Quercetin postconditioning attenuates myocardial ischemia/reperfusion injury in rats through the PI3K/Akt pathway

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Abstract

Quercetin (Que), a plant-derived flavonoid, has multiple benefical actions on the cardiovascular system. The current study investigated whether Que postconditioning has any protective effects on myocardial ischemia/reperfusion (I/R) injury *in vivo* and its potential cardioprotective mechanisms. Male Sprague-Dawley rats were randomly allocated to 5 groups (20 animals/group): sham, I/R, Que postconditioning, Que+LY294002 [a phosphatidylinositol 3-kinase (PI3K)/Akt signaling pathway inhibitor], and LY294002+I/R. I/R was produced by 30-min coronary occlusion followed by 2-h reperfusion. At the end of reperfusion, myocardial infarct size and biochemical changes were compared. Apoptosis was evaluated by both TUNEL staining and measurement of activated caspase-3 immunoreactivity. The phosphorylation of Akt and protein expression of Bcl-2 and Bax were determined by Western blotting. Que postconditioning significantly reduced infarct size and serum levels of creatine kinase and lactate dehydrogenase compared with the I/R group (all P<0.05). Apoptotic cardiomyocytes and caspase-3 immunoreactivity were also suppressed in the Que postconditioning group compared with the I/R group (both P<0.05). Akt phosphorylation and Bcl-2 expression increased after Que postconditioning, but Bax expression decreased. These effects were inhibited by LY294002. The data indicate that Que postconditioning can induce cardioprotection by activating the PI3K/Akt signaling pathway and modulating the expression of Bcl-2 and Bax proteins.

Key words: Ischemia and reperfusion; Quercetin; Postconditioning; PI3K/Akt

Introduction

Acute myocardial infarction is a major cause of death and disability worldwide. Although early reperfusion is essential for myocardial salvage, it induces reperfusion injury, which reduces the benefits of myocardial reperfusion (1,2). Ischemic postconditioning, defined as brief periods of ischemia and reperfusion (I/R) at the onset of reperfusion, has been shown to protect against lethal reperfusion injury in many species including humans (3). However, this strategy is not suitable for patients treated with thrombolytic agents. Thus, a more feasible pharmacological postcondtioning is needed. Many chemicals such as erythropoietin (4), adenosine (5), and hydrogen sulfide (6) are known to be cardioprotective when given at the onset of reperfusion, but none has been widely used.

Quercetin (Que; 3,3',4',5,7-pentahydroxyflavone) is an important member of the flavonoid family that is present in plants and red wine (7). It is nontoxic and has a broad range of pharmacological and biological activities including antioxidative, anticarcinogenic, vasoprotective, anti-inflammatory, antidiabetic, and antiplatelet effects (8-11). Several studies indicated that Que, when given before ischemia (preconditioning), protects the myocardium from I/R injury through its antioxidant and anti-inflammatory activities (12,13). Recently, Bartekova et al. (14) reported a protective role for Que, when administered during reperfusion (postconditioning), in an isolated rat heart model. However, it is unclear whether Que postconditioning has a protective effect against myocardial I/R injury *in vivo*. Moreover, the molecular mechanism underlying Que-mediated cardioprotection is unknown.

Phosphoinositide 3-kinases (PI3Ks) and their downstream effector Akt are key signal transduction enzymes involved in modulating cell proliferation, survival, and apoptosis (15). Several studies have indicated that the PI3K/Akt signaling pathway plays a key role in cardiac protection against I/R injury (16,17). It has also been shown that Que attenuates postischemic neuronal apoptosis by activating the PI3K/Akt pathway (18). However, it

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is unclear whether the PI3K/Akt pathway mediates the cardioprotection of Que postconditioning.

In the present study, Que postconditioning was used as an adjuvant to attenuate myocardial I/R injury in an *in vivo* rat model.

Material and Methods

Animals

Male Sprague-Dawley rats weighing 250 ± 20 g were purchased from Wuhan University Animal Center (Wuhan, China). This study conformed to the Guidelines for the Care and Use of Laboratory Animals by the National Institutes of Health (NIH Publication No. 80-23), and the experimental procedures were approved by the Institutional Animal Ethics Committee of Wuhan University.

In vivo myocardial I/R model

The *in vivo* myocardial I/R model was modified from a previous study (17). Briefly, rats were anesthetized by an intraperitoneal (*ip*) injection of 40 mg/kg 1% pentobarbital sodium. The rats were intubated and mechanically ventilated with room air with a rodent respirator (DV-2000, Shanghai Jia Peng Technology Co., Ltd., China). A left thoracotomy was carried out to expose the hearts. A 5-0 silk ligature was passed under the left anterior descending coronary artery, and a small vinyl tube was placed on top of the vessel to form a snare for reversible coronary occlussion. After 30 min of ischemia, the heart was reperfused for 2 h by releasing the snare.

