₅₀ values of the aqueous and aqueous acetone extracts here were lower than those found for araticum-do-mato, butiazeiro and mandacaru-de-três-quinhas in extracts of mixed pulp using methanol (50%) and aqueous acetone (70%) (Pereira et al., 2013). The findings of this study also showed lower IC₅₀ values when compared to those for ethanol and aqueous extracts of fruit peel and pulp of cagaita (*E. klotzchiana*), (387.47 ± 8.70 and 879.33 ± 11.70 g/g DPPH, respectively) and araticum (49.18 ± 3.13 and 198.28 ± 8.24 g/g DPPH, respectively), using fresh samples.

The ethanol extract was shown to be less statistically efficient in the extraction of phenols compared to the other solvents ($p < 0.01$). The aqueous acetone and aqueous extracts did not differ significantly from each other.

Rocha et al. (2013) found total phenol values of 105-385 mg TAE/100 g in aqueous acetone (70%) extract and of 0.0-252 mg TAE/100 g in ethanol (95%) extract for guapeva (*Pouteria gardneriana*), mama-cadela (*Campomanesia sp.*), cagaita (*E. klotzchiana*), cambuçá (*Pliniaedulis*), gabirola (*Campomanesia sp.*), jaracatiá

Table 1. Antioxidant activity, total phenols and tannins in aqueous acetone, ethanol and aqueous extracts of pulp of jatobá-do-cerrado.

	Aqueous acetone	Ethanol	Aqueous	p value
Antioxidant activity (IC ₅₀ mg/mg DPPH)*	< 26.76 ^a	618.06 ^c	117.07 ^b	<0.001
Total phenols (mg GAE/100 g)	786.18 ^a	444.97 ^b	669.64 ^a	<0.01
Total tannins (mg TAE/100 g)	1838.69 ^a	518.43 ^c	728.14 ^b	<0.01

Different letters in the same row indicate significant differences ($p < 0.01$) according to Tukey's test. *Lower IC₅₀ indicates greater antioxidant potential, ranked by order of letters.

(*Jaracatia spinosa*), pera-do-cerrado (*Eugenia klotzschiana*) and pitanga-do-cerrado (*Eugenia punicifolia*). These values demonstrated that jatobá had a high concentration of phenols, associated with its antioxidant activity potential.

Peres et al. (2013) reported a total phenol level in the ethyl acetate extract of jatobá fruit pulp of $18 \pm 1.54 \mu\text{g GAE/mg}$ of extract, finding a higher concentration of pulp phenolics in jatobá compared to other fruits of the Cerrado, such as jabuticaba (*Myrciaria cauliflora*) ($7.92 \pm 0.56 \mu\text{g GAE/mg}$), but showing better extraction of phenolics from the fruit peel.

Rocha et al. (2013) found higher values of GAE in ethanol extracts compared to aqueous (34.1 and 25.19 mg GAE/100 g, respectively). The values reported by these authors are below those in the present study and also those determined by Dias et al. (2013) in the oil of jatobá pulp ($343 \pm 8.0 \text{ mg GAE/100 g}$).

The difference in the values found may be due to the technique used, fruit characteristics (cultivation, climate, maturation stage at collection, and collection and storage characteristics) and also because of the extractions performed, since Dias et al. (2013) extracted the oil from the pulp, and in their study, aqueous and aqueous acetone extracts showed better antioxidant potential, demonstrating the polar character of the bioactive compounds in this fruit.

According to Rufino et al. (2010), phenol content can be classified as low for $<100 \text{ mg GAE/100 g}$, intermediate for $100\text{-}500 \text{ mg GAE/100 g}$ and high for $>500 \text{ mg GAE/100 g}$ of fresh weight, whereas in dry matter, phenol content is classified as low for $<1000 \text{ mg GAE/100 gm}$ intermediate for $1000\text{-}5000 \text{ mg GAE/100 g}$ and high for $>5000 \text{ mg GAE/100 g}$, indicating that the aqueous and aqueous acetone extracts of jatobá evaluated in this study had high total phenol values.

Tannin levels were expressed in milligrams of tannic acid equivalents (TAE) in 100 g of sample. The aqueous, aqueous acetone and ethanol extracts were statistically different ($p < 0.01$), where the aqueous acetone extract had the highest level, followed by the aqueous and ethanol extracts.

