6 – ORIGINAL ARTICLE ANESTHESIA

Effects of epidural nalbuphine on intraoperative isoflurane and postoperative analgesic requirements in dogs¹

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ABSTRACT

PURPOSE: To measure the change in the minimum alveolar concentration of isoflurane (EtISO) associated with epidural nalbuphine and the postoperative analgesic requirements in dogs after ovariohysterectomy.

METHODS: Twenty four healthy female dogs were randomly assigned to receive saline or nalbuphine at 0.3 or 0.6 mg/kg (n=8 for each group) administered via lumbosacral epidural catheter introduced cranially into the epidural canal. Changes in heart and respiratory rates and arterial blood pressure during surgery were recorded along with the corresponding EtISO. Immediately after tracheal extubation, analgesia, sedation, heart rate, respiratory rate, and arterial blood pressure were measured at predetermined intervals and every 60 min thereafter until the first rescue analgesic.

RESULTS: A significant decrease in EtISO was associated with epidural nalbuphine at 0.3 mg/kg (26.3%) and 0.6 mg/kg (38.4%) but not with saline in ovariohysterectomized dogs. In the postoperative period, VAS and Colorado analgesic scores were lower for the dogs that received the higher nalbuphine dose, which only required supplemental analgesia 10 h following its administration, compared with dogs that received the lower dose.

CONCLUSION: Epidural nalbuphine significantly reduces the intra-operative isoflurane requirement and provides prolonged postoperative analgesia after ovariohysterectomy in dogs.

Key words: Anesthesia. Epidural. Nalbuphine. Isoflurane. Dogs.

Introduction

Epidural analgesia combined with inhalatory anesthesia may provide advantages over the conventional routes of analgesic administration (intravenous or intramuscular), including reduction of the doses of the inhalant agents required to produce anaesthesia and thus the side effects derived from higher doses¹. Epidural methadone in dogs undergoing pelvic limb surgical procedures resulted in greater decreases in the EtISO required to maintain anesthesia compared with the same doses administered intravenously². Other opioids are also used with the same goal, such as fentanyl^{3,4}, sufentanil^{5,6}, oxymorphone⁷, and especially morphine^{1,8-11}.

Epidural use of morphine has been limited though because of the dose-dependent risk of delayed respiratory depression and/or potential urinary retention requires close monitoring of respiratory patterns, which is not always possible in dogs^{11,12} and humans¹³.

Nalbuphine is a low lipophilic (octanol-buffer partition coefficient–9.75) semisynthetic opioid related to both oxymorphone and naloxone; it has relatively potent μ -antagonist and κ -agonist activity. The respiratory depression induced by opioids is primarily mediated by the μ -receptor agonist activity. The μ -antagonist properties of the nalbuphine should produce fewer μ -mediated side effects such as respiratory depression, pruritus, nausea and vomiting 13,14 .

The epidural administration of nalbuphine combined with an inhalant anesthetic agent to produce the inhalant-sparing effect has not been investigated in dogs, nor has the onset and duration of action of postoperative pain relief been assessed. Therefore, we designed this study to determine if epidurally-administered nalbuphine may reduce isoflurane and postoperative analgesic requirements in dogs undergoing ovariohysterectomy that had received epidural nalbuphine.

Methods

This study was approved by the Animal Care Committee of the Federal University of Mato Grosso do Sul (UFMS) and was conducted under Good Clinical Practice guidelines. Twenty-four female dogs (ASA status 1) from the Zoonosis Control Centre of the municipality of Campo Grande to be spayed were used in this clinical trial. These animals were recruited over one year (2011–2012) and randomly assigned to one of three treatment groups. A variety of breeds were represented, with mixed-breed dogs predominating. Exclusion criteria included a body weight <15 kg or >30 kg; presence of uterine diseases, such as pyometra, dead foetuses, or canine transmissible venereal tumour (CTVT), or evidence of

pregnancy; a contraindication to epidural administration (previous pelvic fractures, obesity, dermatitis at the insertion of the epidural needle or coagulopathies); and the presence of pulmonary disease as shown on thoracic radiographs. Additionally, any dog having cardiac arrhythmias, clinical signs of systemic disease, or abnormal laboratory data was excluded from the experiment.

