

Prevention of rocuronium induced mast cell activation with prophylactic oleuropein rich diet in anesthetized rabbits¹

Tuncer Simsek¹, Mesut Erbas¹¹ Basak Buyuk¹¹¹, Cigdem Pala¹¹, Hasan Sahin¹, Betul Altinisik¹¹

¹Assistant Professor, Department of Anesthesiology and Reanimation, Medical Faculty, Canakkale Onsekiz Mart University, Turkey. Design of the study, acquisition of data, manuscript preparation.

"Associate Professor, Department of Anesthesiology and Reanimation, Medical Faculty, Canakkale Onsekiz Mart University, Turkey. Conception and design of the study.

"Assistant Professor, Department of Histology, Canakkale Onsekiz Mart University, Turkey. Histopathological examinations.

^{IV}Associate Professor, Department of Food Engineering, Engineering Faculty, Canakkale Onsekiz Mart University, Turkey. Interpretation of data, technical procedures.

^vAssistant Professor, Department of Anesthesiology and Reanimation, Medical Faculty, Canakkale Onsekiz Mart University, Turkey. Acquisition and interpretation of data.

viAssistant Professor, Department of Anesthesiology and Reanimation, Medical Faculty, Canakkale Onsekiz Mart University, Turkey. Analysis and interpretation of data.

Abstract

Purpose: The effect of a prophylactic oleuropein-rich diet before anesthesia accompanied by the widely-used steroid-based neuromuscular drug rocuronium on mast cell activation was investigated in the study.

Methods: 14 rabbits used in the study. The rabbits in the oleuropein group were given oleuropein-rich extract added to the animals' water at doses of 20 mg/kg oleuropein for 15 days orally. After 15 days, all rabbits in the two groups were given general anesthesia with rocuronium of 1 mg/kg. After 1 day, animals were sacrificed and the liver tissue sections stained with H&E, toluidine blue and tryptase for immunohistochemical study.

Results: There was no statistically significant difference between ALT, AST and albumin averages of the oleuropein and control groups (p> 0.05). The tryptase average of the control group was higher than the tryptase average of the oleuropein group and this difference was statistically significant (p=0.003). The T. blue average in the oleuropein group was higher than the control group. However, there was no statistically significant difference between groups (p=0.482).

Conclusions: Rocuronium adverse effects, like hypersensitivity and anaphylaxis, may limit routine use of this substance. The use of oleuropein reduced the number of inflammatory cells and prevented degranulation.

Key words: Anesthesia. Mast Cells. Rabbits.

Introduction

olive tree contains phenolic compounds with important biological properties and oleuropein is considered the most important of these phenolic compounds¹. The pharmacological benefits of oleuropein in preventing and protecting against diseases are known. Oleuropein has been shown in a variety of scientific studies to have antiinflammatory effects in addition to antioxidant, antihistaminic, cardioproductive and hypolipidemic effects, the effect of inhibiting thrombocyte aggregation, anti-atherogenic effect, anticancerogenic effect, antimicrobic properties against gram-positive and gramnegative bacteria, neuroprotective effect and antiviral effects²⁻⁸. Previous studies have shown that hydroxytyrosol and oleuropein are the most important phenolic components in olive plants9. These phenolic components inhibit mast cell degranulation via induced immune and non-immune pathways. It has been reported that hydroxytirozole and oleuropein may shed light on researchs in terms of preventing mast cell mediated disorders and developing useful tools for their treatment¹⁰.

Mast cells play an important role in the development of allergic reactions due to the release of histamine, cytokines and proteolytic enzymes¹¹. As mast and T cells interact with each other, mast cells especially increase proliferation of T cells and secretion of cytokines¹². Thus, events leading up to the reactions that result in anaphylaxis.

Rocuronium is a widely used steroidal neuromuscular muscle relaxant that is used to ease endotracheal intubation and mechanical ventilation during general anesthesia¹³. Rocuronium often causes mast cell mediated hypersensitivity reactions. Neuromuscular blocker (NMB) drugs are the most important cause of anaphylactic reactions (38.1% of all cases) during general anesthesia. Rocuronium

(50% of cases) is responsible for most anaphylactic reactions related to NMB drugs¹⁴.

The aim of this study is to demonstrate the effect of prophylactic administration of oleuropein for the prevention of mast cell aggregation and degranulation after rocuronium administration given for neuromuscular blockage during general anesthesia.