Compared to other native fruits of the Cerrado, the extracts described in this study showed a very high tannin content when compared to that obtained for the 80% methanol in water extract of jambolão, reported by Faria et al. (2011), with a mean of $3.9 \pm 0.8 \text{ mg TAE /100 g}$

Arapitsas (2012) considered a mixture of water and acetone as the best solvent for the extraction of tannins, since extraction with ethanol can lead to the formation of ethyl gallate corroborating the results of this study.

In a study by Chavan & Singhal (2013), 80% aqueous acetone also showed greater extraction efficacy compared to pure acetone or solvent with lower acetone percentage with betel nut (*Areca catechu*), regarding antioxidant activity and amounts of total phenolics and tannins, corroborating the results found in this study. These results may be related to difference in polarity of solvents, which determines the dissolution of a group of antioxidant compounds, thereby influencing antioxidant activity.

In this work, at the beginning of the *in vivo* experiment, the animals were weighed and there was no significant difference in initial weight between groups ($p = 0.899$). The day before sacrificing, the animals were weighed again, with no significant difference in final weight between groups ($p = 0.4138$). Thus, there was no significant difference in weight gain between groups ($p = 0.5853$), as shown in Table 2.

Tinkov et al. (2014) found a significant difference ($p < 0.05$) in weight of animals fed diets with a high lipid content and animals with the same diet and treated with an aqueous extract of *P. maximum*. The authors attributed the antiadipogenic, antidiabetic, antiinflammatory effects and capacity to normalize the intestinal flora of the animals to bioactive components present in the extract.

In this study, with respect to the aqueous acetone extract formulated for *in vivo* application was tested for its antioxidant capacity, where the IC50 was $<26.44 \text{ mg/mgDPPH}$.

After sacrificing the animals, the brain was removed of animals, and inhibition of oxidation of brain tissue was determined in treated animals relative to the control group. Inhibition of oxidation was greater with increasing concentration of extract (Graph 1).

The Control and Treatment 1 groups did not differ statistically ($p > 0.05$), although the latter showed higher inhibition of oxidation values. Inhibition of oxidation in brain tissue was greater according to the following order: Treatment 3 (T3) > Treatment 2 (T2) > Treatment 1 (T1) > Control (C), as shown in Table 3.

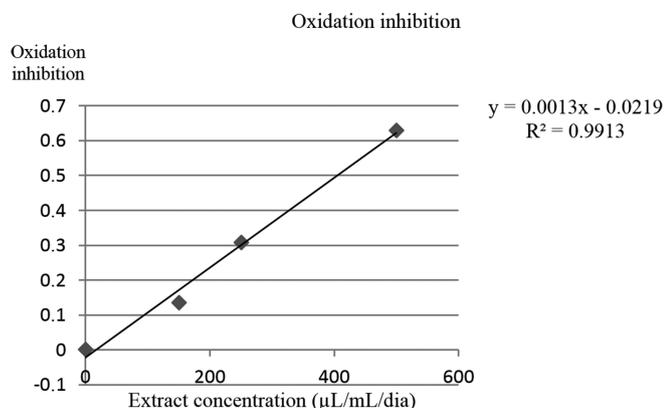
The T2 and T3 groups showed significant difference in inhibition of oxidation compared to the control group ($p < 0.001$). T2 was inhibited lipid peroxidation significantly more than did T1 ($p < 0.05$), and T3 was more effective compared to T2 ($p < 0.05$) and T1 ($p < 0.001$).

The aqueous acetone extract of jatobá showed high antioxidant potential *in vivo*, and antioxidant activity increased with dose, and was thus dose dependent. The doses tested in the animals were equivalent to 3.75, 6.25 and 12.5 mg/day for 150, 250 and

Table 2. Initial weight, final weight and weight gain of the control and treatment groups (n=8), using aqueous acetone (80:20) extract of pulp of jatobá-do-cerrado.