Prior to surgery, the dogs were fasted for 12 h, and water was withheld for 3 h before the surgery. All dogs received a standardized anesthetic protocol consisting of intravenous (cephalic or saphenous vein) pre-anesthesia with acepromazine (Acepran 0.2%; Univet SA, SP, Brazil) 0.05 mg/kg administered via an over the needle catheter (BD Insyte, Becton Dickinson, Cirúrgicas Ltda, Brazil) that was placed prior to the start of the study. Anesthesia was induced with intravenous propofol (Fresofol 1%; Fresenius Kabi AB, Uppsala, Sweden), based on an expected dose of 4–6 mg/kg and given to allow endotracheal intubation. Anesthesia was maintained with isoflurane (Isoforine; Cristália Chemical and Pharmacological Products, Itapira, Brazil) delivered in 100% oxygen to a rebreathing system with a precision vaporizer out-of-the-circuit.

Mechanical ventilation was volume-controlled, with a tidal volume of 8 mL/kg and a respiratory rate set to maintain end-tidal CO₂ (EtCO₂) between 35 and 45 mm Hg. To maintain normocarbia, pancuronium (Pancuron, Cristália Chemicals and Pharmaceuticals, Itapira, Brazil) (0.06 mg/kg) was intravenously administered in all animals after induction of anesthesia. After completion of the surgical procedure, residual neuromuscular block was reversed using neostigmine (0.04 mg/kg) and atropine (0.02 mg/kg), intravenously. End-tidal gas was sampled from the trachea by inserting a flexible polyvinyl feeding tube through a port in the Y piece of the breathing circuit into the lumen of the endotracheal tube (Dixtal; DX 2021, Dixtal Biomédica Ind e Com, Ltda, Manaus, Brazil). The other end of the feeding tube was connected to the monitor to obtain samples that were reflective of alveolar gas concentration. The animals' temperatures were maintained at approximately 37°C-38°C with the use of warming blankets (Ortovet; SP, Brazil). All animals received Ringer's lactated solution (5 mL kg/hour) IV as a maintenance fluid. During the anesthetic procedure, oxygen saturation (pulseoximetry), ECG, blood pressure (oscillometry), body temperature, inspired and end-tidal carbon dioxide and isoflurane were monitored.

Once the animals were induced to anaesthesia and monitored, the surgical site clipped, and baseline measurements taken, a 20- or 18-gauge epidural needle (Perican; B Braun, São Gonçalo, Brazil) was inserted in the lumbosacral space. Correct needle placement was confirmed by the hanging-drop method and by noting no resistance during insertion of a 20- or 18-gauge

epidural catheter (Portex epidural catheter, Smiths Medical ASD, Inc., Keene, NH, USA). At L4–L5 (midlumbar level), the epidural catheter was threaded about 8 to 10 cm into the epidural canal, based on the marks of the catheter and the length of the Tuohy needle inserted to reach the epidural space, with negligible resistance through the same needle site. All epidural catheter placements and injections were performed by the same blinded investigator. At the end of each epidural injection, the epidural catheter was flushed with 1 mL of saline and then removed. At the end of the instrumentation period, EtISO was maintained at 1.8%–1.9% for at least 15 min.

All injected epidural volumes were calculated based on the sum of an equivalent volume of saline plus nalbuphine or saline alone, giving a total of 0.36 mL/kg⁴. Twenty-four dogs were randomly assigned to one of the following treatments groups: (1) isoflurane ISO/SS, 0.36 mL/kg of epidural saline 0.9%; (2) ISO/NB3, 0.3 mg/kg of nalbuphine chloridrate (Nubain; Cristália Chemical and Pharmacological Products, Itapira, Brazil); or (3) ISO/NB6, 0.6 mg/kg of nalbuphine chloridrate. In a pilot study, a dose of 0.1 mg/kg of epidural nalbuphine was tested in two animals, but without alteration in the MAC or postoperative analgesia. All epidural injections were given 15 min after anesthetic induction, and all surgeries began within 30 min from anaesthetic induction. To improve drug diffusion, the dogs were positioned in dorsal recumbency for at least 5 min following epidural administration of the drug until the start of surgery.

In this study, the time points used to measure the EtISO that involved noxious stimulus were baseline (T0; time 0) before the administration of the drugs; (T15) after pre-anesthetic and anesthetic induction, and the EtISO was held constant for at least 15 min (stabilization period); (T30) 15 min after epidural injections; (T45; 43 ± 6 min) skin incision and subcutaneous tissue dissection; (T60; 54 ± 8 min) midline skin incision; (T75; 65 ± 10 min) left and right ovarian pedicles clamping; and (T90; 82 ± 14 min) end of the surgery.