Methods

Study groups and diet protocol

All the protocols described here are in accordance with the European Communities Council Directive (86/609/EEC). Ethics committee approval was gained from the local animal experiment.

Fourteen adult male New Zealand rabbits weighing between 2.7 and 3.8 kg were used for the study. The rabbits were clinically evaluated for behavior, respiratory and cardiovascular system problems and no negative result was found for animals included in the study. All experiments took place between 09.00 and 16.00. During the experiments the animals were fed with standard rabbit food and were given continuous access to water. The temperature of the shelter was kept at 21±2°C.

The animals were randomly divided into two groups: Group C (Control, n:7) were fed with normal diet and water, and group O (Oleuropein, n:7) were fed with normal diet and water containing 20 mg/kg/day of oleuropein. Both groups were fed for 15 days. Oleuropein was extracted from olive leaves as described by Simsek *et al.*¹⁵.

Analysis of oleuropein in olive leaf extract (OLE)

A HPLC (High Pressure Liquid Chromatography) assisted method was used

for analysis of oleuropein content of OLE¹⁶. OLE sample was diluted (1:10, v:v) with ultrapure water and filtered through 0.45µm PTFE syringe filter before injection to HPLC system. With a sample loop of 20µL OLE injected to a photodiode array detector (SPD-M20A, Shimadzu, Japan). Separation of oleuropein was carried on Inertsil ODS 3 (250mm, 4.6ID, 5µ) reverse-phase column under isocratic condition with 80:20 v/v water:acetonitrile (adjusted to pH 3 with acetic acid) as a mobile phase. Column was maintained at 35°C during chromatographic run. Flow rate was 1 mL/min and the DAD detector was set at 200-400 nm to analyze the spectra. Quantification was made at 280 nm using external standard method. Results were expressed as mg of oleuropein/L of OLE based on seven-point standard curve prepared by the oleuropein standard (Sigma-Aldrich, Germany) solutions range in 10-1000 mg/L concentration.

General anesthesia protocol

Rabbits in the study were fasted for 8 hours before the general anesthesia. All rabbits were administered 10 mg/kg ketamine for premedication before the general anesthesia. After 15 minutes, the animals were monitored with ECG. Then a vein was opened in the ear with 26 G branula and 4 L/min O₂ was administered through the mask.

During the general anesthesia, pulse and oxygen saturation measured from the ear by a pulse oximetry device. Body temperature was monitored and held at 37-39°C during the anesthesia.

V-Gel Rabbit (V-gel rabbit R-3 Docsinnovent® Ltd. London, UK) supraglottic airway was inserted to provide reliable airway after the intravenous administration of 2mg/kg ketamine and 1 mg/kg rocuroium bromide. Later the animals were linked to an anesthetic device (Anesthesia Machine w/

O, Flush Model M3000PK Parkland Scientific Lab And Research Equipment. Florida, USA) and ventilated manually. 1 MAC isoflurane used to maintain anesthesia and 50% oxygen, 50% air mix was used with. The rabbits were manually ventilated by the same anesthetist to a respiration count of about 40/minute and pressure of 15 cmH₂O (about 10ml/kg) appropriate for rabbit physiology. The animals were also monitored with pulse oximeter during the anesthesia procedure. General anesthesia duration determined as 30 minute to ensure the data integrity. V-Gel Rabbit was removed and animals were taken to recovery after the observation of spontaneous respiration at sufficient levels.

Euthanasia and sample collection

All animals were observed for 24 hours following rocuronium administration and then underwent euthanasia remifentanil anesthesia. After euthanasia, blood samples were taken for AST, ALT, and albumin analyses and the liver tissues were harvested and stored in 10% neutral formalin solution. Tissues underwent routine tissue processing protocols for light microscopy and were embedded in liquid paraffin. Tissue sections were cut to a thickness of 4 um with a microtome (Leica RM2255; Leica Microsystems, Wetzlar, Germany) and were stained with routine hematoxylin-eosin (HE) and toluidine blue. HE stained sections were graded using the following scoring system for inflammatory cell infiltration; 0, no infiltration; 1, mild infiltration; 2, moderate infiltration; 3, severe infiltration. Tryptase positive cells were investigated using the immunohistochemical method. Two slides from each rabbit were prepared and five portal triads were examined on each slide. Total and tryptase-positive mast cells in all groups were counted as described by Tomak et al.17.