Group/dose	Initial**	Final**	Weight gain**	p value*
Control	50.62 + 2.92	281.50 + 24.04	230.88 + 22.65	0.5853
Treatment 1 - 150 μL /day	49.50 + 3.38	297.62 + 25.38	230.18 + 26.50	
Treatment 2 - 250 μL /day	49.87 + 4.08	269.12 + 27.41	219.25 + 28.23	
Treatment 3 - 500 μL /day	49.50 + 3.11	267.25 + 17.65	217.25 + 18.67	

*Values considered statistically significant when $p < 0.05$. **Weight (g) given as mean \pm standard deviation.



Graph 1. Inhibition of oxidation in brain tissue of rats, according to dose of aqueous acetone extract of Jatobá-do-cerrado ($\mu\text{L}/\text{day}$) administered.

Table 3. Percent inhibition of oxidation in rat brain tissue according to dose of the aqueous acetone extract of jatobá-do-cerrado ($\mu\text{L}/\text{day}$) administered.

Treatment	Inhibition of oxidation - %
Control	- ^c
T1 - 150 $\mu\text{L}/\text{day}$	13.46 ^c
T2 - 250 $\mu\text{L}/\text{day}$	30.77 ^b
T3 - 500 $\mu\text{L}/\text{day}$	62.92 ^a

Different letters indicate significant differences ($p < 0.05$).

500 $\mu\text{L}/\text{day}$, respectively, amounting to 45-155 mL of extract/day in humans, maintaining a concentration of 2.5%, as described in the *in vitro* assay.

In a study on coffee (*Coffea arabica*), Duarte et al. (2009) found 48.60% inhibition of lipid peroxidation in Wistar rats with acute treatment for 7 days and 53.40% with chronic treatment (30 days). Regarding the time of administration, the aqueous acetone extract of jatobá was more effective at inhibiting lipid peroxidation at a concentration of 500 $\mu\text{L}/\text{day}$, causing 62.92% inhibition. The doses of 150 and 250 $\mu\text{L}/\text{day}$ caused 13.46 and 30.77% inhibition, respectively.

In the aqueous extract of the bark of *S. henningsii* Gilg., an African shrub, Oyedemi et al. (2010) found dose-dependent inhibition of lipid peroxidation with eight days of treatment. This extract was able to inhibit lipid peroxidation by up to 47-58%, which was somewhat lower than with the highest dose of the aqueous acetone extract of jatobá.

However, the concentrations used in preparing the coffee and *S. henningsii* extracts were respectively 10 and 6%, while the concentration of the extracts used here was 2.5%. Thus, when compared with these extracts, we see that even a low concentration of the aqueous acetone extract of jatoba had a high antioxidant potential *in vivo* (Duarte et al., 2009); (Oyedemi et al., 2010).

Although the aqueous acetone extract was shown to have higher antioxidant potential *in vitro*, an *in vitro* study does not take into account particular *in vivo* processes such as digestion, absorption and metabolism, the combination of factors to which living organisms may be exposed, and other metabolic factors

that can increase or decrease oxidative stress (the presence of diseases, stress, and combination of endogenous and dietary exogenous antioxidants) (Prior & Wu, 2013).

For the antioxidant potential of jatobá extracts cannot be assigned to a single bioactive component, since both *in vitro* and *in vivo*, antioxidant potential increases in the presence of more than one bioactive compound, because they can act synergistically or antagonistically with each other (Hidalgo et al., 2010). But it is possible to correlate the antioxidant activity described with extraction of total phenols and tannins, highlighting the importance of these bioactive compounds in the antioxidant capacity of these extracts studied. These results corroborate those obtained by Lopes et al. (2012), who found a positive correlation between the amount of phenolics and antioxidant activity.

4 Conclusions

All extracts exhibited antioxidant activity, and the aqueous acetone extract had the lowest IC₅₀, followed by the ethanol and aqueous extracts. The extraction of phenols and tannins followed the same pattern, where the aqueous acetone extract showed higher levels of phenols and tannins than did the aqueous and ethanol extracts.

The aqueous acetone extract inhibited lipid peroxidation in rat brain tissue at all concentrations used and in a concentration-dependent manner.

The aqueous acetone extract showed high antioxidant potential both *in vitro* and *in vivo*, probably due to the diversity and high concentration of bioactive compounds present, which can act synergistically.

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