Thirty min after saline (control) or nalbuphine epidural injections and the start of skin incision, the EtISO concentration was increased or decreased by 0.3% until the next stimulus. Approximately 10 to 15 min were allowed between stimuli for anesthetic equilibration, without positive response. An increase in heart rate (≥15%) or mean blood pressure (>100 mm Hg) within 15 min of skin incision indicated the positive response. If a negative response was detected in the first noxious stimuli step (T45, skin incision and subcutaneous tissue dissection), the EtISO concentration was reduced by 0.3% and the surgical procedure was continued until the next step. This procedure was repeated until the end of surgery.

In dogs that had an initial positive response, the EtISO was increased in a reverse order to provide the necessary depth of anesthesia. The dog was required to remain stable at the lowest possible isoflurane concentration between two noxious stimulus periods. The end-tidal concentration at this time was then recorded as the final percentage of isoflurane required. All ovariohysterectomies were performed by the same surgeon, by a routine method by use of the 3-hemostats technique, and the degree of abdominal muscle relaxation (mild, moderate or intense) was evaluated during surgery by the surgeon.

After surgery, inhalant anesthetic was discontinued, with oxygen continued until spontaneous breathing resumed. Tracheal extubation was done when animals no longer tolerated the tracheal tube. All dogs were monitored for clinical signs of postoperative pain. In the control treatment and in both nalbuphine treatments, postoperative monitoring was done closely in the 90 and 120 min and every hour afterwards until 10 h from the epidural injections by the same investigator. Evaluations of analgesia and sedation were made according to the use of a visual analog scale (VAS), on which 0 represents no pain or no sedation and 10 represents to dogs with worst pain possible, and the scale proposed by the Colorado State University Veterinary Teaching Hospital¹⁵, on which 0 represents no pain and 25 the worst pain possible. The additional need for analgesic therapy in all cases in the postoperative period was determined by pain scores of 4 or higher for VAS or 10 or higher for the Colorado scale. In this case, first rescue analgesia was administered with morphine (Dimorf, 10 mg, Cristália Chemical and Pharmacological Products, Itapira, Brazil) (0.5 mg/kg intramuscularly) by the blinded observer. Additionally, patients received tramadol (Tramadon, 10 mg, Cristália Chemical and Pharmacological Products, Itapira, Brazil) (2 mg/kg orally) every 8 h for three days. Analgesia, sedation, heart rate, respiratory rate, and arterial blood pressure were measured at 90 and 120 min after the epidural injections and every 60 min thereafter until the first rescue analgesic.

Motor function was observed for the first 10 h after surgery and the disconnection of inhalant agent until the end of observation period. The dogs were encouraged to stand up and walk around by their owners and anaesthesiologists participating in the experiment. If this was not possible, the dogs were helped to a standing position. If motor function continued to be abnormal after 6 h, motor function was assessed every 2 h until normal motor function was returned as evidenced by no motor weakness, normal gait and the ability to walk unsupported. The range for return of appetite was also assessed by offering food that dogs were accustomed to eating at home. Other side effects such as vomiting, nausea, pruritus, and urinary retention were recorded during the first 12 h following the end of the surgeries in all animals.

Statistical analysis

The quantitative variables were analyzed within groups and among groups by using analysis of variance (ANOVA) for repeated measures, and the post hoc Tukey method was applied. VAS and Colorado score dependent variables were analyzed within and among groups with the nonparametric Friedman's test (Sigma Stat II, Systat Software Inc., San Jose, USA). For all measurements, mean \pm S.D. values or median \pm confidence interval were determined. For all comparisons,p<0.05 was considered statistically significant.

Results

Twenty-four female dogs were randomized to three treatments as follows: eight to ISO/SS, eight to ISO/NB3, and eight

to ISO/NB6. Mean duration of surgery was 45 \pm 8 min (mean \pm S.D.) and did not differ significantly among treatments (Table 1). A high degree of abdominal relaxation was reported in all treatments by the surgeon.