Statistical analysis

Statistical analysis was performed by using the SPSS software version 20 (SPSS 20 for Windows, IBM, Chicago, Illinois, USA). Mann—Whitney U test used to compare the differences between groups. Data were expressed as the mean ± standard deviation. P value less than 0.05 considered as statistically significant.

Results

None of the rabbits had signs and symptoms of any infection, anaphylactic or allergic reactions during the study period.

The average weight of rabbits in the control group was 3541±150g and the average

weight of OLE group rabbits was 3288±428g with no statistically significant difference. The oxygen saturation levels, mean arterial pressure and heart rate values in both groups were similar during the observation.

The rabbits in study had daily water consumption of 80-100 ml/kg/day calculated according to weight. The amount of oleuropein in OLE was calculated as 4.31±0.18 mg/ml with the amount of OLE found according to rabbit weight for an oleuropein dose of 20 mg/kg.

There were no significant differences between the 2 groups for alanine aminotransferase (ALT), aspartate aminotransferase (AST) and albumin (ALB) mean levels (p>0.05) which were obtained from blood samples (Table 1).

Table 1 - Comparison of the groups for ALT, AST, ALB.

	Group O (n=7)		Group C (n=7)		
Variables	Mean±SD	Median (min/max)	Mean±SD	Median (min/max)	р
ALT	32.5±12.4	30.0 (15.4-54.8)	27.0±7.6	28.0 (18.4-39.6)	0.443
AST	21.7±13.3	15.9 (10.8-46.1)	14.3±3.4	15.1 (8.3-17.9)	0.654
ALB	4.3±0.9	4.2 (3.0-5.9)	4.4±1.2	4.5 (2.2-6.1)	0.848

HE stained sections showed preserved histological tissue architecture in both control and OLE groups, with normal histological structure of hepatocyte cords and vena sentralis on microscopic examinations. The histological appearance of portal triads regions was normal and no destruction was

found. However, inflammatory cell infiltrations were found especially in portal areas in control group rather than the OLE group. According to scoring system, grade 1 inflammatory cell infiltration observed in OLE group when compared to grade 3 infiltration in control group (Figure 1).

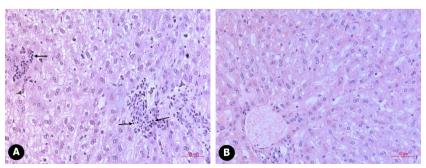


Figure 1 - Microscopic comparison of routine histopathologic changes with Heatoxylin & Eosin staining shown in **A** (group C) and **B** (group O) (Magnification x4). Inflamatuar cell infiltrations are significant in **A** than **B**.

Mean mast cell numbers in Group O were significantly higher than Group C for toluidine blue staining (1774.4±336.0 vs. 1673.6±291.4, respectively), though there was no significant difference between the two groups statistically (p=0.482) (Figure 2).

Tryptase positive cell mean value in group C was significantly higher than group O and the difference was statistically significant (1169.6±137.5 vs. 775.1±180.0, respectively, p=0.003) (Table 2, Figure 3).

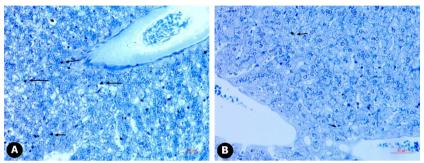


Figure 2 - Microscopic comparison of Toluidine blue staining was performed in both groups for evaluating the mast cell numbers. **A** (group C) and **B** (group O) (Magnification x10) are Toluidine blue stainings of group C and group O sections, respectively. Mast cell accumulation is significant in **A**.

Table 2 - Comparison of the groups in Tryptase and Toluidine Blue.

	Gro	oup O (n=7)	Group C (n=7)		
Variables	Mean±SD	Median (min/max)	Mean±SD	Median (min/max)	р
Triptase	775.1±180.0	792.0 (548.0-1053.0)	1169.6±137.5	1149.0 (949.0-1361.0)	0.003
T-Blue	1774.4±336.0	1860.0 (1304.0-2147.0)	1673.6±291.4	1645.0 (1201.0-2062.0)	0.482

SD: Standart Deviation p: Mann Whitney U Test

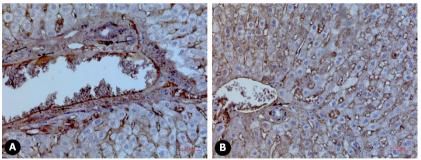


Figure 3 - Microscopic comparison of immunohistochemical evaluation for triptase positive cells is shown in **A** (group C) and **B** (group O) (Magnification x20). Group O demonstrated less count of triptase positive cells.