Experimental groups

A total of 100 rats were randomly allocated in equal numbers (n = 20) to 5 groups. 1) sham: rats were subjected to the surgical procedures without coronary occlusion; 2) I/R: 30-min coronary occlusion and followed by 2-h reperfusion; 3) Que postconditioning (Que): 10 mg/kg Que ip 5 min before reperfusion; 4) Que + LY: 10 mg/kg Que ip 5 min before reperfusion and 0.3 mg/kg LY294002 ip 10 min before reperfusion; 5) LY+I/R: 0.3 mg/kg LY294002 ip 10 min before reperfusion. Que and LY294002 (both Sigma-Aldrich, USA) were dissolved in 0.02% dimethyl sulfoxide. The doses of Que and LY294002 were administered as described previously (19,20).

Determination of myocardial infarct size

After reperfusion, the ligature around the left anterior descending coronary artery was retied. Two milliliters of 2% Evans blue (Sigma-Aldrich) was given intravenously to track the perfused region and the area at risk (AAR). Tissue was collected from the right ventricle and atrium. The left ventricle (LV) was removed, cut into 3-mm transverse slices and incubated in 1% tetrazolium chloride (Sigma-Aldrich) solution for 20 min to identify the infarct

area. The weight of the infarct area (white), AAR (red), and nonischemic zones of the LV (blue) was measured. Infarct size is reported as a percentage of the AAR mass.

Measurement of serum creatine kinase (CK) and lactate dehydrogenase (LDH)

At the end of reperfusion, blood samples were collected and centrifuged at 800 g for 10 min. Serum CK and LDH levels were measured by a colorimetric method using commercial kits (Nanjing Jiancheng Bioengineering Institute, China) according to the manufacturer's protocols. The results are reported as U/L.

Apoptotic assessment

Terminal dUTP nick end-labeling (TUNEL) and caspase-3 activity detection were used to assess apoptosis (21). To examine cardiac myocyte apoptosis, samples of tissue from ischemic zones were fixed in 4% paraformaldehyde, embedded in paraffin, and cut into 5- μ m transverse sections. The TUNEL assay was carried out using an apoptosis detection kit (Roche Applied Science, Germany) according to the manufacturer's protocols. By light microscopy, TUNEL-positive myocardial cells had brown-staining nuclei. Five visual fields were selected randomly from each block, and at least 100 cells per field were counted at 200 \times . The apoptotic index was calculated (apoptotic cells/total cells \times 100%) from a total of 25 fields per sample.

To examine caspase-3 activity in ischemic zones, immunohistochemistry was conducted using antibody against cleaved caspase-3 (Cell Signaling Technology, USA) as described previously. Tissue sections were prepared as described earlier. Slides were viewed under a light microscope at $400 \times$ magnification (Olympus BX50 Microphotographic System, Japan). Three tissue sections were selected randomly for each animal. Immunohistochemical staining intensity was measured as mean optical density using HIPAS-2000 image-analysis software (Qianli Technical Imaging, China).

Western blot analysis

At the end of reperfusion, protein extracts were prepared as previously described (22). Briefly, cardiac samples were homogenized in lysis buffer containing 50 mM Tris-HCl, pH 7.4, 150 mM NaCl, 5 mM EDTA, 1 mM dithiothreitol, 1% Triton X-100, and 1% protease inhibitor cocktail. The homogenates were centrifuged at 12,000 *g* for 15 min, and the supernatants were collected for Western blotting. Protein concentration was determined by a bicinchoninic acid protein assay (Beyotime Biotechnology, Inc., China), and 50-µg protein homogenate samples were separated on 15% SDS-polyacrylamide gels, transferred to nitrocellulose membranes (BioRad, USA), and blocked with 5% nonfat dry milk prepared in Tris-buffered saline containing 0.05% Tween 20 (TBST). The membranes were then incubated overnight at 4°C with

antibodies against phospho-Akt (Ser473), Akt, Bcl-2, or Bax (Cell Signaling Technology). After 4 washes in TBST, the membranes were incubated with a secondary antibody (Santa Cruz Biotechnology, Inc., USA) for 2 h at room temperature. Signals were detected with an enhanced chemiluminescence kit (Beyotime Biotechnology).