The mean final EtISO requirements for the ISO/SS, ISO/NB3, and ISO/NB6 treatments, respectively, were $1.98 \pm 0.10\%$, $1.46 \pm 0.10\%$, and $1.22 \pm 0.10\%$ after the first noxious stimulus (T45; Table 2). EtISO was monitored throughout the anesthetic period until the end of surgery (30–90 min). These values differed significantly among treatments (p<0.05). Compared with the control treatment, EtISO decreased significantly at the time of ovarian clamp (T90) with the ISO/NB3 treatment (26.3% reduction) and with the ISO/NB6 treatment (38.4% reduction). There was a significant reduction (16.4%) between the treatments ISO/NB3 and ISO/NB6.

TABLE 1 - Mean ± SD values for demographic data and variables recorded during anesthesia and recovery from anesthesia in 24 dogs premedicated with acepromazine, induced with propofol and anesthetized with isoflurane that underwent ovariohysterectomy after epidural administration of one of three treatments (n=8).

| Variable | ISO/SS (control) | ISO/NB3 | ISO/NB | |
|----------------------------------------------|------------------|----------------|----------------|--|
| Body weight (kg) | 18 ± 5 | 20 ± 7 | 21 ± 6 | |
| Age (months) | 36 ± 10 | 30 ± 6 | 28 ± 8 | |
| Incision (cm) | 12 ± 2 | 13 ± 2 | 11 ± 3 | |
| Duration of surgery (min) | 47 ± 5 | 50 ± 8 | 49 ± 5 | |
| Body temperature (°C) | 38.0 ± 0.6 | 37.4 ± 0.9 | 37.4 ± 0.7 | |
| EtCO ₂ (mm Hg) | 40 ± 5 | 41 ± 3 | 41 ± 4 | |
| SpO_2 | 99 ± 1 | 100 ± 1 | 99 ± 1 | |
| Interval until first rescue analgesic (min)* | 30–60** | 240–300 | 540–600 | |

ISO/SS, epidural saline solution; ISO/NB3, epidural 0.3 mg/kg of nalbuphine; ISO/NB6 epidural 0.6 mg/kg of nalbuphine; EtCO₂, end-tidal CO₂; SpO₂, arterial oxygen saturation

^{*}Interval from epidural administrations until event.

^{**}Approximately within the range suggested.

TABLE 2 - End-tidal, respiratory rate and cardiovascular values during isoflurane anesthesia in association with epidural nalbuphine in 24 dogs premedicated with acepromazine and induced with propofol that underwent ovariohysterectomy.

| Variables | Treatments | T0 | T15 | T30* | T45 | T60 | T75 | T90 |
|-----------|------------|---------------|---------------|---------------|------------------------|---------------------|------------------------|--------------------|
| EtISO | ISO/SS | 0.0 ± 0.0 | 1.9 ± 0.1 | 1.8 ± 0.2 | 2.1 ± 0.1 | 2.1 ± 0.1 | 2.0 ± 0.2 | 1.9 ± 0.1 |
| | ISO/NB3 | 0.0 ± 0.0 | 1.8 ± 0.1 | 1.9 ± 0.1 | $1.5\pm0.3^{\text{a}}$ | $1.3\pm0.3^{\rm a}$ | $1.3\pm0.2^{\text{a}}$ | 1.3 ± 0.1^{a} |
| | ISO/NB6 | 0.0 ± 0.0 | 1.7 ± 0.3 | 1.8 ± 0.1 | 1.3 ± 0.1 ab | 1.0 ± 0.1 ab | 1.0 ± 0.1 ab | 1.0 ± 0.1^{ab} |
| HR | ISO/SS | 108 ± 20 | 109 ± 19 | 112 ± 8 | 114 ± 8 | 111 ± 8 | 108 ± 12 | 112 ± 9 |
| | ISO/NB3 | 111 ± 20 | 101 ± 7 | 108 ± 14 | 104 ± 12 | 108 ± 11 | 108 ± 12 | 113 ±14 |
| | ISO/NB6 | 118 ± 19 | 110 ± 17 | 94 ± 18 | 110 ± 17 | 111 ± 12 | 114 ±19 | 115 ± 17 |
| SAP | ISO/SS | 104 ± 10 | 102 ± 12 | 100 ± 10 | 105 ± 10 | 107 ± 12 | 102 ± 8 | 89 ± 19 |
| | ISO/NB3 | 108 ± 16 | 98 ± 9 | 99 ± 13 | 109 ± 17 | 120 ± 27 | 112 ± 17 | 109 ± 12 |
| | ISO/NB6 | 113 ± 13 | 111 ± 18 | 93 ± 23 | 95 ± 12 | 112 ± 15 | 101 ± 13 | 102 ± 15 |
| DAP | ISO/SS | 64 ± 6 | 49 ± 19 | 46 ± 13 | 61 ± 13 | 61 ± 12 | 55 ± 8 | 49 ± 9 |
| | ISO/NB3 | 63 ± 13 | 51 ± 13 | 50 ± 12 | 61 ± 16 | 70 ± 26 | 63 ± 12 | 58 ± 10 |
| | ISO/NB6 | 69 ± 16 | 56 ± 14 | 54 ± 16 | 56 ± 12 | 73 ± 7 | 65 ± 11 | 65 ± 6 |
| MAP | ISO/SS | 77 ± 6 | 63 ± 9 | 63 ± 11 | 76 ± 11 | 75 ± 13 | 69 ± 6 | 64 ± 4 |
| | ISO/NB3 | 79 ± 15 | 69 ± 14 | 68 ± 14 | 78 ± 15 | 87 ± 23 | 81 ± 10 | 76 ± 9 |
| | ISO/NB6 | 87 ± 15 | 76 ± 15 | 71 ± 18 | 72 ± 11 | 88 ± 7 | 77 ± 7 | 80 ± 7 |
| RR | ISO/SS | 18 ± 3 | 19 ± 5 | 21 ± 2 | 20 ± 6 | 20 ± 4 | 22 ± 4 | 19 ± 6 |
| | ISO/NB3 | 24 ± 7 | 22 ± 4 | 22 ± 3 | 25 ± 8 | 23 ± 6 | 23 ± 5 | 23 ± 7 |
| | ISO/NB6 | 28 ± 5 | 25 ± 5 | 23 ± 3 | 23 ± 6 | 24 ± 7 | 24 ± 5 | 22 ± 8 |