Discussion

Mast cell aggregation and degranulation effects of rocuronium are known from

previous research¹⁸. Sugammadex can be used to reverse the effects of rocuronium. Tomak *et al.*¹⁷ stated that sugammadex is beneficial for the treatment or prevention of

rocuronium-induced anaphylaxis. However Menéndez-Ozcoidi et al.19 reported allergic reaction after the administration of sugammadex (3.2 mg/kg) to reverse neuromuscular block induced by rocuronium. Tomak et al.17 found that sugammadex could increase the numbers of mast cells. This situation can lead to a predisposition for allergic reactions. Nevertheless, the rocuroniuminduced increase in mast cells is significantly higher than the sugammadex-induced increase in mast cells. According to these findings a preoperative diet formulation could be used to prevent allergic reactions and anaphylaxis rather than drugs which could induce histamine discharge. Previous research has indicated the antiallergic and anti-hystaminic effects of different medicinal plants. Brazilian pepper tree (Schinusterebinthifolius Raddi), Ximenia Americana and Oleuropein are some of the flavonoids previously studied for the inhibition of mast cell degranulation²⁰⁻²². Oleuropein is one of the most studied flavonoid for different medical conditions. The extract is derived from Olea Europea tree leaves which is containing the phenolic compound Oleuropein in high density1.

Recent studies related to Oleuropein is especially focused on its anti allergic and anti inflammatory effects²³. Using this phenolic compound for the prevention of rocuronium induced mast cell activation before the general anesthesia and surgical operation process is a new and continuing stage of the Oleuropein studies.

A variety of studies were taken as reference for the dose of oleuropein used in rabbits. Different animal studies have reported a range of calculations related to the dose of oleuropein that should be administered^{24,25}. According to these calculations, as antioxidant, anti-inflammatory and vasodilatatory effects are clearly observed with doses of 20

mg/kg, we chose to use an OLE formulation containing the same dose of oleuropein. Olive leaf extracts (OLEs) were stored at -20°C to prevent degradation of oleuropein during the experiment. All rabbits in the OLE group consumed the extract mixed with water. Oral tolerance of OLE appeared successful. A study of rats reported the intestinal absorption of oleuropein was rapid with a 20 mg/kg oral dose reaching peak plasma concentration in 2 hours²⁶.

At non-toxic doses oleuropein inhibits the release of hexosaminidase from peritoneal mast cells playing a role as mast cell stabilizer and additionally some in vitro studies have shown that a variety of phenolic diets inhibit mast cell activation^{27,28}. When cells are exposed to calcium ionophore A23187, it is reported that oleuropein inhibits mast cell degranulation more efficiently than sodium cromoglycate and hydroxytyrosol. It is reported that polyphenols dissolve proteins or form insoluble complexes and change the structure of allergenic proteins or reduce their bioavailability to render them hypoallergenic²². In terms of the effect mechanism of oleuropein, a different study reported that oleuropein blocked the phosphorylation of p38 mitogen-activated protein kinase through the up-regulation of annexin A1. It is a potent anti-inflammatory calcium-dependent phospholipid protein also an endogenous mediator of glucocorticoid signaling downstream signal pathways of systolic calcium increase²⁹.

Animals were sacrificed and livers were harvested for tissue preparation 24 hours after general anesthesia according to the pharmacokinetics of rocuronium. As reported in the literature, rocuronium is primarily eliminated by the liver. Plasma concentration decreases according to hepatic intake and bile excretion after the administration of rocuronium and nearly a third of the

rocuronium in circulation is expected to be excreted in urine in 24 hours^{30,31}.

IgE receptors are present on the mast cell membranes. When IgE-coated antigens bind to surface receptors, mast cell degranulation occurs. Increased mast cell tryptase has a central role in inflammatory and immediate allergic reactions initiated by immunoglobulins IgE³².