Statistical analysis

Data are reported as means \pm SD. Differences between groups were analyzed by one-way ANOVA followed by the Bonferroni *post hoc* test. P<0.05 was considered to be statistically significant. Data analyses were performed with the SPSS 13.0 software (SPSS, Inc., USA).

Results

Effects of Que postconditioning on I/R-induced infarct size

The ratio of AAR to LV mass was not significantly different among the groups, suggesting a comparable degree of existing ischemic impairment (Figure 1A). The infarct size following I/R was $50\pm4\%$ in the I/R group. Que postconditioning significantly reduced infarct size compared with the I/R group $(27\pm4~vs~50\pm4\%,~P<0.05)$. LY294002 administration eliminated the Que-mediated reduction in infarct size $(49\pm4~vs~27\pm4\%,~P<0.05)$. Administration of LY294002 alone before reperfusion had no effect on I/R-induced infarct size $(47\pm3~vs~50\pm5\%,~P>0.05;~Figure~1B)$.

Serum CK and LDH were determined as another indicator of myocardial insult induced by I/R. Serum levels of both CK (2679.5 \pm 194.3 $\it vs$ 1141.5 \pm 118.7 U/L, P<0.05) and LDH (2618 \pm 197.7 $\it vs$ 901 \pm 98.7 U/L, P<0.05) were significantly elevated in the I/R group compared with those in the sham group. Que postconditioning reduced the levels of CK (1642.9 \pm 194.3 $\it vs$ 2679.5 \pm 194.3 U/L, P<0.05) and LDH (1273.6 \pm 176.5 $\it vs$ 2618 \pm 197.7 U/L, P<0.05) compared to the I/R group. LY294002 administration eliminated the effects of Que postconditioning on CK (2675.3 \pm 202.5 $\it vs$

1642.9 \pm 194.3 U/L, P<0.05) and LDH (2523.6 \pm 198.8 vs 1273.6 \pm 176.5 U/L, P<0.05). Consistently, administration of LY294002 alone before reperfusion had no effect on I/R-induced CK and LDH release (Figure 1C and D).

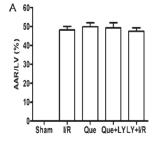
Effects of Que postconditioning on I/R-induced apoptosis

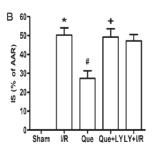
The percentage of TUNEL-positive myocardial cells following I/R was $40.2\pm3.5\%$ in the I/R group. Que postconditioning significantly decreased the percentage of TUNEL-positive myocardial cells compared to the I/R group $(20.6\pm3.4~vs~40.2\pm3.5\%,~P<0.05)$. LY294002 administration eliminated Que-mediated decrease in the percentage of TUNEL-positive cells $(39.2\pm2.9~vs~20.6\pm3.4\%,~P<0.05)$. Administration of LY294002 alone before reperfusion had no effect on I/R-induced cell apoptosis (Figure 2).

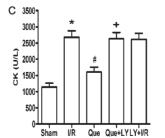
To further determine the occurrence of apoptotic cell death, we measured caspase-3 activity by immunohistochemistry. As shown in Figure 3, caspase-3 immunoreactivity was significantly increased in the I/R group compared to that in the sham group ($70\pm3.8~vs\ 12\pm2.5$, P<0.05). Que postconditioning reduced caspase-3 immunoreactivity compared to the I/R group ($23.8\pm3.0~vs\ 70\pm3.8$, P<0.05). LY294002 administration eliminated Que-mediated reduction in caspase-3 immunoreactivity ($66.3\pm4.8~vs\ 23.8\pm3.0,~P<0.05$). Administration of LY294002 alone before reperfusion had no effect on I/R-induced caspase-3 activation.

Effects of Que postconditioning on Bcl-2 and Bax expression in I/R-induced hearts

As shown in Figure 4, a significant decrease in Bcl-2 expression and an increase in Bax expression occurred following I/R injury (both P<0.05). Que postconditioning significantly increased Bcl-2 and decreased Bax expression compared with that observed in the I/R group (both P<0.05). However, LY294002 abolished the effects of Que postconditioning on Bcl-2 and Bax expression (both P<0.05).







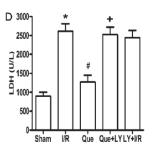


Figure 1. Effect of Que postconditioning on I/R-induced myocardial injury. *A*, AAR is reported as mean \pm SD percentage of the LV mass (n = 5). *B*, Infarct size is reported as mean \pm SD percentage of the AAR mass (n = 5). *C*, Effect of Que postconditioning on serum levels of CK (n = 8). *D*, Effect of Que postconditioning on serum levels of LDH (n = 8). Que: quercetin; I/R: ischemia/reperfusion; LY: LY294002; IS: infarct size; AAR: area at risk; LV: left ventricle; CK: creatine kinase; LDH: lactate dehydrogenase. *P<0.05 vs sham group, $^{\#}$ P<0.05 vs I/R group, $^{+}$ P<0.05 vs Que group (one-way ANOVA followed by the Bonferroni test).