T0, time 0; T15, preanesthetic and anesthetic induction; T30*, epidural injections; T45, skin incision and subcutaneous tissue dissection; T60, midline skin incision; T75, left and right ovarian pedicles clamping; and T90, end of the surgery.

All dogs required the rescue analgesia (VAS score ≥ 4 or Colorado score ≥ 10) at any time point of the postoperative period. The VAS score determined when dogs meet the criteria for rescue analgesia between the two nalbuphine treatments, and it was higher compared to the control treatment. The highest VAS scores in the control treatment were recorded 90 min after surgery (median, 2; range, 0 to 4.0). The time to first rescue analgesic was also significantly different (p<0.05) between the nalbuphine treatments when compared to the control treatment and between the ISO/NB3 treatment (240-300 min) versus ISO/ NB6 (540-600 min) (Figure 1; p<0.05). For sedation values, according to the same criteria (VAS scores), a decrease in both treatments with nalbuphine was observed with time (Figure 2). For analgesia and sedation, the Colorado score had the highest median value 90 min after surgery in the control treatment (median, 8.4; range 7–10) and was significantly lower for dogs

receiving the ISO/NB3 or ISO/NB6 treatment. The Colorado scores were significantly lower in the ISO/NB6 treatment, compared with values for the ISO/NB3 treatment, at 240, 480, and 600 min (Figure 3).

The heart rate, $EtCO_2$, arterial oxygen saturation (SpO₂), and arterial pressure (systolic arterial pressure, diastolic arterial pressure, and mean arterial pressure) did not change significantly among treatments or when compared with the basal values following epidural administration of ISO/SS, ISO/NB3 or ISO/NB6 treatments until the end of anaesthesia. (Tables 1 and 2). Body temperature was kept stable throughout anesthesia and surgery. There were no differences regarding the incidence of postoperative adverse effects. During the first 12 h following surgery none of the dogs had nausea or vomiting. Return to appetite or drinking water was delayed in control dogs (mean \pm S.D., 13 ± 4 h), compared with dogs receiving ISO/NB3 (5 ± 2 h) or ISO/NB6 (3 ± 1 h).

ISO/SS, epidural saline solution (0.36 ml/kg); ISO/NB3, epidural 0.3 mg/kg of nalbuphine; and ISO/NB6 epidural 0.6 mg/kg of nalbuphine.

EtISO, end-tidal isoflurane (%); HR, heart rate (beats/min); SAP, systolic arterial pressure (mm Hg); DAP, diastolic arterial pressure (mm Hg); MAP, mean arterial pressure (mm Hg); RR, respiratory rate (breaths/min).