IgE antibodies generated by are B-lymphocytes in response to the exposure to a specific antigen like rocuronium molecule. IgE molecules bind to high affinity IgE receptors situated on the surface of mast cells and leading to mast cell degranulation, releasing both preformed and newly synthesised Under normal circumstances. mediators. these mediators help for the development of defensive acute inflammatory reactions. When there is massive release of these mediators, bronchoconstriction and vasodilation predominate³². In our study evaluation with toluidine blue staining in all rabbits within the two groups showed significantly increased mast cell counts in hepatic portal areas. This finding supports the results of previous studies about rocuroniums' mast cell aggregation pattern. Our results found that all rabbits in group C had higher numbers of mast cells in hepatic portal areas, in contrast there was no significant difference between the two groups in statistical analysis due to the limited number of rabbits.

Generally, it has been accepted that mast cell tryptase levels show the activation and degranulation of mast cells^{24,25}. For that reason, immunochemical staining methods were used to determine tryptase-positive mast cell counts. Tryptase-positive mast cell numbers were significantly lower in animals receiving rocuronium after an oleuropein-rich diet compared with Group C and this data demonstrates that oleuropein can decrease

mast cell aggregation and degranulation.

Acute toxicity study in rabbits fed with OLE was performed in order to explore any adverse effect resulted from OLE. OLE administered orally revealed no sign of poisoning in rabbits of OLE group. After 15 days of the treatment, no rabbits died and no significant change of weight between the both groups. Several studies have demonstrated that the elevated liver enzymes AST and ALT indicate hepatocellular plasma membrane damage, and these two enzymes are considered good markers for liver inflammation and necrosis. Marmugi et al.33.There was no significant difference between two groups for ALT, AST and ALB levels in biochemical evaluation. This means that neither rocuronium nor oleuropein has any toxic effects on hepatic functions.

Previous research has illustrated the antiallergic effects of oleuropein¹⁴ via the inhibition of mast cell degranulation. Our results support these previous studies with the decrease in tryptase-positive mast cell counts in rabbits fed with an oleuropein rich diet.

Conclusions

This study assessed the histological and immunological effects of a prophylactic oleuropein-rich diet in terms of mast cell numbers and degranulation levels in liver cells developing after intravenous administration of rocuronium in rabbits undergoing general anesthesia. Our findings show that the use of oleuropein-rich diet reduced the number of inflammatory cells and prevented degranulation. These findings show that using prophylactic oleuropein-rich diet would be useful to improve preoperative anesthesia approaches to prevent anaphylactic reactions according to the administered neuromuscular

blockers as rocuronium during the general anesthesia.

References

- 1- Japon-Lujan, R, J. Luque-Rodríguez JM, Luque de Castro MD. Dynamic ultrasound assisted extraction of oleuropein and related biophenols from olive leaves. J Chromatogr A. 2006;1108:76–82. doi: 10.1016/j. chroma.2005.12.106.
- 2- Andreadou I, Iliodromitis EK, Mikros E, Constantinou M, Agalias A, Magiatis P, Skaltsounis AL, Kamber E, Tsantili-Kakoulidou A, Kremastinos DT. The olive constituent oleuropein exhibits anti-ischemic, antioxidative, and hypolipidemic effects in anesthetized rabbits. J Nutr. 2006;136:2213–9. PMID: 16857843.
- 3- Omar S.H. Oleuropein in olive and its pharmacological effects. Sci Pharm. 2010;78:133-54. doi: 10.3797/scipharm.0912-18.
- 4- Visioli F, Galli C. Antiatherogenic components of olive oil. Curr Atheroscler Rep. 2001;3:64–7. doi: 10.1007/s11883-001-0012-0.
- 5- Hamdi HK, Castellon R. Oleuropein, a nontoxic olive iridoid, is an anti-tumor agent and cytoskeleton disruptor. Biochem Biophys Res Commun. 2005;334:769–78. doi: 10.1016/j.bbrc.2005.06.161.
- 6- Bazoti FN, Bergquist J, Markides K, Tsarbopoulos A. Noncovalent interaction between amyloid-β-peptide (1–40) and oleuropein studied by electrospray ionization mass spectrometry. J Am Soc Mass Spectrom. 2006;17:568–75. doi: 10.1016/j.jasms.2005.11.016.
- 7- Kiraz A, Simsek T, Tekin SZ, Elmas S, Tekin M, Sahin H, Altinisik HB and Pala C. Investigation of the effects of oleuropein rich diet on rat enteric bacterial flora. Bratisl Lek Listy. 2016;117: 734-7. doi: 10.4149/BLL_2016_141.
- 8- Ahmadvand H, Shahsavari G, Tavafi M, Bagheri S, Moradkhani MR, Kkorramabadi RM, Khosravi P, Jafari M, Zahabi K, Eftekhar R. Protective effects of oleuropein against renal injury oxidative damage in alloxan-