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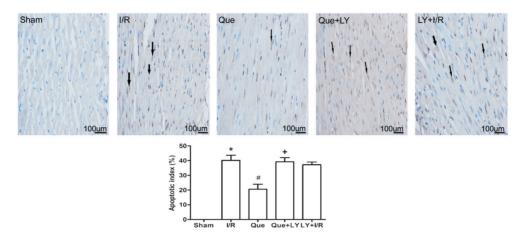


Figure 2. Effect of Que postconditioning on I/R-induced cardiomyocyte apoptosis determined by TUNEL (n=5). Arrows indicate TUNEL-positive cells. Data are reported as means \pm SD. Que: quercetin; I/R: ischemia/reperfusion; LY: LY294002; TUNEL: terminal dUTP nick end-labeling. *P<0.05 vs sham group, $^{\#}$ P<0.05 vs I/R group, $^{+}$ P<0.05 vs Que group (one-way ANOVA followed by the Bonferroni test).

Effects of Que postconditioning on Akt phosphorylation in I/R-induced hearts

As shown in Figure 5, the ratio of P-Akt/Akt expression was significantly increased in I/R and Que postconditioning groups compared to the sham group (P<0.05). LY294002 administration inhibited the phosphorylation of Akt in Que-treated hearts (P<0.05).

Discussion

There are two major findings in the current study. First, we confirmed that Que postconditioning significantly reduced infarct size and decreased cardiomyocyte apoptosis following myocardial I/R injury in rats *in vivo*. Second, we confirmed that the cardioprotective effects of

Que postconditioning involved activation of the PI3K/Akt pathway by using a PI3K/Akt signaling pathway inhibitor (LY294002), which could abolish the cardioprotective effects of Que postconditioning.

The onset of myocardial ischemia is usually unpredictable clinically. Therefore, agents that confer cardioprotection at the onset of reperfusion after myocardial ischemia provide a promising approach to attenuate myocardial I/R injury. Que is a member of the flavonoid family and is widely present in the plant kingdom (7). Que manifests a wide range of beneficial biological activites and can mediate cardioprotective effects (7,23). For example, an increased intake of Que has been suggested to reduce risk of cardiovascular diseases (23). Previous studies have also indicated that Que could reduce infarct

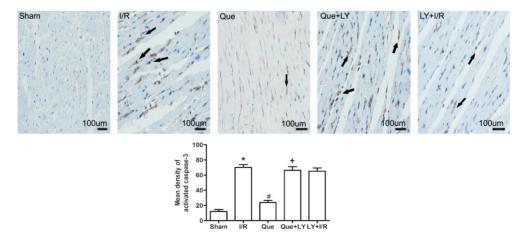


Figure 3. Immunohistochemical staining for activated caspase-3 in the myocardium after I/R injury. Arrows indicate the positive expression of activated caspase-3. Data are reported as means \pm SD. Que: quercetin; I/R: ischemia/reperfusion; LY: LY294002. *P<0.05 vs sham group, #P<0.05 vs I/R group, *P<0.05 vs Que group (one-way ANOVA followed by the Bonferroni test).

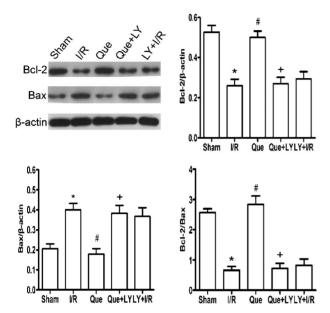


Figure 4. Effect of Que postconditioning on Bcl-2 and Bax expression after myocardial I/R injury (n=5). Data are reported as means \pm SD. Que: quercetin; I/R: ischemia/reperfusion; LY: LY294002. *P<0.05 vs sham group, #P<0.05 vs I/R group, *P<0.05 vs Que group (one-way ANOVA followed by the Bonferroni test).

size and improve functional recovery in an acute myocardial I/R model when given prior to ischemia (24,25). Similar results were observed in a myocardial infarction model induced by infusion of isoproterenol (26,27). Recently, Bartekova et al. (14) described the protective effects of Que postconditioning in an isolated rat heart model of I/R injury. Our data are consistent with that study (14), and we further demonstrated that Que postconditioning exerted remarkable cardioprotective effects against myocardial I/R injury in an *in vivo* rat