The values for ISO/SS, ISO/NB3 and ISO/NB6 differ significantly (p<0.05) from the baseline (B1-MAC_{BAB}); time 0) values.

^bSignificant differences among treatments were detected (p<0.05).

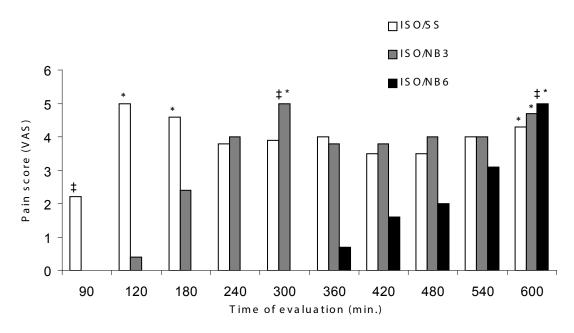


FIGURE 1 - Median pain scores from dogs administered epidurally saline (ISO/SS; 0.36 mL/kg), nalbuphine 0.3 mg/kg (ISO/NB3) or nalbuphine 0.6 mg/kg (ISO/NB6) determined between 90 and 600 min following ovariohysterectomy. Pain was scored using visual analogue system (VAS) on a scale of 0–10; increased scores were indicative of greater pain. ‡Time of the first analgesic rescue. *Significantly (p<0.05) different from value at 90 min.

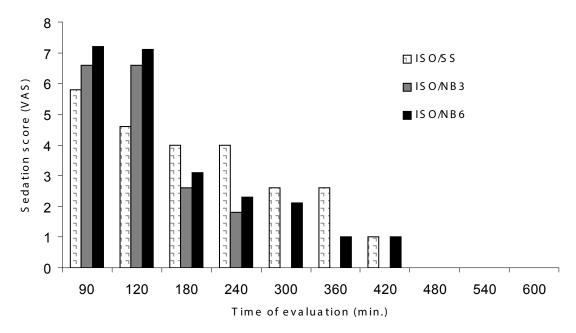


FIGURE 2 - Median sedation score in epidural saline (ISO/SS; 0.36 mL kg⁻¹), nalbuphine 0.3 mg/kg (ISO/NB3) or nalbuphine 0.6 mg/kg (ISO/NB6) in dogs 90–600 min following ovariohysterectomy. Sedation was scored using visual analogue system (VAS, 0–10), where 10 is deep sedation and 0 represents agitation.

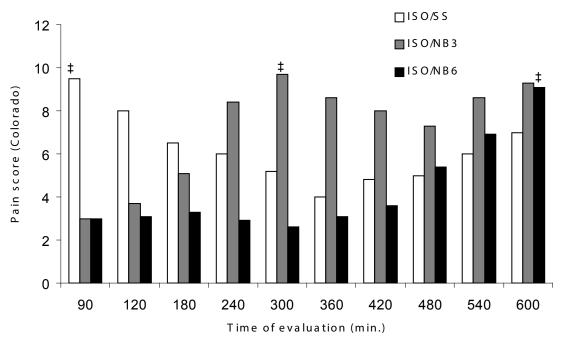


FIGURE 3- Median pain and sedation assessment (Colorado State University Veterinary Teaching Hospital scale) in the same dogs as in Figures 1 and 2. In this proposed score on a scale of 0–25; increased scores were indicative of greater pain and deep sedation. ‡Time of the first analysesic rescue. *Significantly (p<0.05) different from value at 90 min.

Discussion

This study showed a sparing effect when the inhalant anesthetic agent isoflurane was combined with nalbuphine administered by the epidural route to dogs that underwent ovariohysterectomy. The MAC of isoflurane in our dogs receiving saline solution was 1.98% at approximately 2 h, which was higher than that measured by others for this species^{16,17}. The ovarian stimulation model is an adequate and repeatable means of producing visceral stimulation to determine MAC in dogs¹⁸. Previous studies used electrical stimulation or mechanical stimuli (tail clamp) based on clinical signs such as palpebral response, jaw tone, and vigour of spontaneous movement in response to noxious stimulation, indicating that these dogs were not deeply anesthetized at the measured concentrations 16,17,19,20. Movement at skin incision has been proposed for determining the potency of intravenous anaesthetics²¹; however, it is also used as a standard measure for assessment of inhalant anesthetic agents²².