- induced diabetic rats; a histological and biochemical study. J Nephropathol. 2017;6:204. doi: 10.15171/jnp.2017.34.
- 9- El SN, Karakaya S. Olive tree (*Olea europaea*) leaves: potential beneficial effects on human health. Nutr Rev. 2009;67(11):632–8. doi: 10.1111/j.1753-4887.2009.00248.x.
- 10-Persia FA, Mariani ML, Fogal TH, Penissi AB. Hydroxytyrosol and oleuropein of olive oil inhibit mast cell degranulation induced by immune and non-immune pathways. Phytomedicine. 2014;21:1400–5 doi: 10.1016/j.phymed.2014.05.010.
- 11-Marone G, Genovese A, Granata F, Forte V, Detoraki A, De Paulis A, Triggiani M. Pharmacological modulation of human mast cells and basophils Clin Exp Allergy. 2002;32(12):1682-9. doi: 10.1046/j.1365-2222.2002.01535.x.
- 12-Nakae S, Suto H, Kakurai M, Sedgwick JD, Tsai M, Galli SJ. Mast cells enhance T cell activation: importance of mast cell derived TNF. Proc Natl Acad Sci USA. 2005;102(18):6467-72. doi: 10.1073/pnas.0501912102.
- 13-Bom A, Bradley M, Cameron K, Clark JK, Egmond JV, Feilden H, MacLean EJ, Muir AW, Palin R, Rees DC, Zhang MQ. A novel concept of reversing neuromuscular block: chemical encapsulation of rocuronium bromide by a cyclodextrin-based synthetic host. Angew Chem Int Ed Engl. 2002;41:266–70. doi: 10.1002/1521-3773(20020118)41.
- 14-Meng J, Rotiroti G, Burdett E, Lukawska JJ. Anaphylaxis during general anaesthesia: experience from a drug allergy centre in the UK. Acta Anaesthesiol Scand. 2017;61:281-9. doi: 10.1111/aas.12858.
- 15-Şimşek T, Altınışık U, Erşan İ, Şahin H, Altınışık B, Erbaş M, Pala Ç. Prevention of intraocular pressure elevation with oleuropein rich diet in rabbits, during the general anaesthesia. SpringerPlus. 2016;5(1):952. doi: 10.1186/s40064-016-2402-3.
- 16-Al-Rimawi F. Development and validation of a simple reversed-phase HPLC-UV method for determination of oleuropein in olive leaves. J Food Drug Anal. 2014;22:285–9. doi: 10.1016/j.jfda.2013.10.002.
- 17-Tomak Y, Yılmaz A, Bostan H, Tümkaya L, Altuner D, Kalkan Y, Erdivanlı, B. Effects of

- sugammadex and rocuronium mast cell number and degranulation in rat liver. Anaesthesia. 2012;67(10):1101-4. doi: 10.1111/j.1365-2044.2012.07264.x.
- 18-Rose M, Fisher M. Rocuronium: high risk for anaphylaxis? Br J Anaesth. 2001;86(5):678-82. doi: 10.1093/bia/86.5.678.
- 19-Menéndez-Ozcoidi L, Ortiz-Gómez JR, Olaguibel-Ribero JM, Salvador-Bravo MJ. Allergy to low dose sugammadex. Anaesthesia. 2011;66:217–9. doi: 10.1111/j.1365-2044.2010.06611.x..
- 20-Castro Souza Junior Neto J, Estevão LR, Baratella-Evêncio L, Vieira MG, Simões RS, Florencio-Silva R, Evêncio-Luz L, Evêncio-Neto J. Mast cell concentration and skin wound contraction in rats treated with Ximenia americana L. Acta Cir Bras. 2017;32(2):148-56. doi: 10.1590/s0102-865020170207.
- 21-Estevão LR, Medeiros JP, Simões RS, Arantes RM, Rachid MA, Silva RM, Mendonça FS, Evêncio-Neto J. Mast cell concentration and skin wound contraction in rats treated with Brazilian pepper essential oil (Schinus terebinthifolius Raddi). Acta Cir Bras. 2015;30(4):289-95. doi: 10.1590/S0102-865020150040000008.
- 22-Persia FA, Mariani ML, Fogal TH, Penissi AB. Hydroxytyrosol and oleuropein of olive oil inhibit mast cell degranulation induced by immune and non-immune pathways, Phytomedicine. 2014;21:1400–5. doi: 10.1016/j.phymed.2014.05.010.
- 23-Gambino CM, Accardi G, Aiello A, Candore G, DC Giovanni, Mirisola M, Procopio A, Taormina G, Caruso C. Effect of extra virgin olive oil and table olives on the immuneinflammatory responses: potential clinical applications. Endocr Metabol Immune Disord Drug Targets. 2018;18:14-22. doi: 10.2174/187153031766617111411 3822.
- 24-Andreadou I, Iliodromitis EK, Mikros E, Constantinou M, Agalias A, Magiatis P, Skaltsounis AL, Kamber E, Tsantili-Kakoulidou A, Kremastinos D. The olive constituent oleuropein exhibits anti-ischemic, antioxidative, and hypolipidemic effects in anesthetized rabbits. J Nutr. 2006;136:2213–9. doi: 10.1093/jn/136.8.2213.