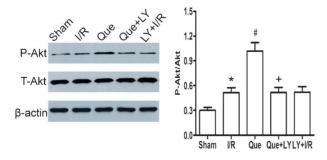


Figure 5. Effect of Que postconditioning on P-Akt expression during myocardial I/R (n=5). Data are reported as means \pm SD. Que: quercetin; I/R: ischemia/reperfusion; LY: LY294002; P-Akt: phospho-Akt; T-Akt: total-Akt. *P<0.05 vs sham group, $^{\#}P$ <0.05 vs I/R group, ^{+}P <0.05 vs Que group (one-way ANOVA followed by the Bonferroni test).

model showing diminished infarct size and reduced CK and LDH release.

It is well documented that apoptosis is dominant in the pathogenesis of myocardial I/R injury. Suppression of apoptosis could reduce loss of cardiomyocytes and limit myocardial damage caused by I/R (28). Ischemic post-conditioning, which consists of brief periods of I/R at the onset of reperfusion, has been shown to inhibit cardiomyocyte apoptosis and attenuate myocardial I/R injury (29). Similar to ischemic postconditioning, the results from TUNEL staining and immunohistochemisty for activated caspase-3 in our current study showed that Que postconditioning could significantly decrease cardiomyocyte apoptosis after I/R injury. These results suggest that antiapoptosis might be one of the major mechanisms underlying the cardioprotective effects of Que postconditioning.

Bcl-2 family members regulate apoptosis by modulating mitochondrial membrane permeability (30). The antiapoptotic protein Bcl-2 is located in the mitochondrial wall and prevents mitochondrial release of cytochrome c. while the proapoptotic protein Bax resides in the cytosol but can be translocated to mitochondria to induce cytochrome c release. Therefore, the ratio of the Bcl-2/ Bax protein is a key factor in determining cell apoptosis or survival after apoptotic stimuli (31). Experimental studies have demonstrated that ischemic postconditioning reduces apoptosis by modulating Bcl-2 and/or Bax expression that prevents cardiomyocyte apoptosis following myocardial I/R (29). In the current study, we found that Que postconditioning significantly decreased Bax expression and increased Bcl-2 expression and the ratio of Bcl-2/Bax. These results indicate that antiapoptosis of Que postconditioning might be related to changes in the expression of Bcl-2 and Bax.

It is not clear how Que modulates Bcl-2 and Bax expression during myocardial I/R in vivo. The antioxidant action of Que may contribute to this modulation (32), since many antioxidants affect the expression of Bcl-2 family proteins during myocardial I/R (33,34). Alternatively, activation of PI3K/Akt may be responsible for the changes in Bcl-2 and Bax expression (35). PI3K/ Akt is an important component of the intracellular signaling pathway regulating cell growth, survival and apoptosis. Activating PI3K/Akt at the time of reperfusion has been demonstrated to provide protection against myocardial I/R injury (4,18,36,37). Tsang et al. (36) showed that ischemic postconditioning increased phosphorylation of Akt and inhibition of Akt activity partly abolished the cardioprotective effects of ischemic postconditioning. Many cardioprotectants, including erythropoietin (4), sulfentanil (17), and KB-R7943 (37), confer their cardioprotective effects through the PI3K/Akt pathway. In this study, Que postconditioning significantly increased phosphorylation of Akt and reduced cardiomyocyte apoptosis by regulating the Bcl-2 family. The cardioprotective effects of Que postconditioning were 866 Y. Wang et al.

blocked by the PI3K/Akt signaling pathway inhibitor LY294002. These data indicate that Que postcondtioning induces cardioprotection through the PI3K/Akt signaling pathway. It has been shown that previously activated Akt could inhibit expression of proapoptotic proteins such as Bad, Bax, and caspase-9 (15), but maintain high levels of antiapoptotic protein Bcl-2 (38). Akt has also been suggested to increase the production of nitric oxide (39). The protective roles of nitric oxide during ischemic postconditioning and pharmacological postconditioning have already been confirmed (37,38). Additionally, signaling through the PI3K/Akt pathway could activate PKC, which plays a key role in cardiac protection against myocardial I/R injury (40). Therefore, it appears that the PI3K/Akt pathway contributes to the recruitment of

various endogenous cardioprotective signals to attenuate myocardial I/R injury. Furture research is needed to identify other signaling elements involved in Que-induced cardioprotection against myocardial I/R injury.

In the present study, we demonstrated that Que postconditioning attenuates myocardial I/R injury in rats via activating the PI3K/Akt pathway and modulating Bcl-2 and Bax expression.

Acknowledgments

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