Currently, clinical and laboratory studies demonstrate the need to treat pain according to individual needs, pain source, pain mechanisms and drug properties²³. Epidural opioids have been shown to decrease the MAC of inhalant anaesthetics in dogs in a number of studies^{1,2,7,24}. Epidural methadone significantly reduces the isoflurane requirement compared with intravenous methadone during femoro-tibial joint surgery². In experimental studies

using electrical stimulus, morphine and methadone administered epidurally in dogs reduced the anaesthetic requirement for halothane and isoflurane, respectively^{1,24}. In our study, the MAC of isoflurane in dogs was reduced by 26.3% and 38.4% by epidural nalbuphine at 0.3 and 0.6 mg/kg, respectively, when compared with the saline treatment. When compared between nalbuphine treatments, the reduction was 16.4%. The nalbuphine administered intravenously in dogs with a wide dose range (0.5, 1.5, 5.0, and 20.0 mg/kg) produced significant reduction of enflurane MAC (8%) with the lowest dose, with no additional reduction at higher doses²⁵. To our knowledge, this is the first clinical study to verify that nalbuphine epidural administration reduces isoflurane requirements in surgical procedures in dogs. Previous studies with epidural nalbuphine in humans were only related to the control of postoperative pain^{13,26} and the avoidance of side effects induced by other epidural opioids²⁷.

Our study showed a dose-dependent analgesic effect of 0.3 and 0.6 mg/kg epidural nalbuphine compared with saline control in dogs undergoing ovariohysterectomy. This was reflected by the time required to first rescue analgesia (4.5 h compared with 9.5 h following nalbuphine 0.3 mg/kg or 0.6 mg/kg, respectively). Both nalbuphine treatments were associated with significantly lower pain scores on the VAS and Colorado pain scales compared with the control treatment in the first 10 h after epidural injections. The analgesia mediated by epidural nalbuphine is probably caused by potent μ -antagonist and κ -agonist opioid activity. Methadone

administered by the epidural route to dogs that underwent femorotibial joint surgery had analgesia duration of approximately 7.5 h based on a numerical pain scoring system². These differences in the duration of analgesia may be related to the different drugs, and different subjective pain scales used in our study and those of others^{2,24}. The cortisol concentration is objective for determining physiologic or neurohumoral stress response in dogs; however, routine measurement of cortisol is clinically impractical²⁸.

One of the main advantages of epidural administration of opioids is that a small dose of the drugs provides pain relief in the postoperative period. However, there may be side effects. Side effects that occur more frequently after administration of epidural opioids are respiratory depression, pruritus, nausea, vomiting and urinary retention^{1,7,12,13}. Studies on opioids alone or in combination with local anaesthetics report conflicting results on respiratory depression. For instance, one study showed that epidural administration of morphine in dogs anesthetized with halothane was not associated with respiratory depression or other side effects¹. However, in another study respiratory depression was found to occur after epidural oxymorphone in dogs anesthetized with halothane⁷. Nalbuphine's μ-antagonism should produce fewer μ-mediated side effects such as respiratory depression¹³. In our study with nalbuphine epidural, although respiratory function was not thoroughly investigated, respiratory rate, SpO2, and EtCO2 remained within the physiological range. Mechanical ventilation used during general anesthesia, contributed to the respiratory parameters remained within the physiological parameters.

Despite systemic absorption and supraspinal effects, epidural administration of opioids is not expected to have a significant cardiovascular effect in healthy patients. Epidural administration of morphine in halothane- or isoflurane-anaesthetized dogs did not induce significant cardiovascular alterations^{1,29}. But other studies with other opioids have reported conflicting results for cardiovascular parameters. Epidural administration of oxymorphone in combination with bupivacaine in halothane-anesthetized dogs showed decreased heart rate and blood pressure⁷. Another study showed a decrease in heart rate after epidural methadone in isoflurane-anesthetized dogs, but an increase in blood pressure (systolic arterial pressure)²⁴. We did not observe any alteration in the cardiovascular system with either dose of nalbuphine epidural in the present study during the anaesthetic period.

Conclusions

The nalbuphine administered epidurally at 0.3 and 0.6 mg/kg diluted in saline reduced the anesthetic requirement for isoflurane in ovariohysterectomized dogs in a dose-dependent manner. Similarly,

nalbuphine epidural provided improved postoperative analgesia by increasing the time until the first rescue analgesia was needed. There were no side effects observed in this study.

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