- 25-Al-Azzawie HF, Saeed Alhamdani MS. Hypoglycemic and antioxidant effect of oleuropein in alloxan-diabetic rabbits. Life Sci. 2006;78:1371–7. doi: 10.1016/j. lfs.2005.07.029.
- 26-Del Boccio P, Di Deo A, De Curtis A, Celli N, lacoviello L, Rotilio D. Liquid chromatographytandem mass spectrometry analysis of oleuropein and its metabolite hydroxytyrosol in rat plasma and urine after oral administration. J Chromatogr B Analyt Technol Biomed Life Sci. 2003;785(1):47-56. PMID: 12535837.
- 27-Middleton E, Kandaswami C, Theoharides C. The effects of plant flavonoidson mammalian cells: implications for inflammation, heart disease, and cancer. Pharmacol Rev. 2000;52:673–751. doi: 0031-6997/00/5204-0673\$03.00/0.
- 28-Singh A, Holvoet S, Mercenier A. Dietary polyphenols in the prevention and treatment of allergic diseases. Clin Exp Allergy. 2011;41:1346–59. doi: 10.1111/j.1365-2222.2011.03773.x.
- 29-Giner E, Andújar I, Recio MC, Ríos JL, Giner RM. Oleuropein protects against dextran sodium sulfate-induced chronic colitis in mice. J Nat Prod. 2013;13:1113–20. doi: 10.1021/np400175b.
- 30-Proost JH, Eriksson LI, Mirakhur RK, Roest G, Wierda JM. Urinary, biliary and faecal excretion of rocuronium in humans. Br J Anaesth. 2000;85:717–23. PMID: 11094587.
- 31-Mirakhur RK. Safety aspects of nondepolarizing neuromuscular blocking agents with special reference to rocuronium bromide. Eur J Anaesthesiol. 1994;9:133– 40. PMID: 7523105.
- 32-Payne V, Kam PC. Mast cell tryptase: a review of its physiologyand clinical significance. Anesthesia. 2004;59:695-703. doi: 10.1111/j.1365-2044.2004.03757.x.
- 33-Marmugi A, Ducheix S, Lasserre F, Polizzi A, Paris A, Priymenko N, Bertrand-Michel J, Pineau T, Guillou H, Martin PG, Mselli-Lakhal L. Low doses of bisphenol A induce gene expression related to lipid synthesis and trigger triglyceride accumulation in adult mouse liver. Hepatology. 2012;55(2):395-407. doi: 10.1002/hep.24685.

Acknowledgements

Special thanks to Çanakkale Onsekiz Mart University Experimental Research Application, and Research Center Chief Veterinarian Dr. Sait Elmas.

Correspondence:

Tuncer Şimşek Department of Anesthesiology and Reanimation Canakkale Onsekiz Mart University, Medicine Faculty

Canakkale Turkey

Phone: +90 543 408 89 13 rtksege@hotmail.com

Received: July 03, 2018 Review: Sept 06, 2018 Accepted: Oct 09, 2018 Conflict of interest: none

Financial source: Çanakkale Onsekiz Mart University Scientific Research Projects Coordination Unit

¹Research performed at Canakkale Onsekiz Mart University Experimental Research Application and Research Center, Turkey.

